A Synthesis of 6-Myoporol

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An approach to (\pm) -6-myoporol (1) featured stereocontrol by equilibration of 2,6-disubstituted cyclohexanones to mainly the *cis* isomer. The synthesis was completed by reaction of a lactone intermediate with 3-lithiofuran, Mitsunobu reaction, and hydrolysis.

6-Myoporol (1) is a stress metabolite of the furanoterpenoid type, isolated from slices of the root of sweet potato (Ipomoea batatas) that had been treated with 0.1% mercuric chloride or with spores of Ceratocystis fimbriata.1 The relative stereochemistry of 6-myoporol was established through a highly diastereoselective hydroboration² of (*E*)-2,6-dimethyl-1,4-heptadiene to afford a diol intermediate in which the secondary methyl substituent and the hydroxyl group are syn-related. After a rapid hydroboration of the terminal double bond, the second step became intramolecular, which followed a steric course as determined by conformational factors. Later, the absolute configuration of the natural (-)-6myoporol as (4S,6R)-1-(3-furanyl)-6-hydroxy-4,8-dimethyl-1-nonanone was determined by another synthesis³ featuring elaboration of (R)-2,6-dimethyl-1-hepten-4-ol by means of Lewis-acid-induced cleavage of a chiral 4,6dimethyl-1,3-dioxane with methallyltrimethylsilane as a nucleophile and subsequent disengagement of the γ -hydroxyalkyl unit inherited from the C_2 -symmetric diol. Hydroboration of the trifluoroacetate led to a 3:1 diastereomeric mixture, which could be separated into the components by HPLC. The major product was identified as the dextrorotatory form of the nine-carbon diol obtained from the previous synthesis.

Our synthetic approach to racemic 6-myoporol was based on the conformational behavior of 2,4-disubstituted cyclohexanones. Following the same stereochemical relationship of the corresponding cyclohexanes the cis isomers are expected to be more stable because both substituents are equatorial. Accordingly, we started our work by alkylation of the pyrrolidino enamine of 4-methylcyclohexanone with methallyl chloride to afford 2. The ratio of the *cis* and *trans* isomers was estimated by NMR to be 40:60, axial alkylation being favored. The same ratio was retained after hydrogenation. However, when the product was subjected to equilibration with 10% NaOH in hot MeOH, 3 was obtained in a cis-trans ratio of 84:16 according to ¹H-NMR data. As we feared the separated isomers would undergo isomerization again, the mixture was directly used in the subsequent Baeyer-Villger oxidation. With *m*-chloroperbenzoic acid in refluxing CHCl₃, the cis-lactone (4) was obtained in 73% yield. Based on the original content the actual yield from the cis-ketone was actually close to 87%. The trans-lactone was not isolated. (See Scheme 1.)

Reaction of the lactone (4) with 3-lithiofuran at -100 °C furnished *epi*-6-myoporol (5), which was separated from the very small amount of impurities.

Scheme 1. Synthesis of Racemic 6-Myoporol (1)

The configuration of the carbinolic center of the major product required inversion to complete the synthesis. Thus, a Mitsunobu reaction⁴ with benzoic acid as the nucleophile, was performed. The product was *dl*-6-myoporol benzoate (**6**), which was promptly saponified with lithium hydroxide in MeOH. *dl*-6-Myoporol (**1**) was obtained, showing spectral data consistent with those reported.²

Experimental Section

General Experimental Procedures. IR spectra of liquid samples were sandwiched between NaCl plates and taken from a JASCO FT/IR-200 spectrophotometer. $^1\text{H-}$ and $^{13}\text{C-NMR}$ spectra were recorded in CDCl $_3$ on a Varian UNITY-300 instrument at 300 and 75 MHz, respectively; chemical shifts are reported in parts per million downfield from TMS. The EIMS were obtained from a TRIO-2000 mass spectrometer operating at 70 eV ionization voltage, and HRMS were measured on a JEOL JMX-HX 110 mass spectrometer. Melting points were uncorrected. Merck Kieselgel 60 of 70–230 mesh was used for column chromatography. During workup of reactions, solutions were dried over anhydrous Na2-SO4 and evaporatred *in vacuo* using a Büchi rotary evaporator.

