

Nonvolatile Flash Memory Based on Biologically Integrated Hierarchical Nanostructures

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S Supporting Information

[ABSTRACT:](#page-5-0) The first six peptides of multifunctional titanium binding peptide-1 bestowed recombinant L-ferritin, minT1-LF, was genetically engineered and used to fabricate multilayered nanoparticle architecture. The multifunctionality of minT1-LF enables specific binding of nanoparticleaccommodated minT1-LF to the silicon substrate surface and wet biochemical fabrication of gate oxide layer by its biomineralization activity. Three-dimensional (3D) nanoparticle architecture with multilayered structure was fabricated by the biological layer-by-layer method and embedded in a

metal oxide−semiconductor device structure as a charge storage node of a fl[ash memory device. The 3D-integrated multilayered](http://pubs.acs.org/action/showImage?doi=10.1021/la402742f&iName=master.img-000.jpg&w=238&h=98) nanoparticle architecture successfully worked as a charge storage node in flash memory devices that exhibited improved charge storage capacity compared with that of a conventional monolayer structure device.

Controlled fabrication of three-dimensional (3D) arrays of bioinorganic hybrid materials using the ability of biomolecules to biomineralize is drawing considerable attention because of its potential to produce hierarchical structures for nanoelectronics and surface architectures for sensing devices and structural analysis. Hybrid bioinorganic matrices that are generated using a 3D protein template array encapsulated with biomineralized inorganic material allow versatile use of the inherent functionality of biomaterials for different applications.¹ Targeting of specific materials is one such functionality of biomaterials. Peptide aptamers that target inorganic materia[ls](#page-5-0) have inspired remarkable advances in engineering during the past decade, particularly with bottom-up fabrication at the nanometer scale.² The goal of biofunctionality-assisted fabrication is the self-organization of nanoscale building blocks to allow downsizi[ng](#page-5-0) of large-scale integration (LSI) technology.3[−]⁵ The sophisticated ability of biomolecules to selforganize in the nanometer regime has encouraged us to fabr[icat](#page-5-0)e hierarchical nanoarchitectures using the automated molecular interactions of functional biomolecules.^{6−8}

Peptide evolution systems have played an important role in the effort to realize this goal because they use [a n](#page-6-0)umber of target inorganic materials that can be specifically recognized by biomolecules.^{9,10} We previously reported isolation of peptide aptamer titanium binding peptide-1 (TBP-1) that discriminates and binds to [the](#page-6-0) surface of specific materials.^{11,12} For example,

TBP-1 selectively recognizes and binds to Ti, Ag, and Si surfaces, but not to those of Au, Cr, Pt, Sn, Zn, Cu, or Fe. TBP-1 also promotes mineralization of titanium oxide (titania), silicon oxide (silica), and Ag.^{12,13} Thus, TBP-1 is a bifunctional peptide, as is often the case for peptide aptamers that target inorganic materials. Althoug[h TB](#page-6-0)P-1 is composed of 12 amino acids, it has been revealed that the first hexapeptide in the sequence, Arg-Lys-Leu-Pro-Asp-Ala, mediates specific binding and the mineralization function of this peptide aptamer.¹¹ We have named this hexapeptide aptamer "minTBP-1". The bifunctionality of minTBP-1 can be endowed on [o](#page-6-0)ther molecules both by genetic and chemical display at their Nterminals.14−¹⁶ We equipped a protein with the bifunctionality of minTBP-1 by addition of the peptide aptamer to a protein molecule [by m](#page-6-0)aking a recombinant protein, minT1-LF [Figure S1, Supporting Information (SI)]. This protein possesses genetically displayed minTBP-1 on the surface of recombinant L-fer[ritin molecules. MinT1-L](#page-5-0)F showed a greater affinity for target silica and drastically lowered the necessary protein concentration for silica mineralization compared with original $minTBP-1$ peptide.^{14,16} The biomineralization function of the aptamer enabled a novel method to produce multilayered

