# 國立交通大學

# 奈米科技研究所

## 碩士論文

藉由範圍性奈米結構控制 H9c2 心肌纖維母細胞功能、增生、型態和貼附

Functional modulation of proliferation, morphology, and adhesion for H9c2 cardiomyoblasts by nanostructures

研究生:隋玉蘋

學 號:9752512

指導教授:黃國華 教授

中華民國九十九年七月

### 藉由奈米結構調控 H9c2 cardiomyoblasts

#### 功能、增生、型態和貼附

Functional modulation of proliferation, morphology, and adhesion for

H9c2 cardiomyoblasts by nanostructures

研究生:隋玉蘋 Student: Yu-Ping Sui

指導教授:黃國華 Advisor: Dr. Guewha Steven Huang

國立交通大學

奈米科技研究所

月工論义

A The sis

Submitted to Institute of Nanotechnology

College of Engineering

National Chiao Tung University in partial Fulfillment of the Requirements

for the Degree of

Master

in

Institute of Nanotechnology

July 2010

Hsinchu, Taiwan, Republic of China

中華民國九十九年七月

#### 藉由奈米結構調控 H9c2 cardiomyoblasts

#### 功能、增生、型態和貼附

學生:隋玉蘋 指導教授:黃國華 教授

#### 國立交通大學奈米科技研究所

#### 碩士論文

#### 摘 要

奈米尺度表面形態會直接的影響細胞的行為。為了明確定義出調控細胞生長的結構確切尺度範圍和型態,本研究將 H9c2 心肌纖維母細胞養在 10-nm 至 200-nm 的奈米點陣列上。奈米點陣列結構是藉由陽極氧化鋁製程(AAO)在 TaN 表面的矽晶圓上製備而成。

實驗結果發現,即使表面細胞數已經達飽和,cardiomyoblasts 在 50-nm 尺度上的 nanodots 仍有最佳的存活能力;100-nm 以及 200-nm 表面,細胞培養三天後數量明顯減少了 53.7%以及 72.6%。50-nm 表面的細胞於表面有較大的貼附面積、並且較多的伸展細胞本體、細胞較快的成長速率;100-nm 和 200-nm 表面細胞數目有明顯減少,並且伴隨著凋亡的產生。

根據螢光染色的 vinculin 和 actin filament 分布探討奈米表面對 focal adhesion 的影響,結構尺寸低於 50-nm(含 50-nm 尺度結構)會促進細胞的貼附、細胞骨架的完整,尤其是在 50-nm 結構上的細胞具顯著差異。Bromodeoxyuridine (BrdU) proliferation assay 也指出 50-nm 結構也會促進細胞的增生。

我們也更進一步使用 RT-PCR 和 Western blot, 針對基因還有蛋白質的分析,

結果指出,在 100-nm 的奈米點表面會誘發心肌細胞肥大和纖維化,我們推測可能是因為此奈米結構促成細胞的收縮功能降低,所引起的細胞肥大,纖維化的部分則是因為細胞間基質的降解所導致;高 vinculin 表現量則是發生在 50-nm nanodots,表示心肌纖維母細胞的貼附狀況良好。

根據實驗結果顯示, nanodots 結構調控 cardiomyoblasts 生長是與奈米結構尺寸大小有關的。心肌纖維母細胞生長在 50-nm 結構表面具有最佳的生存能力、型態、貼附、和增生能力;細胞生長在 100-nm 以上的結構表面則會降低細胞的生長和貼附。未來,預期可以利用此研究的發現,應用到人工植體之生物醫學層面。



Functional modulation of proliferation, morphology, and adhesion for

H9c2 cardiomyoblasts by nanostructures

Student: Yu-Ping Sui

Advisor: Dr. Guewha Steven Huang

Institute of Nanotechnology National Chaio Tung University

**ABSTRACT** 

Surface topology at nanoscale encodes information that directs cell behavior. To identify

the exact ranges in size and shape that modulates cell growth, H9c2 cells were cultured onto

nanodot arrays with dot diameters ranging from 10 to 200-nm. The nanodot arrays were

fabricated by anodic aluminum oxide (AAO) processing on TaN-coated wafers.

1896

Optimized growth occurred when cardiomyoblasts were cultured on nanodot arrays with

dot size at 50-nm, on which maximum viability was maintained even when the cell density

reached saturation. Nanodots of 100-nm and 200-nm prevented viable growth of

cardiomyocytes with 53.7% and 72.6% reduction on day 3, respectively. Cells seeded on

50-nm nanodots showed flat morphology, with largest surface area, most extended

lamellipodia, and fastest growth rate. Cell grown on flat surface remained stable in culture

dish, while apoptosis-like growth was observed with 100-nm and 200-nm nanodots with

significant reduction in the surface area. Immunostaining against vinculin and actin filament

iii

indicated that nanodots smaller than 50-nm promoted cell adhesion and cytoskeleton organization. Best adhesion occurred at 50-nm. Nanodots of 100-nm retarded the formation of focal adhesions while 200-nm inhibited the organization of cytoskeleton. Incorporation of Bromodeoxyuridine (BrdU) indicates proliferating growth of the cells. BrdU was applied to differentiate the newly proliferated cells from pre-existing culture. Maximum proliferation occurred for cells grown on 50-nm nanodots, which is approximately 2-fold compared to flat surface. We also utilize RT-PCR and Western blot to indicate fibrosis and hypertrophy were induced on 100-nm nanodots. We hypothesize the cardiomyoblasts seeded on 100-nm nanodots induced extracellular matrix-degrading metalloproteinases and promote fibrosis. High expression of vinculin was occurred in 50-nm nanodots. It means that 50-nm nanodots have good focal adhesion for cardiomyoblasts.