4-Methyl-2-(2-methyl-2-propenyl) cyclohexanone (2). A solution of 4-methylcyclohexanone (1.12 g, 10 mmol), pyrrolidine (0.78 g, 11 mmol), and a trace of p-toluenesulfonic acid in benzene (20 mL) was refluxed under a Dean-Stark trap for 8 h. After evaporation of solvent, the residual enamine was redissolved in dioxane (5 mL) and treated with 3-chloro-2-methylpropene (0.91 g) in dioxane (5 mL). The mixture was again refluxed for 6 h to complete the allylation, then hydrolyzed by the addition of 50% HOAc (10 mL) by continuing heating at reflux for 4 h. Dioxane was removed *in vacuo*, and the remaining mixture was taken up in Et_2O and washed with H_2O and aqueous NaHCO₃.

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The dried organic solution was evaporated to give a slightly yellow oil (1.36 g, 82% yield): IR (neat) ν 1712, 1650 cm⁻¹; NMR (300 MHz, CDCl₃) δ 0.96, 1.03 (total 3H, two d, J=6.6 Hz), 1.62 (3H, br s), 1.8–2.6 (10H, m), 4.64 (1H, d, J=12.3 Hz), 4.74 (1H, br s); M^{+•} calcd for C₁₁H₁₈O 166.1358, found 166.1353.

4-Methyl-2-(2-methylpropyl)cyclohexanone (3). 4-Methyl-2-(2-methyl-2-propenyl)cyclohexanone (1.0 g, 6 mmol) was dissolved in MeOH (10 mL), added to a flask containing 10% Pd-C (50 mg) and a stir bar, deaerated by evacuation, and placed under hydrogen. After 5 h the catalyst was filtered, and the solution was evaporated. The residue was redissolved in MeOH (20 mL), to which 10% aqueous NaOH (200 mg) was introduced, and the mixture was refluxed for 18 h. On cooling to room temperature, the base was neutralized with 10% HCl. and MeOH was removed in vacuo. The residue was distributed between H₂O and CH₂Cl₂, and the organic layer was separated, dried, and evaporated. Chromatography gave 3 as an oil (0.98 g, 98% yield): IR (neat) ν 1714 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.82 (3H, d, J = 10.2 Hz), 0.84 (3H, d, J = 10.2 Hz), 0.96 (3H, d, J = 6.6 Hz), 1.2 - 1.4 (1H, m), 1.4 - 1.7 (3H, m)m), 1.8-2.1 (3H, m), 2.2-2.5 (3H, m); M⁺ calcd for C₁₁H₂₀O 168.1515, found 168.1507.

cis-4,8-Dimethyl-6-nonanolide (4). A mixture of ketone **3** (1.0 g, 6 mmol), *m*-chloroperbenzoic acid (1.86 g, 10.8 mmol) in CHCl₃ (4 mL) was refluxed for 40 h. The cooled reaction mixture was filtered, filtrate evaoprated, the residue dissolved in Et₂O, washed with aqueous K₂CO₃ and brine, and dried. Concentration of the solution and chromatography of the resulting residue over a silica gel column furnished the oily lactone **4** (0.8 g, 73%): IR (neat) ν 1737 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.88 (3H, d, J = 6.6 Hz), 0.89 (3H, d, J= 6.6 Hz), 0.94 (2.52H, d, J = 6.6 Hz), 1.02 (0.48H, d, J= 7.5 Hz, 1.1-1.4 (4H, m), 1.6-2.0 (6H, m), 2.61 (2H, m)m), 4.29 (0.84H, m), 4.43 (0.16H, m); ¹³C NMR (75 MHz, $CDCl_3$) δ 20.50, 21.40, 21.99, 22.95, 30.12, 32.56, 33.58, 42.35, 44.16, 75.77, 173.67; M⁺ calcd for C₁₁H₂₀O₂ 184.1464, found 184.1462.

