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Synthesis of 4-Oxo- and 4-*anti*-Formyl-8,10,12,13-tetraoxapentacyclo-[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecanes

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Abstract: The synthesis of 4-oxo- and 4-*anti*-formyl-8,10,12,13tetraoxapentacyclo $[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]$ tridecanes has been accomplished. Ozonolysis of compounds 10a,b and 12a-c in dichloromethane at -78 °C followed by reduction with dimethyl sulfide gave the title compounds, 4oxo-tetraoxa-cages 11a,b and 14a-c, in moderate yields. Ozonolysis of the *endo-syn* isomers 15a,b and 18a,b under the same reaction conditions gave 4-*anti*-formyl-tetraoxa-cages 17a,b and 20a,b, respectively. © 1997 Published by Elsevier Science Ltd.

Introduction

The synthesis and chemistry of polycyclic cage compounds have attracted considerable attention in recent years.¹ The vast majority of the work reported in this area has dealt with carbocyclic cage compounds, such as triprismane,² tetraprismane (cubane),³ pentaprismane,⁴ homopentaprismane,⁵ hexaprismane,⁶ dodecahedrane,⁷ heptacyclotetradecane (HCTD),⁸ pogodane,⁹ and fullerenes.¹⁰ On the other hand, the synthesis and chemistry of heterocyclic cage compounds have received less attention. However, there are some reports regarding the chemistry¹¹ and synthesis¹²⁻¹⁷ of oxa-cage compounds in the literature. This class of heterocyclic cage compounds is synthesized by intramolecular alkene-oxirane (2σ - 2π) photocycloaddition,¹² by transannular cyclization of suitable compounds,¹³ by tandem cyclization,¹⁴ by dehydration of diols having the proper stereochemistry,¹⁵ by base-promoted rearrangement,¹⁶ and by intramolecular etherification of an alkene bond with organoselenium reagents.¹⁷

Recently, we developed new methods for the synthesis of a series of oxa-cage compounds, such as diacetal trioxa-cages,¹⁸ triacetal trioxa-cages,¹⁹ tetraacetal tetraoxa-cages,²⁰ tetraacetal pentaoxa-cages,²¹ and pentaacetal pentaoxa-cages (the pentaoxa[5]peristylanes).²² We also investigated the chemical nature of the acetal groups of tetraoxa-cages and discovered a novel hydride rearrangement and one-pot conversion from oxa-cages to aza-cages.²³ As part of a program that involves the synthesis, chemistry, and applications of new heterocyclic cages, we report here the synthesis of 4-oxo- and 4-*anti*-formyl-8,10,12,13-tetraoxapentacyclo-[$5.5.1.0^{2,6}.0^{3,11}.0^{5,9}$]tridecanes. 4-Oxo-8,10,12,13-tetraoxapentacyclo[$5.5.1.0^{2,6}.0^{3,11}.0^{5,9}$]tridecane is a new system for the facial selectivity study on carbonyl group. We have investigated the facial selectivity of a series of oxa-cages, and the results will be reported soon.

Results and Discussion

Reaction of 6,6-dimethylfulvene 1 (commercially available) with (Z)-3-hexene-2,5-dione 2a^{20a} in dichloromethane at 25 \degree for 24 h gave the endo adduct 3 (10%) and compound 4 (50%) (Scheme 1). When the reaction time was prolonged at 25 °C for 72 h, compound 4 was obtained in 70% yield. Refluxing the adduct 3 in dichloromethane for 24 h gave 4 in 80% yield. No detectable amount of the other regioisomer 5 was obtained. The regiochemistry of 4 was determined by ${}^{1}H^{-1}H$ correlated two-dimensional NMR spectral analysis. Proton H₁ (δ 6.53) showed strong coupling to proton H₂ (δ 5.74), which, in turn, displayed coupling to proton H₃ (δ 5.30). Proton H₃ exhibited strong coupling to proton H₄ (δ 3.69), which, in turn, showed coupling to H_5 (δ 3.03), and H_5 displayed coupling to H_6 (δ 4.87). The stereochemistry of 4 was determined on the basis of NOE experiments. Irradiating the H_4 proton gives 4.8% enhancement for the H_3 proton absorptions and 4.2% enhancement for the H_5 proton absorptions. Reaction of 1 with (Z)- γ -oxo- α , β -unsaturated thioesters **6a**- c^{20g} in CH₂Cl₂ at 25 °C for 24 h gave compounds 8a-c in 60-65% yields. The amount of the endo adducts 7a-c was too small to be isolated. We proposed that compounds 4 and 8 were obtained by a [3,3] sigmatropic rearrangement from 3 and 7, respectively.

To improve the yields of the *endo* adducts, 6-acetoxyfulvene 9 was prepared for the Diels-Alder reaction as a diene. Reaction of 9 with the ene-diones 2a,b in dichloromethane at 25 °C for 48 h gave the *endo* adducts 10a,b in 50-55% yields, with unreacted starting compounds. Ozonolysis of 10a,b in dichloromethane at -78 °C followed by reduction with dimethyl sulfide gave 4-oxo-tetraoxa-cage compounds 11a,b in 60-66% yields (Scheme 2). Ozonolysis of 3 under the same reaction conditions gave 11a in 65% yield. Thus, we have accomplished the synthesis of new tetraoxa-cages with a carbonyl group on the apex carbon, 4-oxo-8,10,12,13tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecanes, in a short sequence.

Scheme 1



Scheme 2



Reaction of 9 with 6a-c in dichloromethane at 25 $^{\circ}$ C for 48 h gave the *endo* adducts 12a-c (30%) and compounds 13a-c (35%) (Scheme 3). Both compounds 12 and 13 contained two regioisomers in each case from their ¹H and ¹³C NMR spectra. Ozonolysis of 12a-c in dichloromethane at -78 $^{\circ}$ C followed by reduction with dimethyl sulfide gave 4-oxo-tetraoxa-cage compounds 14a-c in 60-65% yields.