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Figure 1. Schematic diagram of the BioLBL process used to fabricate MOS-based flash memory devices. The first monolayer of Fe@minT1-LF is formed on the cleane[d silicon substrate. A silica layer is deposited on top of the layer of minT1-LF via mineralization mediat](http://pubs.acs.org/action/showImage?doi=10.1021/la402742f&iName=master.img-001.jpg&w=352&h=131)ed by minTBP-1. The newly formed silica layer then serves as a binding target for a second Fe@minT1-LF layer. By repeating these binding and mineralization cycles, multilayered structures are fabricated. Following this BioLBL process, ferritin shells were removed by UV oxidation. After the layers of Fe-BNDs were embedded in a control gate layer by plasma CVD, aluminum electrodes were deposited on both sides of the substrate.

Figure 2. Fabrication [of multilayers of BND by BioLBL. \(A\) TEM image of aurothioglucose-stained Fe@minT1-LF. M](http://pubs.acs.org/action/showImage?doi=10.1021/la402742f&iName=master.img-002.jpg&w=349&h=271)inT1-LF shells were unstained (in white). (B) Time-dependent changes in resonance frequency (f) during QCM analysis. The values on the longitudinal axis represent the negative of Δf . Changes associated with adsorption of minT1-LF and silicification are shown as green and gray lines, respectively. Green and gray arrows represent the respective time points at which minT1-LF and silicic acid, respectively, were added to the measurement chamber. The black arrows indicate the time points at which the QCM measurement cell was washed with buffer solution. SEM images of samples after fabrication of the (C) first, (D) second, and (E) third Fe@minT1-LF layer.

protein−inorganic nanoparticle architectures by biomimetic layer-by-layer assembly (BioLBL) to be developed.¹⁶ BioLBL alternately utilizes the binding and silicification capabilities of minT1-LF to assemble multilayered nanostructures [co](#page-6-0)mposed of layers of minT1-LF and intercalating silica. In addition, we have established the X−Y positioning of mutant ferritin displaying a peptide aptamer and controlled stacking in the Z-dimension through the specific binding of minT1-LF and BioLBL.¹³

Ferritin is a cage-shaped protein with an interior space of 7− 8 nm.¹⁷ [It](#page-6-0) is able to accommodate a wide variety of inorganic nanoparticles including hydrated and oxidized metals and

semiconductors via biomineralization.^{18−20} MinT1-LF also showed the ability to incorporate ferrihydrite, cobalt oxide, and cadmium selenide into its interior [space](#page-6-0) via biomineralization.¹⁶ Multiple layers of inorganic nanoparticles separated by intervening layers of ferritin can be constructed by BioLBL. The [m](#page-6-0)ineral layer between protein layers prevents interlayer diffusion of minT1-LF, and nanoparticles in ferritin cages are aligned in the layer adjacent to the substrate with the same density. Fabrication of such alternating nanometer-scaled layered structures is difficult to achieve using conventional layer-by-layer methods because of interlayer diffusion.¹⁶ The satisfactory layer control of BioLBL allows us to fabricate

heterogeneous multilayer structures using minT1-LF containing different nanoparticles.¹⁶

In this work, we propose a novel approach to fabricate selfassembled nanoparticle a[rch](#page-6-0)itectures by BioLBL to produce metal oxide−semiconductor (MOS) flash memory devices. We fabricate nanoparticle multilayers of ferrihydrite bionanodot (Fe-BND)-containing minT1-LF (Fe@minT1-LF) by BioLBL and evaluate its quality for use in electronic devices. The fabricated hierarchical nanoparticle multilayer structure is embedded in a MOS capacitor structure for use as the charge storage node of floating nanodot gate-type flash memory (Figure 1). Fabrication of nanoparticle-based charge storage nodes by BioLBL offers advantages compared to conventional chemica[l](#page-1-0) vapor deposition $(CVD)^{21,22}$ and ion implantation methods,²³ in which control of the size, shape, position, and density of nanoparticles is difficult [to a](#page-6-0)chieve. Use of a sizerestricte[d p](#page-6-0)rotein template allows control of nanoparticle size, and specific chemical and biological modification of the substrate surface and protein outer surface enables controlled placement in a desired area on the nanometer scale.^{24−26} Furthermore, BioLBL enables the 3D integration of a charge storage node layer in the Z -direction, 13 which is quite di[ffi](#page-6-0)c[ult](#page-6-0) using conventional fabrication methods but can be achieved by simple chemical reaction in solution [usi](#page-6-0)ng a BioLBL approach. We fabricate a MOS memory device with a 3D-integrated memory node and confirm that the 3D node can be used as a charge storage node of flash memory.

