

國立交通大學

生物資訊研究所

碩士論文

從蛋白質序列預測殘基相對溶劑可接觸性

**Prediction of Protein Relative Solvent Accessibility
from Amino Acid Sequence**



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中文摘要

從序列資訊來預測蛋白質三級結構是目前生物學研究上非常重要的目標之一，而正確的預測蛋白質相對溶劑可接觸性則可以提供蛋白質三級結構相關的資訊。蛋白質相對溶劑可接觸性(RSA)代表著蛋白質上某一個氨基酸和溶劑接觸的程度。通常蛋白質的結合處會位於它的表面，因此，若能正確的預測蛋白質位於表面的氨基酸位置，就能夠更進一步的瞭解該蛋白質的功能。此外，一個蛋白質位於表面和包埋在蛋白質內部的氨基酸分佈，也被觀察到和蛋白質在細胞內的位置有很大的關連性。

我的方法是利用支持向量機將局部和整體的蛋白質資訊，其中最好的結果是利用位置加權矩陣(PSSM)、二級結構特徵值(secondary structure profile)和氨基酸親水程度(hydrophathy indexes)作為輸入向量。這個方法對於RS126 資料群在以 25% 為分類閾值時，可以達到 77.2%，和最近幾年的在這方面的研究成果 75%-78.3% 達到相近的程度，而在將 RSA 分成十類的預測結果中也可達到 15.2% 平均絕對誤差，預測值和實驗值達到 0.51 的相關性。

Prediction of Protein Relative Solvent Accessibility from Amino Acid Sequence

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Abstract

The prediction of the three-dimensional structure from its sequence is probably one of the most important goals of modern biology. The accurate prediction of protein relative solvent accessibility is useful for the prediction of tertiary structure of a protein. Amino acid solvent accessibility is the degree to which a residue in a protein is accessible to a solvent molecule. Because the binding sites of a protein are usually located on its surface, accurately predicting the surface residues can be regarded as an important step toward determining its function. On the other hand, it has been observed that the distribution of surface residues of a protein is correlated with its subcellular environments; consequently, information of surface residues may improve the prediction of protein subcellular localization.

Presently, our best method is based on the support vector machines using as the input feature vectors, the sliding window that includes the local environment descriptors such as PSSM, secondary structure profile and hydrophathy indexes. In my work, relative solvent accessibility based on a 2-state model, for 25%, 16%, 5%, and 0% accessibility are predicted at 77.2%, 77.1%, 80.4%, and 88.4% accuracy, respectively. Furthermore, solvent accessibility prediction methods have in recent years reached accuracy in the range of 75.0-78.3% at 25% threshold. And the results based in a 10-state model can reach 15.2% mean absolute error and 0.51 correlations.

誌謝

在求知的路上總是要一直不斷的經歷各種學習的過程，才能不斷的成長進步。在研究所二年的時間裡，各方面我都是收穫良多，真的很慶幸能身處在這樣一個優質的學習環境中體驗研究生生活。

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1. Introduction

Knowledge of protein's three-dimensional (3D) structure is essential for a full understanding of its functionality. However, only a small fraction of the enormous number of sequenced proteins has their structure determined. In order to reduce the gap between sequence and structure, developing reliable and applicable structure prediction methods has become a more important task in computational biology. Thus simplification of the problem from 3D structures to 1D feature may be useful as a first-step. The relative solvent accessibility of an amino acid in protein is a real value that represents the solvent exposed surface area of this residue in relative terms. The prediction of secondary structure is more familiar and well-defined aspect of the problem and the prediction of residue solvent accessibility is another aspect. Secondary structures and solvent accessibilities of amino acid residues give a useful insight into the structure and function of a protein. In particular, the knowledge of solvent accessibility has assisted alignments in regions of remote sequence identity for threading.

In addition to providing insight into the conformation of 3D structure, prediction of residue solvent accessibility has many other applications. For example, it has been observed that the distribution of surface residues of a protein is correlated with its subcellular environment. Many studies[1] have suggested that hydrophobic core residues are likely sites of deleterious mutations. The residues in site of deleterious mutations may be critical for protein stability. Wang and Molt[2] found that the vast majority of disease mutations affected protein stability rather than function and could be predicted using straightforward rules.

There are various approaches have been developed, typically by examining a window of residues centered at the test residue and using sequence input or sequence profile as input vectors. These include neural network[3-7], Bayesian statistics[8], linear regressions[9, 10],

information theory[11], support vector machines[12, 13] and fuzzy k-nearest neighbor method[14]. However, in contract to the secondary structure, there is no widely accepted criterion for classifying the experimentally determined solvent accessibility into a finite number of discrete states such as buried, intermediate and exposed states. Also, the prediction accuracies of solvent accessibility are lower than those for secondary structure prediction, since the solvent accessibility is less conserved than secondary structure[3], although there has been some progress recently.