Scheme 3



Hydrolysis of the *endo* adducts 10a,b with a catalytic amount of sodium carbonate in aqueous methanol (1:1) at 25 $^{\circ}$ C gave the *endo-syn* isomers 15a,b in 80-90% yields (Scheme 4). The amount of the *endo-anti* isomers 16a,b was too small to be isolated. The stereochemistry of the formyl group on the apical carbon of 15 was determined by the following chemical transformation. Ozonolysis of 15a,b in dichloromethane at -78 $^{\circ}$ C followed by reduction with dimethyl sulfide gave the tetraaectal tetraoxa-cages 17a,b in 75-85% yields. Scheme 4



Hydrolysis of the mixtures of the two regionsomers of 12a,b with a catalytic amount of sodium carbonate in aqueous methanol (1:1) at 25 °C gave the *endo-syn* isomers 18a,b and the *endo-anti* isomers 19a,b in ratios of 7-8:1 in 80-85% yields (Scheme 5). The stereochemistry of the formyl group on the apical carbon of 18 and 19 was determined by the following chemical transformation. Ozonolysis of the *endo-syn* isomers 18a,b in dichloromethane at -78 °C followed by reduction with dimethyl sulfide gave the methylthio group substituted tetraacetal tetraoxa-cages 20a,b in 80-85% yields. As expected, the thioester group of 18a,b participated the cyclization process.^{20d}

Scheme 5



Conclusion

We have accomplished the synthesis of 4-oxo- and 4-anti-formyl-8,10,12,13tetraoxapentacyclo $[5.5.1.0^{2.6}.0^{3.11}.0^{5.9}]$ tridecanes in a short sequence. Ozonolysis of compounds 10a,b and 12a-c in dichloromethane at -78 °C followed by reduction with dimethyl sulfide gave the title compounds, 4-oxo-tetraoxa-cages 11a,b and 14a-c, in moderate yields. Ozonolysis of the *endo*-syn isomers 15a,b and 18a,b under the same reaction conditions gave 4-anti-formyltetraoxa-cages 17a,b and 20a,b, respectively. The formation of tetraoxa-cages can aslo be used as a probe for determining the stereochemistry of the formyl group on the apical carbon.

Experimental Section

General. Melting points were determined in capillary tubes with a Laboratory Devices melting point apparatus and uncorrected. Infrared spectra were recorded in $CHCl_3$ solutions or on neat thin films between NaCl disks. ¹H NMR spectra were determined at 300 MHz, and ¹³C NMR were determined at 75 MHz on Fourier transform spectrometers. Chemical shifts are reported in ppm relative to TMS in the solvents specified. The multiplicities of ¹³C signals were determined by DEPT techniques. High resolution mass values were obtained with a high resolution mass spectrometer at the Department of Chemistry, National Tsing Hua University. Elemental analyses were performed at the microanalysis laboratory of National Taiwan University. For thin-layer chromatography (TLC) analysis, precoated TLC plates (Kieselgel 60 F_{254}) were used, and column chromatography was done by using Kieselgel 60 (70-230 mesh) as the stationary phase. THF was distilled immediately prior to use from sodium benzophenone ketyl under nitrogen.

Reaction of 6,6-Dimethylfulvene with (Z)-3-Hexene-2,5-dione 2a.

To a solution of (Z)-3-hexene-2,5-dione $2a^{20a}$ (1.1 g, 9.5 mmol) in dichloromethane (20 mL) was added 6,6-dimethylfulvene (1.0 g, 9.4 mmol) at 25 °C. The reaction mixture was stirred at 25 °C for 24 h. The solvent was evaporated and the crude product was purified by column chromatography to give the *endo* adduct **3** (0.21 g, 10%) and compound **4** (1.02 g, 50%). Spectral data for **3**: pale yellow oil; IR (neat) 2980, 1710, 1380 cm⁻¹,¹H NMR (300 MHz, CDCl₃) δ 6.33 (brs, 2H), 3.62 (brs, 2H), 3.35 (brs, 2H), 2.10 (s, 6H), 1.57 (s, 6H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 206.43 (2CO), 144.34 (C), 137.89 (C), 134.55 (2CH), 57.08 (2CH), 46.41 (2CH), 29.97 (2CH₃), 19.44 (2CH₃); LRMS m/z (rel inten) 218 (M⁺, 36), 203 (40), 112 (100); HRMS (EI) calcd for C₁₄H₁₈O₂ 218.1307, found 218.1301.

Spectral data for 4: pale yellow oil; IR (neat) 2980, 1705, 1360 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.53 (d, J = 6.0 Hz, 1H), 5.74 (dd, J = 6.0, 1.5 Hz, 1H), 5.30 (dd, J = 8.1, 1.5 Hz, 1H), 4.87 (d, J = 4.5 Hz, 1H), 3.69 (dd, J = 6.6, 4.5 Hz, 1H), 3.03 (dd, J = 8.1, 6.6 Hz, 1H), 2.16 (s, 3H), 1.74 (s, 3H), 1.70 (s, 3H), 1.65 (s, 3H), ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 206.57 (CO), 151.91 (C), 140.43 (C), 133.67 (CH), 132.36 (CH), 123.01 (C), 96.64 (CH) 81.55 (CH), 49.26 (CH), 40.06 (CH), 27.18 (CH₃), 20.56 (CH₃), 20.24 (CH₃), 20.13 (CH₃); LRMS m/z (rel inten) 218 (M⁺, 21), 203 (100); HRMS (EI) calcd for C₁₄H₁₈O₂ 218.1307, found 218.1312.

Refluxing the endo Adduct 2a in Dichloromethane. To a solution of 2a (1.0 g, 4.6 mmol)

in dichloromethane (50 mL) was refluxed at 45 °C for 48 h. After cooling, the solvent was evaporated and crude product was purified by column chromatography to give 3a (0.80 g, 80%).