■ RESULTS AND DISCUSSIONS

Natural biomolecules usually need a high concentration of alkaline metal ions such as sodium and potassium present to allow mineralization in solution; TBP-1 and minT1-LF are no exception.^{13,16} However, contamination with alkaline metal ions degrades a performance of the resulting LSI devices because o[f the](#page-6-0) current leakage caused by persisting metal ions. Therefore, we need to decrease ion concentration and optimize solution conditions of the BioLBL process so that they are compatible with LSI. We first searched for an appropriate solution composition that allows minT1-LF to form minerals from prehydrolized tetramethoxysilane (TMOS) without alkaline metal ions. We found that including 50−100 mM of Tris-phosphate buffer enabled silica to mineralize without alkaline metal ions. Although minT1-LF showed inferior binding and mineralization activity under these conditions compared with those of conventional mineralization involving a millimolar concentration of alkaline ions in solution, sufficient binding of minT1-LF to the substrate and subsequent silica film formation were confirmed. This indicates that use of 50−100 mM Tris-phosphate buffer is suitable for fabrication of memory nodes for LSI devices. We confirmed that BioLBL can also be performed using minT1-LF containing Fe-BNDs, Fe@minT1- LF (Figure 2A). Formation of multilayered nanostructures during the BioLBL process was confirmed by monitoring the frequency ch[an](#page-1-0)ges of a quartz crystal microbalance (QCM). A profile of resonance frequency changes of the QCM (Figure 2B) was similar to that observed previously.¹⁶ Initial injection of Fe@minT1-LF solution into the QCM measurement cell [ca](#page-1-0)used the resonance frequency to decrease [b](#page-6-0)y approximately 100 Hz; this reduction in resonance frequency corresponds to binding of Fe@minT1-LF molecules at 1.7 μ g/cm², equivalent to monolayer deposition.¹⁶ To confirm this, we also observed surfaces of the first Fe@minT1-LF layer by SEM (Figure 2C). After the first Fe@minT[1-L](#page-6-0)F monolayer formed, we injected a

solution containing prehydrolyzed TMOS, which caused an initial rapid decrease followed by a gradual decrease in resonance frequency, reflecting the formation of a silica layer by minT1-LF-mediated silicification. We continued the silicification reaction for approximately 20 min, which was twice as long as that required previously because the rate of silica formation under alkali ion-free conditions was about half that under conventional conditions in the presence of alkali ions. When the QCM cell was refilled with Fe@minT1-LF solution after washing out the TMOS solution, the resonance frequency again decreased by 100 Hz, indicating the deposition of a second monolayer of Fe@mint1-LF on top of the silica layer. Subsequent cycles of injection of TMOS and Fe@minT1- LF solutions caused the changes in resonance frequency, indicating the successful stepwise deposition of alternate layers of Fe@minT1-LF and silica. The frequency change caused by the binding of Fe@minT1-LF is negligible for each layer formation, and therefore, the amount of Fe@minT1-LF adsorbed on each layer is considered to be almost the same. The reproducibility of this BioLBL process was excellent. Those facts suggest that we can produce 3D nanoparticle arrays with consistent node density for use as memory devices.