In this work, relative solvent accessibility based on a 2-state model in RS126 dataset, for 25%, 16%, 5%, and 0% accessibility are predicted at 77.2%, 77.1%, 80.4%, and 88.4% accuracy, respectively. And the results based in a 10-state model can reach 15.2% mean absolute error and 0.51 correlations.



2. Material and Methods

2.1 Definition and thresholds of solvent accessibility

The relative solvent accessibility of an amino acid is the degree that present the residue in a protein is accessible to solvent molecules. The relative solvent accessibility can be calculated by the formula as follows,

$$RelACC = 100 \cdot \frac{ACC}{MaxACC} \quad (1)$$

RelACC : relative solvent accessibility

ACC : solvent accessible surface areas

MaxACC : maximum value of solvent accessible surface areas of each kind of residue

Where ACC of *i*th residue is the solvent accessible surface areas calculated by the dictionary of protein secondary structure program (DSSP) [15], and the MaxACC are taken from MOLSIM program (Table 1). RelACC is derived from normalizing ACC by dividing the DSSP accessibility by the maximum accessibility of amino acid residues corresponding to accessibility for a Gly-X-Gly extended tripeptide conformation. RelACC can hence adopt values between 0% and 100%, with 0% corresponding to a fully buried and 100% to a fully accessible residue, respectively.

Various thresholds have been used to classify residues as buried and exposed (2-state prediction) or buried, intermediate and exposed (3-state prediction) in previously published results. In this work, thresholds of 0, 5, 16, 25% for buried/exposed in the 2-state predictions, thresholds of 9, 36% for buried/intermediate/exposed in the 3-state predictions and 10, 20, 30, 40, 50, 60, 70, 80, 90% for classifying residues in 10 classes are used.

2.2 Training and data sets

The solvent accessibility was predicted for three sets of proteins in order to evaluate the performance of our method. They are a 126-protein dataset of Rost and Sander[3] (Table 2), a set of 215 protein chains selected by Naderi-Manesh[11] (Table 3), and 480 proteins from Kim and Park[13] (Table 4). Each of these data sets consists of protein sequence with less than 25% homology. To perform 7-fold cross validation test, we randomly divided these data sets into 7 groups, each containing a roughly equal number of protein sequences. One group was chosen as the testing set, while others were merged into the training set.

2.3 Feature vectors

In the first phase, we have individually benchmarked 8 different local descriptor types in prediction experiment in RS126 dataset. Our results revealed that the five best-performing sets of local descriptors were using PSSM (position-specific scoring matrices), secondary structure profiles, hydropathy indexes, amino acid composition and amino acid properties by their R groups. In second phase, the five top-performing local descriptors were combined to obtain a new method with improved performance. And then, we add 2 different global descriptors types into the method to see if global descriptors are useful for solvent accessibility prediction.

2.3.1 Local descriptors

We represent the local environment of sequences by using sliding window coding scheme. Increasing the window size can provide more local information and it is reasonable to expect that prediction accuracy would increase with the enlargement of the window size. But the past research[12] found that window size has a very limited effect on prediction accuracy.

Therefore, we followed the paper and selected 15 for the window size for the prediction of protein relative solvent accessibility

2.3.1.1 PSSM (position-specific scoring matrices)

We use PSI-BLAST[16] against non-redundant protein sequence database for 5 iterations to produce PSSM, which has $20*m$ elements, where m is the length of query sequences and each element represents the log-likelihood of that specific residue substitution at that position in the template (based on a weighted average of BLOSUM62 matrix scores for the given alignment position). PSI-BLAST uses a simple but effective scheme for weighting the contribution of locally different numbers of sequences to the resulting profiles. The profile matrix elements are scaled to the required 0-1 range by using the standard logistic function as below, where x of the equation is the raw profile matrix value.

$$Score = \frac{1}{1 + e^{-x}} \quad (2)$$



Then, the input vector has $20*15$ components (Illustrated in Figure 1). Each component can be expressed as $PSSM_{ijk}$, where i is the predicted residue, j is the location in the sliding window and k is the type of amino acid.

2.3.1.2 Secondary structure profiles

The secondary structure profile has $3*m$ elements. The elements describe the probabilities of each residue in the three kinds of secondary structures which are Helix, Sheet and Coil. These profiles were generated from the PSIPRED[17] program, and each input vector has $3*15$ components (Illustrated in Figure 2). Each component can be expressed as SS_{ijk} , where i is the predicted residue, j is the location in the sliding window and k is the type of secondary structure including helix,

strand and coil.