General Procedure for Reaction of 6,6-Dimethylfulvene with (Z)- γ -Oxo- α , β unsaturated Thioesters 6a-c. To a solution of (Z)-methyl- γ -oxo-2-pententhioate 6a (1.36 g, 9.4 mmol) in dichloromethane (20 mL) was added 6,6-dimethylfulvene (1.0 g, 9.4 mmol) at 25 $^\circ$ C. The reaction mixture was stirred at 25 °C for 48 h. The solvent was evaporated and the crude product was purified by column chromatography to give 8a in 65% yield with unreacted starting compounds. Spectral data for 8a: pale yellow oil; IR (neat) 2960, 2880, 1685, 1380 cm⁻¹, ¹H NMR (300 MHz, CDCl₃) δ 6.52 (d, J = 5.7 Hz, 1H), 5.78 (dd, J = 5.7, 1.5 Hz, 1H), 5.29 (dd, J = 5.7 7.2, 1.5 Hz, 1H), 4.83 (d, J = 4.5 Hz, 1H), 3.66 (dd, J = 6.9, 4.5 Hz, 1H), 3.16 (dd, J = 7.2, 6.9 Hz, 1H), 2.28 (s, 3H), 1.76 (s, 3H), 1.75 (s, 3H), 1.70 (s, 3H); 13 C NMR (75 MHz, CDCl₃, DEPT) δ 202.26 (COS), 153.34 (C), 139.96 (C), 133.76 (CH), 133.06 (CH), 124.81 (C), 96.78 (CH), 81.66 (CH), 49.87 (CH), 42.36 (CH), 21.20 (CH₃), 20.88 (CH₃), 20.59 (CH₃), 12.08 (SCH₃); LRMS m/z (rel inten) 250 (M⁺, 18), 203 (100); HRMS (EI) calcd for C₁₄H₁₈O₂S 250.1028, found 250.1024. Spectral data for 8b: pale yellow oil; yield 63% IR (neat) 2960, 1685, 1380 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.53 (d, J = 5.7 Hz, 1H), 5.72 (dd, J = 5.7, 1.5 Hz, 1H), 5.32 (dd, J = 7.2, 1.5 Hz, 1H), 4.90 (d, J = 4.8 Hz, 1H), 3.77 (dd, J = 6.9, 4.8 Hz, 1H), 3.14 (dd, J = 7.2, 6.9 Hz, 1H), 2.26 (s, 3H), 2.24-2.17 (m, 1H), 1.75 (s, 3H), 1.74 (s, 3H), 1.04 (d, J = 6.6 Hz, 3H), 1.03 (d, J = 6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 202.64 (COS), 161.76 (C), 140.29 (C), 134.14 (CH), 132.83 (CH), 124.43 (C), 95.04 (CH), 81.84 (CH), 49.58 (CH), 43.35 (CH), 32.57 (CH), 21.03 (CH₃), 20.85 (CH₃), 19.63 (CH₃), 19.34 (CH₃), 12.11 (SCH₃); LRMS m/z (rel inten) 278 (M⁺, 8), 231 (100);

Spectral data for 8c: pale yellow oil; IR (neat) 2970, 1685, 1370 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.52 (d, J = 6.0 Hz, 1H), 5.75 (dd, J = 6.0, 1.5 Hz, 1H), 5.29 (dd, J = 7.2, 1.5 Hz, 1H), 4.85 (d, J = 4.5 Hz, 1H), 3.70 (dd, J = 6.9, 4.5 Hz, 1H), 3.14 (dd, J = 7.2, 6.9 Hz, 1H), 2.27 (s, 3H), 2.05-1.98 (m, 2H), 1.75 (s, 3H), 1.72 (s, 3H), 1.48-1.25 (m, 4H), 0.89 (t, J = 7.5 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 202.35 (COS), 156.95 (C), 140.17 (C), 133.90 (CH), 133.06 (CH), 124.58 (C), 96.52 (CH), 81.69 (CH), 49.79 (CH), 42.79 (CH), 34.14 (CH₂), 28.25 (CH₂), 22.22 (CH₂), 21.15 (CH₃), , 20.83 (CH₃), 13.80 (CH₃), 12.08 (SCH₃); LRMS m/z (rel inten) 292 (M⁺, 11), 245 (100); HRMS (EI) calcd for C₁₇H₂₄O₂S 292.1497, found 292.1489.

HRMS (EI) calcd for $C_{16}H_{22}O_2S$ 278.1341, found 278.1338.

General Procedure for Reaction of 6-Acetoxyfulvene with (Z)-2-Ene-1,4-diones 2a,b. To a solution of 2a (0.82 g, 7.4 mmol) in dichloromethane (20 mL) was added 6-acetoxyfulvene (1.0 g, 7.3 mmol) at 25 °C. The reaction mixture was stirred at 25 °C for 48 h. The solvent was evaporated and the crude product was purified by column chromatography to give the endo adduct 10a (0.98 g, 55%). In the case of 10b, the product contained two regioisomers. Spectral data for 10a: pale yellow oil; IR (neat) 2980, 2880, 1755, 1710, 1225 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.60 (s, 1H), 6.32 -6.29 (m, 2H), 3.87 (brs, 1H), 3.50-3.42 (m, 3H), 2.13 (s, 3H) 2.11 (s, 3H) 2.10 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 205.41 (CO), 205.35 (CO), 168.17 (CO), 136.76 (C), 134.49 (CH), 133.50 (CH), 116.45 (CH), 57.33 (CH), 56.14 (CH), 46.70 (CH), 44.75 (CH), 29.97 (CH₃), 29.89 (CH₃), 20.59 (CH₃); LRMS m/z (rel inten) 248 (M⁺, 26), 233 (100); HRMS (EI) calcd for C₁₄H₁₆O₄ 248.1049, found 248.1044.