(±)-*epi*-6-Myoporol (5). A solution of 3-lithiofuran formed by treatment of 3-bromofuran (600 mg, 4 mmol) in THF (8 mL) with *n*-butyllithium (2.5 M in hexanes, 1.4 mL) was stirred at -100 °C for 0.5 h; the temperature was raised to -78 °C for 1 h and recooled to -100 °C. The lactone (4) was added via syringe, and the reaction was allowed to proceed for 1 h at the same temperature, then quenched with saturated NH₄Cl solution. Ether extraction workup followed by column chromatography (eluent, 1:4 EtOAc—hexane) afforded (±)-*epi*-6-myoporol (5) (449.5 mg, 65.5%): mp 59–61 °C IR ν 3407, 1681 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ

0.88–1.0 (9H, m), 1.15–1.3 (2H, m), 1.3–1.5 (3H, m), 1.6–1.8 (4H, m), 2.74 (2H, t, J=7 Hz), 3.77 (1H, m), 6.74 (1H, d, J=2.1 Hz), 7.41 (1H, d, J=2.1 Hz), 8.00 (1H, s); 13 C NMR (75 MHz, CDCl₃) δ 19.16, 22.04, 23.21, 24.40, 28.98, 31.86, 37.92, 44.92, 47.51, 67.29, 108.46, 127.46, 143.98, 147.04, 195.52; M^{+*} calcd for $C_{15}H_{24}O_{3}$ 252.1726, found 252.1727.

(\pm)-6-Myoporol Benzoate (6). To a cold (-13 °C), magnetically stirred mixture of *epi*-6-myoporol (30 mg), triphenylphosphine (170 mg), and benzoic acid (60 mg) in THF (1 mL) under a nitrogen atmosphere was slowly added a solution of diethyl azodicarboxylate (128 mg) in THF (1 mL) via a syringe. After 1 h the reaction was allowed to warm to room temperature and stand for 1 day. The solvent was evaporated, and the residue was chromatographed (eluent, 1:9 EtOAc-hexane) to give (\pm)-6-myoporol benzoate (36.6 mg, 86%): IR (neat) ν 1714, 1675 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.88 (6H, d, J = 4.5 Hz), 0.95 (3H, d, J = 5.7 Hz), 1.2–2.0 (8H, m), 2.68 (2H, t, J = 7.5 Hz), 5.30 (1H, m), 6.70 (1H, s), 7.40 (3H, t, J = 7.5 Hz), 7.49 (2H, t, J = 7.5 Hz), 7.99 (1H, s), 8.00 (1H, d, J = 7.5 Hz); ¹³C NMR (75 MHz, $CDCl_3$) δ 19.89, 22.14, 23.24, 24.64, 29.42, 31.20, 38.19, 42.18, 43.73, 71.73, 108.58, 128.25, 129.48, 130.64, 132.68, 144.01, 147.10, 166.19, 195.12; M⁺• calcd for C₂₂H₂₈O₄ 356.1988, found 356.1968.

(±)-6-Myoporol (1). A mixture of **6** (97 mg) and lithium hydroxide (31.5 mg) in MeOH (4 mL containing 0.5 mL of H₂O) was refluxed for 18 h. After removal of solvent the product was dissolved in Et₂O, which was washed with NaHCO₃ solution and H₂O, dried over anhydrous NaSO₄, and evaporated. Chromatography (eluent, 1:4 EtOAc—hexane) furnished **1** (51 mg, 75%): IR (neat) ν 3440, 1674 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.86—0.92 (9H, m), 1.19 (2H, m), 1.37 (4H, m), 1.65 (1H, m), 1.80 (1H, m), 1.88 (1H, m), 2.75 (2H, t, J= 7.5 Hz), 3.81 (1H, m), 6.72 (1H, d, J= 2.1 Hz), 7.39 (1H, d, J= 2.1 Hz), 8.01 (1H, s); ¹³C NMR (75 MHz, CDCl₃) δ 20.35, 22.04, 23.50, 24.50, 28.98, 29.80, 37.75, 45.41, 46.99, 67.06, 108.58, 127.63, 144.04, 147.01, 195.41; M⁺⁺ calcd for C₁₅H₂₄O₃ 252.1725, found 252.1707.

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References and Notes

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