A commercially available p-Si (100) substrate (~10 Ω cm) with a 3-nm-thick thermally oxidized tunneling $SiO₂$ layer was used to fabricate MOS memory. The $SiO₂$ surface was cleaned with piranha solution and UV/ozone treatment before the BioLBL process. The cleaned silicon substrate was incubated in Fe@minT1-LF solution (0.2 mg/mL) in 100 mM Trisphosphate buffer at pH 8.0 for 10 min at room temperature and then washed with the same buffer solution. The substrate was then incubated in silicification solution containing 100 mM TMOS, which was prehydrolyzed with 1 mM inorganic phosphate, and 100 mM Tris-phosphate buffer for 20 min at room temperature. After terminating the silicification reaction, the substrate was thoroughly washed with buffer solution to remove residual chemicals. The BioLBL procedure was repeated until the desired number of Fe@minT1-LF layers was formed. Figure 2C−E shows scanning electron microscopy (SEM) images of deposited Fe-BNDs immobilized on the substrate during th[e](#page-1-0) BioLBL process. Small particles observed in SEM images correspond to Fe-BNDs of Fe@minT1-LF. An array of multilayered Fe-BNDs was successfully fabricated an increasing density of particles with each layer number increase. Analysis of the SEM images revealed that the aerial density of Fe-BNDs in a single layer is ~6.7 × 10^{11} cm⁻², which is comparable to the adsorption density of BNDs for MOS memory devices that showed large charge storage capacity reported previously.27,28 Arial density is an important issue to observe for memory operation on floating nanodot gate-type memory. If the dot [num](#page-6-0)ber is too small, we cannot observe memory operation even though BNDs can store the charge. The fabricated BioLBL memory contains a sufficiently large number of BNDs in each layer, so we can expect to observe charge storage behavior in the BioLBL memory. Following the BioLBL process, outer protein shells were removed by UV oxidation under ozone. Even though the Fe-BNDs are covered with a thin film of mineralized silica, X-ray photoemission spectra (XPS) of carbon and nitrogen suggest that organic residues are removed by chemical decomposition upon UV irradiation (Figure S2, SI). Because the protein shell can be removed by oxidation, the mineralized silica layer is considered porous compared with [t](#page-5-0)hermally oxidized $SiO₂$. Fabricated layers of Fe-BNDs were embedded in a $SiO₂$ control gate layer

Figure 3. Cros[s-sectional TEM images of BND structures fabricated by BioLBL. TEM image of \(A\) monolayer and \(B\) bilayer BND](http://pubs.acs.org/action/showImage?doi=10.1021/la402742f&iName=master.img-003.jpg&w=400&h=337) structures for use in flash memory devices. High-resolution TEM images of (C) monolayer and (D) bilayer BND structures, respectively. (E) EDS mapping of a cross-section of the bilayer BND structure in a flash memory device.

with a thickness of 20 nm, and aluminum electrodes were deposited on both sides of the substrate to make a MOS capacitor device. After deposition of electrodes, devices were annealed at 450 °C under reductive gas atmosphere ($H_2:N_2$ = 10%:90%) for 1 h to improve electrode contact and convert Fe-BNDs to Fe metal.²⁹ The fabricated MOS capacitor structure was evaluated by transmission electron microscope (TEM) observation of ultr[ath](#page-6-0)in cross sections of the prepared MOS capacitors prepared by ion-milling. TEM images of a monolayer of Fe-BNDs in the MOS stacked structure depicted in Figure 3A,C clearly show that the embedded Fe-BNDs were almost the same shape and size as prepared ones. The BNDs formed a monolayer in the gate $SiO₂$ without merging after device fabrication. Cross-sectional TEM images of a bilayer MOS capacitor structure show that a multilayered architecture of Fe-BNDs can be fabricated easily using BioLBL (Figure 3B,D). Structural analysis of the MOS capacitor by high-resolution scanning TEM (STEM) analysis coupled with energydispersive X-ray spectroscopy (EDS) elemental mapping (Figure 3E) clearly illustrates that embedded BNDs did not diffuse into the gate oxide; thus, a layered nanoparticle structure may not be fabricated by both the conventional layer-by-layer technique and conventional LSI process. TEM analysis also indicated that the nanostructure fabricated by BioLBL was robust through the device integration process. These results confirm that multilayered charge storage node arrays for MOS

memory devices can be fabricated by a simple, wet process BioLBL.