Spectral data for 10b: pale yellow oil; yield 55% IR (neat) 2980, 1755, 1720, 1710, 1225 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.55 and 9.54 (d, J = 3.0 Hz, 1H), 6.66 and 6.62 (s, 1H), 6.56-6.48 (m, 1H), 6.23-6.16 (m, 1H), 4.02 and 3.90 (brs, 1H), 3.77-3.71 (m, 1H), 3.64 and 3.50 (brs, 1H), 3.10-3.03 (m, 1H), 2.23 and 2.20 (s, 3H), 2.16 and 2.13 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 205.43 (CO), 205.39 (CO), 200.64 (CHO), 200.60 (CHO), 168.06 (2CO), 136.98 (CH), 136.73 (C), 136.51 (C), 135.80 (CH), 133.51 (CH), 132.41 (CH), 116.82 (CH), 116.80 (CH), 59.85 (CH), 58.76 (CH), 55.93 (CH), 54.80 (CH), 47.12 (CH), 45.41 (CH), 45.14 (CH), 43.59 (CH), 28.88 (CH₃), 28.83 (CH₃), 20.66 (CH₃), 20.62 (CH₃); LRMS m/z (rel inten) 234 (M⁺, 12), 219 (100); HRMS (EI) calcd for C₁₃H₁₄O₄ 234.0892, found 234.0896.

Synthesis of Tetraoxa-Cages 11a,b from Ozonolysis of 10a,b. A solution of 10a (1.0 g, 4.0 mmol) in dichloromethane (30 mL) was cooled to -78 °C, and ozone was bubbled through it at -78 °C until the solution turned light blue. To this solution was added dimethyl sulfide (0.56 g, 9.0 mmol) at -78 °C. Then, the reaction mixture was stirred at room temperature for 5 h. The solvent was evaporated, and the crude product was purified by column chromatography to give the tetraacetal oxa-cage compound 11a (0.59 g, 66%).

1,7-Dimethyl-4-oxo-8,10,12,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecanes 11a: white waxy solid; mp 122-123 °C; IR (CHCl₃) 2880, 1765, 1380, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 5.87 (d, J = 5.7 Hz, 2H), 3.28-3.16 (m, 4H), 1.64 (s, 6H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 204.97 (CO), 121.00 (2C) , 107.27 (2CH), 55.09 (2CH), 50.40 (2CH), 24.90 (2CH₃); LRMS m/z (rel inten) 224 (M⁺, 23), 196 (100); HRMS (EI) calcd for C₁₁H₁₂O₅ 224.0685, found 224.0689; Anal, calcd for C₁₁H₁₂O₅: C, 58.91; H,5.40, found: C, 58.82; H, 5.45.

1-Methyl-4-oxo-4,8,10,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecane 11b: white waxy solid; mp 94-95 °C; yield 60%; IR (CHCl₃) 2980, 2880, 1765, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ 6.15 (d, J = 5.1 Hz, 1H), 5.87 (d, J = 6.6 Hz, 2H), 3.62-3.56 (m, 1H), 3.22-3.10 (m, 3H), 1.65 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 205.00 (CO), 121.35 (C), 112.52 (CH), 107.54 (CH), 107.39 (CH), 55.06 (CH), 54.59 (CH), 49.20 (CH), 47.08 (CH), 24.53 (CH₃); LRMS *m/z* (rel inten) 210 (M⁺, 37), 182 (100); HRMS (EI) calcd for C₁₀H₁₀O₅ 210.0528, found 210.0531; Anal. calcd for C₁₀H₁₀O₅: C, 57.13; H, 4.80, found: C, 57.01; H, 4.88.

Synthesis of Tetraacetal Tetraoxa-Cage 11a from Ozonolysis of 3. The same reaction conditions and procedure as for the ozonolysis of 10a,b were applied for the ozonolysis of 3 to give the tetraoxa-cage 11a in 65% yield.

General Procedure for the Reactions of 6-Acetoxyfulvene with (Z)- γ -Oxo- α , β unsaturated Thioesters 6a-c. To a solution of 6a (1.1 g, 7.4 mmol) in dichloromethane (30 mL) was added 6-acetoxyfulvene (1.0 g, 7.3 mmol) at 25 °C. The reaction mixture was stirred at 25 $^{\circ}$ C for 48 h. The solvent was evaporated, and the crude product was purified by column chromatography to give the endo adduct 12a (0.60 g, 30%) and compound 13a (0.70 g, 35%). In each case, both the endo adducts 12a-c and compounds 13a-c contained two regioisomers, and their spectral data were taken as mixtures of two regioisomers. Spectral data for 12a: pale yellow oil; IR (neat) 2980, 1755, 1710, 1690, 1220 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.62 and 6.59 (s, 1H), 6.58-6.53 (m, 1H), 6.16-6.09 (m, 1H), 3.93 and 3.83 (brs, 1H), 3.81-3.76 (m, 1H), 3.57 and 3.42 (brs, 1H), 3.31-3.25 (m, 1H), 2.30 and 2.29 (s, 3H), 2.15 and 2.12 (s, 3H), 2.05 and 2.04 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) & 205.27 (2CO), 197.24 (2COS), 168.14 (CO), 168.16 (CO), 137.10 (CH), 136.48 (C), 136.13 (C), 135.99 (CH), 132.52 (CH), 131.47 (CH), 116.71 (CH), 116.69 (CH), 58.05 (CH), 57.11 (CH), 56.85 (CH), 56.09 (CH), 48.59 (CH), 46.56 (CH), 46.01 (CH), 44.16 (CH), 30.65 (CH₃), 30.62 (CH₃), 20.65 (CH₃), 20.60 (CH₃), 11.76 (SCH₃), 11.73 (SCH₃); LRMS m/z (rel inten) 280 (M⁺, 21), 233 (100); HRMS (EI) calcd for C₁₄H₁₆O₄S 280.0769, found 208.0761.