Here we show that the ability of nanodot arrays to modulate the growth of cardiomyoblasts is size-dependent. Optimized growth with the best viability, morphology, adhesion, and proliferation occurred at size of 50-nm. Retardation of growth was observed when the dot size was larger than 100-nm. Possible application of nanostructure on the artificial implants is expected.

*Keywords:* Cell adhesion; Nanotopography; viability; proliferation; cytoskeleton; cardiomyoblast; fibrosis ;hypertrophy ;Cardiovascular sten

#### 誌 謝

兩年的碩士班生活,即將進入尾聲。在這兩年中,很榮幸的跟隨 黃國華教授研究 奈米生醫,讓我多學習到生物這方面的領域。畢竟,現在奈米生醫是很熱門的研究領域, 而且現今社會最需要多方面領域擅長並且會做有效結合和創造的人才。雖然說自己並沒 有非常專精,但是過程當中,我學會了很多不只是生物方面的知識,也學會了做實驗的 態度和方法。

在研究生活當中,除了感謝日以繼夜、焚膏繼晷的教導我們的 黃國華教授之外,也很感謝師母 洪孟燕 女士,常常給予我鼓勵和關懷,我也不會忘記每當師母撥冗來看我們時,還會特別準備素食的餐點請我用,對待我們,就像家人一樣,也常常關懷我的身體狀況,自己雖然不是待在家裡,可是有受到像在家裡一樣的溫暖,我心裡真的充滿感激和感動。很感謝 洪耀欽 醫師常常關懷我的實驗還有身體狀況,自己之前實驗一直失敗的時候,常常抗壓性不足難以壓抑自己情緒,可是洪醫師就像爸爸一樣給我關懷和鼓勵,讓我打起精神來繼續努力,真的非常感謝洪醫師的關懷和照顧。

還有,也很感謝打從我一進入實驗室就開始帶我的敘安學長,雖然他對我很嚴格,以前當下心裡有點不是滋味,但是我很感謝他每一個實驗的起頭都是他在帶領我、陪我待到半夜、陪我一起想辦法解決問題,讓不是生物背景的我有方向去找尋問題的答案…等等很多很多,帶我突破困難;也謝謝大勳學長不厭其煩的幫忙我實驗的統整、借儀器等等;還有我同屆的夥伴:佳慧、洪寧、家偉、順華,陪我一起做實驗做到很晚、遇到挫折鼓勵我;還有感謝在我自己趕data時,碰到儀器壞掉,謝孟哲學弟一直幫忙借儀器、配溶液、幫忙做實驗…等等。還有,重要的是,我很感謝這兩年中一直很擔心我狀況的爸爸媽媽爺爺奶奶姑姑舅舅等家人和親戚,給我引導和鼓勵,送補品還有水果來給我。還有很感謝我的男朋友—董亦凇,他自己本身實驗也很忙碌,可是也常常給遇到困難挫折埋怨的我鼓勵,給予我實驗上的建議,陪我走過每一個關卡,一起成長一起提昇。

最後,我想說的是,還有很多很多我要感謝的人,也因為有了 黃國華教授、師母 洪孟燕女士、洪耀欽醫師、學長同學學弟、爸爸媽媽家人、好朋友…等等你們大家鼓勵 和栽培,才會有今日的隋玉蘋。



### Table of Contents

Chi	nese Abstract	i
Eng	lish Abstract	iii
Ack	nowledgement	V
Tab	le of Content	
List	of Tables	
List	of Figures.	
I.	Introduction	1
II.	Experiment and Methods	5
	2.1 Cell culture	5
	2.2 Chemicals	5
	2.3 Fabrication of nanodot arrays	5
	2.4 The cells viability assay	6
	2.5 Scanning electron microscopy (SEM)	7
	2.6 Immunostaining	7
	2.7 Bromodeoxyuridine (BrdU) proliferation assay	7
	2.8 Quantitative real-time RT-PCR	8
	2.9 Western blot	9
III.	Results and Discussions	11
	3.1Nanotopography modulated cell viability and proliferation of	
	Cardiomyoblasts	11
	3.2 Nanotopography modulated morphology of cardiomyoblasts	19
	3.3 Nanotopography modulated cell adhesion and cytoskeleton organization of	
	Cardiomyoblasts	24
	3.4 Nanotopography influence cell survival mRNA expression	28
	3.5 Nanotopography influence cell apoptosis mRNA expression	29
	3.6 Nanotopography influence cell hypertrophic and fibrosis mRNA expression	30
	3.7 Nanotopography influence vinculin, PAI-1 protein expression	31
IV.	Conclusion	35
Ref	Ference	35