The charge storage characteristics of the fabricated singlelayer BioLBL memory were evaluated by high-frequency capacitance−voltage (C−V) measurements (Figure 4A). A C−V curve obtained under a gate voltage (V_G) sweep between −1 and +1 V, a voltage range in which no charge storag[e](#page-4-0) can be achieved, shows a sharp transition from accumulation to inversion without hysteresis and corresponds well with the C− V plot of a control sample without Fe-BNDs. Namely, the observed curve corresponds to the quasineutral state (QNS) of embedded BND memory nodes. In contrast, when positive (+10 V, 10 s) or negative (−10 V, 10 s) pulsed V_G is applied before a V_G sweep, the C−V curve shifted toward the positive (blue line in Figure 4A) or negative (red line in Figure 4A) direction, respectively, versus the QNS. A C−V curve shift, i.e., a flat-ba[n](#page-4-0)d voltage (V_{FB}) (V_{FB}) (V_{FB}) shift, is typically observed when an electron and/or hole is injected to the floating gate. Therefore, the observed V_{FB} shifts clearly indicate that V_G polaritydependent electron and hole confinement occurs in the embedded BNDs, as we previously observed for BNDembedded MOS memory. 2^7 We should note that MOS memory without Fe-BNDs did not exhibit any V_{FB} shift caused by charge storage in the B[ND](#page-6-0)s. Negligible change in memory window size upon ac frequency variation from 1 kHz to 1 MHz during $C-V$ measurement also suggests that the observed V_{FB} shift can be attributed to charge confinement in the embedded

Figure 4. (A) Capacitance−voltage characteristics of single-layer Fe-BN[D-embedded BioLBL memory. Black, blue, and red lines de](http://pubs.acs.org/action/showImage?doi=10.1021/la402742f&iName=master.img-004.jpg&w=214&h=437)pict QNS C−V without charge injection and C−V after charge injection with +10 V and -10 V gate pulses for 10 s, respectively. (B) $C-V$ curves of BioLBL memory with different layer numbers. V_G was swept from -8 V to +8 V and then returned to -8 V. (C) Stored charge density for BioLBL memories with different layer numbers depicted in part B.

BNDs rather than surface charge trapping. Thus, the observed charge confinement-induced V_{FB} shift clearly indicates that the memory node array fabricated by BioLBL does work as a memory node. We also found that a plasma CVD process and postmetal annealing (PMA) could improve the quality of the porous silica layer formed by silicification, which is generally considered a poor gate oxide. The C−V characteristics of this MOS device showed a low interface density of 1.0×10^{11} cm⁻² eV[−]¹ , no frequency dispersion of capacitance density, and no hysteresis in the wide frequency range from 1 kHz to 1 MHz. Therefore, the quality of the control gate is high enough for memory applications, which is probably caused by the conformal deposition of control gate oxide by plasma CVD. $SiO₂$ deposited by plasma CVD can fill the pores in the silicified

silica layer and prevent undesired charge trapping in the gate oxide.