Spectral data for 13a: pale yellow oil; IR (neat) 2980, 1760, 1685, 1215, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.29 and 7.03 (s, 1H), 6.68 and 6.29 (d, J = 5.7 Hz, 1H), 6.08 and 5.95 (dd, J = 5.7, 2.1 Hz, 1H), 5.20 and 5.11 (dd, J = 8.1, 2.1 Hz, 1H), 4.84 and 4.72 (d, J = 5.1 Hz, 1H), 3.84-3.80 (m, 1H), 3.53-3.46 (m, 1H), 2.32 and 2.31 (s, 3H), 2.16 and 2.15 (s, 3H), 1.75 and 1.74 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 201.21 (COS), 201.14 (COS), 167.40 (CO), 167.27 (CO), 153.63 (2C), 135.36 (CH), 134.88 (CH), 133.17 (CH), 132.06 (C), 130.76 (CH), 130.35 (C), 129.00 (CH), 127.21 (CH), 96.30 (CH), 93.94 (CH), 80.80 (CH), 79.34 (CH), 48.43 (CH), 47.46 (CH), 40.41 (CH), 39.52 (CH), 20.58 (2CH₃), 20.47 (2CH₃), 11.85 (2SCH₃); LRMS m/z (rel inten) 280 (M⁺, 8), 233 (100); HRMS (EI) calcd for C₁₄H₁₆O₄S 280.0769, found 280.0756.

Spectral data for 12b: pale yellow oil; IR (neat) 2980, 1755, 1710, 1690, 1220 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.63 and 6.59 (s, 1H), 6.60-6.54 (m, 1H), 6.15-6.09 (m, 1H), 3.93 and 3.80 (brs,

1H), 3.79-3.72 (m, 1H), 3.55 and 3.37 (brs, 1H) 3.52-3.45 (m, 1H), 2.48-2.38 (m, 1H), 2.28 and 2.27 (s, 3H), 2.15 and 2.12 (s, 3H), 1.09 and 1.07 (d, J = 6.6 Hz, 3H), 1.04 and 1.02 (d, J = 6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 211.87 (2CO), 197.18 (2COS), 168.22 (2CO), 137.41 (CH), 136.61 (C), 136.39 (C), 136.35 (CH), 132.09 (CH), 131.03 (CH), 116.58 (CH), 116.52 (CH), 57.39 (CH), 56.45 (CH), 54.63 (CH), 53.51 (CH), 48.46 (CH), 46.38 (CH), 46.26 (CH), 44.39 (CH), 41.40 (2CH), 20.66 (2CH₃), 19.80 (CH₃), 19.71 (CH₃), 17.28 (2CH₃), 11.73 (SCH₃), 11.70 (SCH₃); LRMS m/z (rel inten) 308 (M⁺, 12), 261 (100); HRMS (EI) calcd for C₁₆H₂₀O₄S 308.1082, found 308.1078.

Spectral data for 13b: pale yellow oil; yield 33 %; IR (neat) 2980, 1760, 1685, 1215, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.28 and 7.02 (s, 1H), 6.66 and 6.28 (d, J = 6.0 Hz, 1H), 6.02 and 5.91 (dd, J = 6.0, 2.1 Hz, 1H), 5.23 and 5.14 (dd, J = 7.2, 2.1 Hz, 1H), 4.91 and 4.81 (d, J = 4.8 Hz, 1H), 3.95-3.91 and 3.74-3.69 (m, 1H), 3.63-3.60 and 3.53-3.50 (m, 1H), 2.34 and 2.31 (s, 3H), 2.30-2.20 (m, 1H), 2.17 and 2.16 (s, 3H), 1.02 and 1.00 (d, J = 6.6 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 201.60 (2COS), 167.45 (CO), 167.37 (CO), 162.32 (2C), 135.36 (CH), 134.92 (CH), 133.37 (CH), 132.31 (C), 130.89 (CH), 130.80 (C), 128.80 (CH), 127.14 (CH), 94.95 (CH), 93.23 (CH), 81.32 (CH), 80.19 (CH), 48.30 (CH), 47.26 (CH), 41.51 (CH), 40.56 (CH), 32.55 (CH), 32.48 (CH), 19.73 (CH₃), 19.69 (CH₃), 19.53 (CH₃), 19.41 (CH₃), 11.96 (CH₃), 11.94 (CH₃), 11.69 (CH₃), 11.44 (CH₃); LRMS m/z (rel inten) 308 (M⁺, 14), 261 (100); HRMS (EI) calcd for C₁₆H₂₀O₄S 308.1082, found 308.1088.

Spectral data for 12c: pale yellow oil; yield 31%; IR (neat) 2980, 1755, 1710, 1690, 1220 cm⁻¹;¹H NMR (300 MHz, CDCl₃) δ 6.61 and 6.58 (s, 1H), 6.57-6.50 (m, 1H), 6.17-6.10 (m, 1H), 3.92 and 3.79 (brs, 1H), 3.78-3.71 (m, 1H), 3.53 and 3.40 (brs, 1H) 3.35-3.25 (m, 1H), 2.28 and 2.26 (s, 3H), 2.28-2.20 (m, 2H), 2.15 and 2.12 (s, 3H), 1.60-1.20 (m, 4H), 0.88 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 207.74 (2CO), 197.25 (2COS), 168.22 (2CO), 137.04 (CH), 136.61 (C), 136.33 (C), 135.96 (CH), 132.53 (CH), 131.49 (CH), 116.62 (CH), 116.59 (CH), 57.21 (CH), 57.07 (CH), 56.21 (CH), 55.90 (CH), 48.46 (CH), 46.40 (CH), 46.17 (CH), 44.30 (CH), 43.27 (2CH₂), 25.85 (2CH₂), 22.27 (2CH₂), 20.68 (CH₃), 20.64 (CH₃), 13.82 (2CH₃), 11.79 (SCH₃), 11.76 (SCH₃); LRMS m/z (rel inten) 322 (M⁺, 16), 275 (100); HRMS (EI) calcd for C₁₇H₂₂O₄S 322.1239, found 322.1235.

Spectral data for 13c: pale yellow oil; yield 31%; IR (neat) 2980, 1760, 1685, 1215, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.28 and 7.03 (s, 1H), 6.68 and 6.29 (d, J = 5.7 Hz, 1H), 6.05 and 5.93 (dd, J = 5.7, 2.1 Hz, 1H), 5.22 and 5.12 (dd, J = 7.2, 2.1 Hz, 1H), 4.87 and 4.76 (d, J = 3.6 Hz, 1H), 3.90-3.86 and 3.70-3.65 (m, 1H), 3.55-3.45 (m, 1H), 2.33 and 2.31 (s, 3H), 2.18 and 2.16 (s, 3H), 2.05 -1.98 (m, 2H), 1.46-1.23 (m, 4H), 0.90 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃)

DEPT) δ 201.46 (COS), 201.40 (COS), 167.53 (CO), 167.43 (CO), 157.47 (2C), 135.47 (CH), 135.01 (CH), 133.32 (CH), 132.30 (C), 130.83 (CH), 130.70 (C), 128.96 (CH), 127.25 (CH), 96.34 (CH), 94.21 (CH), 81.07 (CH), 79.92 (CH), 48.80 (CH), 47.46 (CH), 41.03 (CH), 40.10 (CH), 34.09 (CH₂), 34.05 (CH₂), 28.45 (CH₂), 28.30 (CH₂), 22.19 (2CH₂), 20.68 (2CH₃), 13.84 (2CH₃), 11.96 (2SCH₃); LRMS m/z (rel inten) 322 (M⁺, 13), 275 (100); HRMS (EI) calcd for C₁₇H₂₂O₄S 322.1239, found 322.1230.

Synthesis of Tetraoxa-Cages 14a-c from Ozonolysis of 12a-c. The same reaction conditions and procedure as for the synthesis of tetraoxa-cages 11a,b from ozonolysis of 10a,b were applied for the synthesis of tetraoxa-cages 14a-c from ozonolysis of 12a-c.

1-Methylthio-7-methyl-4-oxo-8,10,12,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecane 14a: white waxy solid; mp 88-89 °C; yield 62%; IR (CHCl₃) 2880, 1765, 1380, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ 5.94 (d, J = 6.0 Hz, 1H), 5.89 (d, J = 6.0 Hz, 1H), 3.62 (dd, J = 9.9, 6.0 Hz, 1H), 3.32 (dd, J = 9.9, 6.0 Hz, 1H), 3.28-3.16 (m, 2H), 2.26 (s, 3H), 1.68 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 203.74 (CO), 125.63 (C), 122.06 (C), 107.98 (CH), 107.42 (CH), 55.01 (CH), 54.61 (CH), 52.79 (CH), 50.02 (CH), 24.58 (CH₃), 12.94 (SCH₃); LRMS m/z (rel inten) 256 (M⁺, 32), 209 (100); HRMS (EI) calcd for C1₁₁H₁₂O₅S 256.0405, Found 256.0411; Anal. calcd for C1₁₁H₁₂O₅S: C, 51.55; H 4.72, found: C,51.47; H, 4.77.

1-Methylthio-7-isopropyl-4-oxo-8,10,12,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]-

tridecane 14b: white waxy solid; mp 65-66 °C; yield 60%; IR (CHCl₃) 2880, 1765, 1380, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ 5.94 (d, J = 6.6 Hz, 1H), 5.91 (d, J = 6.6 Hz, 1H), 3.56 (dd, J = 10.2, 6.6Hz, 1H), 3.32 (dd, J = 10.2, 6.6 Hz, 1H), 3.26 (dd, J = 7.2, 6.6 Hz, 1H), 3.08 (dd, J = 7.2, 6.6 Hz, 1H), 2.26 (s, 3H), 2.22-2.14 (m, 1H), 1.03 (d, J = 6.6 Hz, 3H), 1.01 (d, J = 6.6 Hz, 3H); ¹³C NMR (75MHz, CDCl₃, DEPT) δ 203.96 (CO), 126.64 (C), 125.75 (C), 107.95 (CH), 107.44 (CH), 55.29 (CH), 54.66 (CH), 52.30 (CH), 46.49 (CH), 34.77 (CH), 17.14 (CH₃), 17.07 (CH₃), 13.01 (SCH₃); LRMS m/z (rel inten) 284 (M⁺, 19), 237 (100); HRMS (EI) calcd for C₁₃H₁₆O₅S 284.0718, found 284.0723; Anal. calcd for C₁₃H₁₆O₅S: C, 54.92; H, 5.68, found:C, 54.84; H, 5.74.

1-Methylthio-7-butyl-4-oxo-8,10,12,13-Tetraoxapentacyclo[5.5.1.0^{2.6}.0^{3.11}.0^{5.9}]tridecane 14c: white waxy solid; mp 60-61 °C; yield 64%; IR (CHCl₃) 2980, 2880, 1765, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ 5.94 (d, J = 6.0 Hz, 1H), 5.90 (d, J = 6.0 Hz, 1H), 3.58 (dd, J = 10.2, 6.0 Hz, 1H), 3.34-3.25 (m, 2H), 3.17 (dd, J = 7.2, 6.0 Hz, 1H), 2.26 (s, 3H), 1.93-1.86 (m, 2H), 1.70-1.62 (m, 2H), 1.40-1.32 (m, 2H), 0.92 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 203.89 (CO), 125.60 (C), 124.24 (C), 107.97 (CH), 107.38 (CH), 55.16 (CH), 54.61 (CH), 52.45 (CH), 48.33 (CH), 37.09 (CH₂), 26.07 (CH₂), 22.53 (CH₂),13.90 (CH₃), 12.99 (SCH₃); LRMS m/z (rel inten) 298 (M⁺, 14), 251 (100); HRMS (EI) calcd for C₁₄H₁₈O₅S 298.0875, found 298.0870; Anal. calcd for C14H18O5S: C, 56.36; H, 6.09, found: C, 56.29; H, 6.14.

General Procedure for The Hydrolysis of 10a,b with Na₂CO₃ in Aqueous Methanol. To a solution of 10a (1.0 g, 4.0 mmol) in methanol (10 mL) and water (10 mL) was added Na₂CO₃ (0.040 g, 0.40 mmol) at 25 °C. The reaction mixture was stirred at 25 °C for 2 h. The solvent evaporated, and saturated NH₄Cl solution (10 mL) was added. The reaction mixture was extracted with ether (5 x 30 mL). The organic layer was washed with brine, dried over MgSO₄, and evaporated, and the residue was purified by column chromatography to give the *endo-syn* isomer 15a (0.74 g, 90%). The amount of the *endo-anti* isomer 16a was too small to be isolated.