Interestingly, in contrast to previously reported Fe-BND memory^{29,30} and cobalt oxide BND memory,^{25,27} the BioLBL memory shows an asymmetric V_{FB} shift in CV curves. In previou[s case](#page-6-0)s, V_{FB} shifted symmetrically for [electr](#page-6-0)on and hole charging. In contrast, BioLBL memory shows a larger V_{FB} shift for electron charging than for hole. This suggests that BioLBL memory has inferior hole charging or retention ability compared with that for electrons. A similar tendency was observed for the charge retention characteristics of the device (Figure S3, SI), in which dissipation of programmed electrons and holes was monitored by change in capacitance and converted to V_{FB} change.²⁵ Electron and hole programming was perfor[med](#page-5-0) at +10 and −10 V, respectively, with 3 s pulsed bias. Time-dependent cha[ng](#page-6-0)e in capacitance was monitored at +1.0 and −0.4 V for electrons and holes, respectively. After electron programming, V_{FB} gradually decreased for the first 5000 s and then decreased rapidly after 5000 s. The difference between the V_{FB} of the electron-programmed device (+1.4 V) and V_{FB} of the QNS (−0.1 V) at 10⁴ s, i.e., the memory window size (ΔV_{FB}) , was still 1.4 V. From the aerial density of the embedded Fe-BNDs (\sim 6.7 × 10¹¹ cm⁻²), the stored charge density can be calculated as 1.8×10^{12} cm⁻². In contrast, V_{FB} of the hole-programmed device rapidly shifted to the positive voltage side and showed a small $\Delta V_{\rm FB}$ of 0.23 V at 10⁴ s. Although hole retention time was less than that of electrons, both charges can be retained for more than 10^7 s from extrapolation of the V_{FB} shift (inset in Figure S3, SI).

To determine the effect of the 3D-integrated multilayered array structure on device properties, BND me[mo](#page-5-0)ries with different numbers of layers, mono-, bi-, and trilayer, were fabricated and their CV characteristics measured, as shown in Figure 4C. One V_G sweep between -8 and $+8$ V starting from negative voltage was applied. The V_{G} -sweep-induced V_{FB} shift depends on the polarity of the applied voltage, as observed in Figure 4A, with anticlockwise hysteresis found for all devices. These results indicate that effective charge injection to the BNDs in the upper layers, namely, the second and third layers, can occur, so the BNDs effectively work as charge storage nodes, even though they are further away from the Si substrate surface than the first layer. As seen in Figure 4B, the curves of the fabricated multilayered memory devices showed a monotonic increase of ΔV_{FB} depending on the number of layers. Figure 4C depicts the change in charge density for devices with different layer number. The charge density for each device was calculated from the aerial density of embedded Fe-BNDs. Stored charge densities for single-, double-, and triple-layer memory devices was calculated to be 7.6 \times 10¹¹, 12.3×10^{11} , and 15.8×10^{11} cm⁻². SEM observation and QCM results indicate that the number density of Fe-BNDs in the fabricated memory increases linearly with layer number. Therefore, the observed linear increase of stored charge density, i.e., memory window size, is interrupted by the increase in the number of embedded BNDs in the charge storage node. This result clearly indicates that multilayered memory node arrays fabricated by the BioLBL method can be used as integrated memory nodes. We also made multilayered BND-embedded memory with a field effect transistor, which has been reported elsewhere.³¹ This study demonstrates that nanofabrication processes using a biological function can be adapted for LSI devices. De[vic](#page-6-0)e properties can be improved

using our BioLBL method because of the vertical buildup of charge storage elements.

■ CONCLUSION

A novel process to fabricate hierarchical nanoarchitectures of multilayered nanodots is demonstrated, and the nanoarchitectures are applied in memory devices. Memory nodes made up of biochemically synthesized BNDs in protein cavities were fabricated by BIoLBL using the specific binding and mineralization function of peptide aptamer minTBP-1 attached to the exterior of recombinant minT1-LF. MinT1-LF successfully aligned to form separate layers using the wet chemical process of BioLBL. The fabricated Fe-BND array was embedded into a stacked MOS device structure as a charge storage node. The MOS capacitor showed memory operation, and an increase of BND layer number increased the charge storage ability of the memory device. In this study, we only examined Fe@minT1-LF in MOS capacitors; however, ferritin can incorporate various kinds of nanoparticles in its cavity, not only by biomineralization but also by nanoparticle-directed assembly. By using different nanoparticles encapsulated in minT1-LF, we can fabricate heterogeneous layers of different nanoparticles with divergent electronic properties such as band gap.¹⁶ This approach can be used to develop novel types of multilayered memory devices. Moreover, applications of het[ero](#page-6-0)geneous nanoparticle multilayers made by the BioLBL method are not restricted to memory devices but can be applied to optoelectronics, battery cells, and so on.32[−]³⁴