2,3-Bis-endo-diacetyl-7-syn-formylbicyclo[**2.2.1**]-**5-heptene 15a**: pale yellow oil; IR (neat) 2980, 2880, 1720, 1710 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.60 (s, 1H), 6.26 (brs, 2H), 3.46 (brs, 2H), 3.32 (bres, 2H), 2.53 (brs, 1H), 2.10 (s,6H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 205.61 (2CO), 200.10 (CHO), 134.66 (2CH), 68.00 (CH), 54.68 (2CH), 45.74 (2CH), 30.00 (2CH₃); LRMS m/z (rel inten) 206 (M⁺, 17), 128 (100); HRMS (EI) calcd for C₁₂H₁₄O₃ 206.0943, found 206.0948. **2-endo-Acetyl-3-endo-7-syn-diformylbicyclo**[**2.2.1**]-**5-heptene 15b**: pale yellow oil; IR (neat) 2980, 2880, 1720, 1710 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.60 (s, 1H), 9.54 (d, J = 2.1 Hz, 1H), 6.47 (dd, J = 6.0, 3.0 Hz, 1H), 6.13 (dd, J = 6.0, 3.0 Hz, 1H), 3.71 (dd, J = 9.3, 3.6 Hz, 1H) 3.58 (brs, 1H), 3.51 (brs, 1H), 2.96-2.91 (m, 1H), 2.60 (s, 1H), 2.19 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 205.83 (CO), 200.35 (CHO), 199.61 (CHO), 136.87 (CH), 133.55 (CH), 68.60 (CH), 56.78 (CH), 53.51 (CH), 46.07 (CH), 44.55 (CH), 28.88 (CH₃); LRMS m/z (rel inten) 192 (M⁺, 12), 114 (100); HRMS (EI) calcd for C₁₁H₁₂O₃ 192.0786, found 192.0789.

Ozonolysis of 15a,b. Formation of Tetraoxa-Cages 17a,b. The same reaction conditions and procedure as for the ozonolysis of 10a,b were applied for the ozonolysis of 15a,b to give the tetraoxa-cages 17a,b.

1,7-Dimethyl-4-anti-formyl-8,10,12,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{8,11}.0^{6,9}]tridecane 17a: white waxy solid; mp 55-56 °C; yield 85%; IR (CHCl₃) 2970, 1720, 1070 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.73 (s, 1H), 5.63 (d, J = 5.7 Hz, 2H), 3.22-3.18 (m, 5H), 1.54 (s, 6H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 199.52 (CHO), 117.65 (2C) , 102.79 (2CH), 56.20 (2CH), 55.44 (CH), 45.85 (2CH), 24.96 (2CH₃); LRMS m/z (rel inten) 238 (M⁺, 41), 223 (100); HRMS (EI) calcd for C₁₂H₁₄O₅ 238.0841, found 238.0835; Anal. calcd for C₁₂H₁₄O₅; C, 60.48; H, 5.93, found ; C, 60.40; H, 5.97.

1-Methyl-4-anti-formyl-8,10,12,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecane 17b: highly viscous oil; yield 75%; IR (CHCl₃) 2980, 1720, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ

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9.75 (s, 1H), 5.85 (d, J = 5.4 Hz, 1H), 5.63 (d, J = 6.0 Hz, 1H), 5.62 (d, J = 6.0 Hz, 1H), 3.51-3.46 (m, 1H), 3.28-3.04 (m, 4H), 1.55 (s, 3H); ¹³C NMR (75MHz, CDCl₃, DEPT) δ 199.43 (CHO), 118.02 (C), 109.60 (CH), 103.06 (CH), 102.63 (CH), 55.47 (CH), 55.08 (CH), 52.76 (CH), 45.82 (CH), 45.38 (CH), 24.59 (CH₃); LRMS m/z (rel inten) 224 (M⁺, 27), 209 (100); HRMS (EI) calcd for C₁₁H₁₂O₅ 224.0685, found 224.0689; Anal. calcd for C₁₁H₁₂O₅: C, 58.91; H, 5.40, found: C, 58.80; H, 5.47.

General Procedure for The Hydrolysis of 12a,b with Na₂CO₃ in Aqueous Methanol. The same reaction conditions and procedure as for the hydrolysis of 10a,b were applied for the hydrolysis of 12a,b to give the *endo-syn* isomers 18a,b as the major products and the *endo-anti* isomers 19a,b as the minor products.

Spectral data for **18a**: pale yellow oil; yield 75%; IR (neat) 2980, 2880, 1720, 1710, 1690 cm⁻¹;¹H NMR (300 MHz, CDCl₃) δ 9.60 (s, 1H), 6.54 (dd, J = 6.0, 3.0 Hz, 1H), 6.06 (dd, J = 6.0, 3.0 Hz, 1H), 3.75 (dd, J = 9.0, 3.6 Hz, 1H), 3.51 (brs, 1H), 3.41 (brs, 1H), 3.14 (dd, J = 9.0, 3.6 Hz, 1H), 2.52 (brs, 1H), 2.28 (s, 3H) 2.02 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 205.34 (CO), 199.94 (CHO) , 197.59 (COS), 137.30 (CH), 132.64 (CH), 68.14 (CH), 55.56 (CH), 54.60 (CH), 47.61 (CH), 45.22 (CH), 30.78 (CH₃), 11.81 (SCH₃); LRMS m/z (rel inten) 238 (M⁺, 24), 191 (100); HRMS (EI) calcd for C₁₂H₁₄O₃S 238.0664, found 238.0667.

Spectral data for **19a**: pale yellow oil; yield 10%; IR (neat) 2980, 2880, 1720, 1710, 1690 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.59 (d, J = 2.1 Hz, 1H), 6.50 (dd, J = 6.0, 3.0 Hz, 1H), 6.05 (dd, J = 6.0, 3.0 Hz, 1H), 3.83 (dd, J = 9.0, 3.6 Hz, 1H), 3.60 (brs, 1H), 3.49 (brs, 1H), 3.31 (dd, J = 9.0, 3.6 Hz, 1H), 2.38 (brs, 1H), 2.28 (s, 3H) 2.04 (s, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 204.96 (CO), 203.01 (CHO) , 197.06 (COS), 135.48 (CH), 130.80 (CH), 69.10 (CH), 57.60 (CH), 56.57 (CH), 49.85 (CH), 47.41 (CH), 30.66 (CH₃), 11.81 (SCH₃); LRMS m/z (rel inten) 238 (M⁺, 32), 191 (100); HRMS (EI) calcd for C₁₂H₁₄O₃S 238.0664, found 238.0670.

Spectral data for 18b: pale yellow oil; yield 70%; IR (neat) 2980, 2880, 1720, 1710, 1690, 1380 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.62 (s, 1H), 6.54 (dd, J = 5.7, 3.0 Hz, 1H), 6.06 (dd, J = 5.7, 3.0 Hz, 1H), 3.72 (dd, J = 9.6, 3.6 Hz, 1H), 3.51 (brs, 1H), 3.36-3.32 (m, 2H), 2.53 (brs, 1H), 2.43-2.36 (m, 1H), 2.26 (s, 3H), 1.06 (d, J = 6.0 Hz, 3H), 1.03 (d, J = 6.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 211.95 (CO), 200.04 (CHO), 197.53 (COS), 137.53 (CH), 132.34 (CH), 68.28 (CH), 54.93 (CH), 52.13 (CH), 47.38 (CH), 45.44 (CH), 41.48 (CH), 19.86 (CH₃), 17.32 (CH₃), 11.75 (SCH₃); LRMS m/z (rel inten) 266 (M⁺, 27), 219 (100); HRMS (EI) calcd for C₁₄H₁₆O₃S 266.0977, found 266.0974.

Spectral data for 19b: pale yellow oil; yield 10%; IR (neat) 2980, 2880, 1720, 1710, 1690 cm⁻¹; ¹H

NMR (300 MHz, CDCl₃) δ 9.60 (d, J = 2.1 Hz, 1H), 6.50 (dd, J = 6.0, 3.0 Hz, 1H), 6.06 (dd, J = 6.0, 3.0 Hz, 1H), 3.80 (dd, J=10.2, 3.6 Hz, 1H), 3.61 (brs, 1H), 3.54 (dd, J = 10.2, 3.6 Hz, 1H), 3.44 (brs, 1H), 2.46-2.38 (m, 2H), 2.27 (s, 3H) 1.09 (d, J = 6.6 Hz, 3H), 1.06 (d, J = 6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 211.49 (CO), 203.12 (CHO) , 196.99 (COS), 135.66 (CH), 130.46 (CH), 69.18 (CH), 56.91 (CH), 54.11 (CH), 49.61 (CH), 47.65 (CH), 41.39 (CH), 19.75 (CH₃), 17.27 (CH₃), 11.72 (SCH₃); LRMS m/z (rel inten) 266 (M⁺, 21), 219 (100); HRMS (EI) calcd for C₁₄H₁₈O₃S 266.0977, found 266.0985.

General Procedure for the Ozonolysis of 18a,b. Formation of Tetraoxa-Cages 20a,b. The same reaction conditions and procedure as for the ozonolysis of 10a,b were applied for the ozonolysis of 18a,b to give the tetraoxa-cages 20a,b..

1-Methylthio-7-Methyl-4-anti-formyl-8,10,12,13-tetraoxapentacyclo[5.5.1.0^{2,6}.0^{3,11}.0^{5,9}]tridecane 20a: highly viscous oil; yield 85%; IR (CHCl₃) 2970, 1720, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ 9.74 (s, 1H), 5.71 (d, J = 6.0 Hz, 1H), 5.65 (d, J = 6.0 Hz, 1H), 3.58 (dd, J = 10.8, 5.4 Hz, 1H), 3.34-3.18 (m, 4H), 2.20 (s, 3H), 1.57 (s, 3H); ¹³C NMR (75MHz, CDCl₃, DEPT) δ 199.22 (CHO), 122.74 (C), 118.71 (C), 103.68 (CH), 102.95 (CH), 58.49 (CH), 55.63 (CH), 55.06 (CH), 45.82 (CH), 45.47 (CH), 24.53 (CH₃), 12.87 (SCH₃); LRMS m/z (rel inten) 270 (M⁺, 48), 223 (100); HRMS (EI) calcd for C₁₂H₁₄O₅S 270.0562, found 270.0567; Anal. calcd for C₁₂H₁₄O₅S: C, 53.32; H, 5.22, found: C,53.24; H, 5.28.

1-Methylthio-7-isopropyl-4-anti-formyl-8,10,12,13-tetraoxapentacyclo-

[5.5.1.0^{2.6}.0^{3.11}.0^{5.9}]tridecane 20b: highly viscous oil; yield 80%; IR (CHCl₃) 2980, 1720, 1380, 1070 cm⁻¹; ¹H NMR (300 Hz, CDCl₃) δ 9.75 (s, 1H), 5.70 (d, J = 5.7 Hz, 1H), 5.66 (d, J = 5.7 Hz, 1H), 3.50 (dd, J = 10.5, 5.4 Hz, 1H), 3.30-3.20 (m, 3H), 3.13-3.08 (m, 1H), 2.21 (s, 3H), 2.08-2.01 (m, 1H), 0.99 (d, J = 6.0 Hz, 3H), 0.96 (d, J = 6.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃, DEPT) δ 199.33 (CHO), 123.24 (C), 122.82 (C), 103.62 (CH), 102.87 (CH), 57.99 (CH), 55.13 (CH), 51.89 (CH), 46.01 (CH), 45.55 (CH), 34.52 (CH), 17.10 (CH₃), 17.00 (CH₃), 12.94 (SCH₃); LRMS m/z (rel inten) 298 (M⁺, 24), 251 (100); HRMS (EI) calcd for C₁₄H₁₈O₅S 298.0875, found 298.0869; Anal. calcd for C₁₄H₁₈O₅S: C, 56.36; H, 6.09, found: C, 56.24; H, 6.16.

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