■ METHODS

Preparation of minT1-LF. Construction, expression, and purification of minT1-LF have been described previously.¹⁴ Purified apo-minT1-LF molecules were filled with ferrihydrite nanoparticles as described to obtain Fe@minT1-LF.¹⁴ To remove alkaline [me](#page-6-0)tal ions, we carried out extensive dialysis against 100 mM Tris-phosphate buffer at pH 8.0. The concentration of Fe[@m](#page-6-0)inT1-LF was determined using the Bradford method using apo-minT1-LF as a calibration standard.

BioLBL without Alkaline Metal Ions. Silicification activities of apo-minT1-LF were monitored as described previously, 14 and we found buffer conditions for silicification that were alkaline metal ion free. We then monitored the BioLBL process using n[ew](#page-6-0) solution conditions on the quartz sensor of a QCM-D300 (Q-sense AB, Göteborg). The temperature of the chamber was kept at 25.00 ± 0.05 °C, and QCM data were collected at 14.8 MHz. We used a titaniumcoated sensor, which was purchased from Q-sense AB. Experimental conditions in Figure 2B were as follows. We first applied the Ti-coated sensor after filling the measurement cell with 0.2 mg/mL of Fe@ minT1-LF in 100 mM Tris-phosphate buffer at pH 8.0 and incubation for about 5 min. Th[e](#page-1-0) measurement cell was washed with same buffer solution, and then we introduced a silicification solution composed of 0.1 M TMOS, which was prehydrolyzed with 1 mM inorganic phosphate and 100 mM Tris-phosphate buffer. After about 20 min incubation, we washed the measurement cell with buffer solution. We repeated these steps of Fe@minT1-LF binding and silicification three times.

Preparation and Evaluation of MOS Capacitor with Floating Nanodot Gate. Commercially available p-Si (100) substrate (~10 Ω cm) with a 3-nm-thick thermally oxidized tunneling $SiO₂$ layer was used as the substrate for MOS capacitor fabrication. The $SiO₂$ surface of the substrate was made hydrophilic by UV irradiation under ozone prior to BioLBL. Layered memory nodes were fabricated by sequential binding and silicification steps following the BioLBL process described above. The protein component of Fe@minT1-LF, which is an impurity that could affect device operation, was removed by UV irradiation under ozone at 115 °C for 60 min. Each fabricated Fe-BND array was buried in a 17-nm-thick control gate oxide by plasma CVD

using a tetraethoxysilane as a source gas. Aluminum metal was used as the metal gate electrode of the MOS capacitors. PMA was performed in a reductive gas atmosphere (10% H_2 in 90% N_2) at 450 °C for 1h. It should be noted that the partial reduction of Fe-BNDs occurred during PMA under the reductive atmosphere. Electrical characterization of the fabricated Fe-BND-embedded BioLBL memory arrays was performed by capacitance−voltage (C−V) measurements with an HP 4284A LCR meter for MOS capacitors. All measurements were performed at room temperature.

Electron Microscope Observation. We conducted highresolution SEM (SEM7400F, JEOL) measurements to observe nanoparticle deposition by BioLBL. The SEM was operated at a low acceleration voltage of 5 kV to observe the spatial distribution of Fe-BNDs in the mineralized silica layer. Samples for cross-sectional samples for TEM and STEM observations were prepared as described previously (outsourced to TOPCON, Tokyo, Japan).¹⁶

■ ASSOCIATED CONTENT

S Supporting Information

More extensive characterization of multilayered devices, the elemental analysis, and the charge retention properties. This material is available free of charge via the Internet at http:// pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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