2.3.1.3 Hydrophathy indexes

In this study, we use the hydrophathy indexes to represent each residue (Table 5). In general, the hydrophathy indexes can be used to measure the tendency of an amino acid to seek an aqueous environment or a hydrophobic environment. The hydrophathy indexes refer to the book “Lehninger Principles of Biochemistry”. Each input vector has 1*15 components (Illustrated in Figure 3). Each component can be expressed as H_{ij} , where i is the predicted residue and j is the location in the sliding window.

2.3.1.4 Amino acid in binary coding

We code the protein sequences in binary mode, such as “1 0” represents A (alanine). Each input vector has 20*15 components (Illustrated in Figure 4). Each component can be expressed as AA_{ijk} , where i is the predicted residue, j is the location in the sliding window and k is the type of amino acid.

2.3.1.5 Amino acid properties in binary coding

Knowledge of chemical properties of the standard amino acids is central to an understanding of biochemistry. The chemical properties can be simplified by grouping the standard amino acids into 5 main classes based on the properties of their R groups, and others into the sixth class.(Table 6) The classification refer to the book “Lehninger Principles of Biochemistry”. Each input vector has 6*15 components (Illustrated in Figure 5). Each component can be expressed as $PROP_{ijk}$, where i is the predicted residue, j is the location in the sliding window and k is the property of amino acid.

2.3.1.6 Local amino acid composition

The amino acid percentage is the percentage of each residue in the sliding window. Each input vector has 20 components (Illustrated in Figure 6). Each component can be expressed as LAA_{ik} , where i is the predicted residue and k is the type of amino acid.

2.3.1.7 Local amino acid properties composition

The amino acid properties percentage is the percentage of each residue property in the sliding window. Each input vector has 6 components (Illustrated in Figure 7). Each component can be expressed as $LPROP_{ik}$, where i is the predicted residue, j is the location in the sliding window and k is the property of amino acid.

2.3.1.8 Residue size

We use the volume of amino acid as the input vector (Table 1). Each input vector has 1*15 components (Illustrated in Figure 8). Each component can be expressed as R_{ij} , where i is the predicted residue and j is the location in the sliding window.

2.3.2 Global descriptors

We also represent the global environment of sequences by using global sequence composition and global character composition method.

2.3.2.1 Global amino acid composition

The amino acid percentage is the percentage of each residue in a given protein sequence totally. Each input vector has 20 components (Illustrated in Figure 9). Each component can be expressed as GAA_k , where k is the type of amino acid.

2.3.2.2 Global amino acid properties composition

The amino acid properties percentage is the percentage of each residue property in a given protein sequence totally. Each input vector has 6 components (Illustrated in Figure 10). Each component can be expressed as $GPROP_k$, where k is the property of amino acid.

2.4 Performance measures

In order to compare with other works, we use different measures to evaluate 2-state, 3-state and 10-state prediction methods as below.

2.4.1 In 2-state prediction and 3-state prediction

In this work, two measures are used to evaluate the performance of prediction methods. One is accuracy, the percentage of correctly classified residues, and the other is Matthew's correlation coefficients (MCC). These measures can be calculated by the following equation:

$$Accuracy = \frac{\sum_i^c p_i}{N} \quad (3)$$

$$MCC_i = \frac{p_i n_i - o_i u_i}{\sqrt{(p_i + o_i)(p_i + u_i)(n_i + o_i)(n_i + u_i)}} \quad (4)$$

Where N is the total number of residues, and c is the class number. And p_i , n_i , o_i and u_i are the number of true positives, true negatives, false positives and false negatives for class i,

respectively. The MCCs have the same value for the two classes in the case of the 2-state prediction.

2.4.2 In 10-state prediction

Here, we applied several different measures, including the mean absolute error (MAE) of prediction defined as the absolute difference between the predicted and experimental value of relative solvent accessibility, per residue.

$$MAE = \frac{\sum |(ASA)_{pred} - (ASA)_{exp}|}{N} \quad (5)$$

Where summation is carried out for all residues and N is the total number of predictions. In addition, Pearson's "r" is also used in some places, and it is calculated as the ratio of the covariance between the predicted and experimental relative solvent accessibility as below.

$$r = \frac{N(\sum (ASA)_{pred} (ASA)_{exp}) - (\sum (ASA)_{pred})(\sum (ASA)_{exp})}{\sqrt{[N\sum (ASA)_{pred}^2 - (\sum (ASA)_{pred})^2][N\sum (ASA)_{exp}^2 - (\sum (ASA)_{exp})^2]}} \quad (6)$$

2.5 Support vector machine

Support vector machine (SVM), first proposed by Vapnik and co-workers[18] based on statistical learning theory, has quickly become one of the most popular classification and regression methods, due to its flexibility in choosing a similarity function, the ability to handle large feature spaces and accuracy. It has been used in various area, such as protein structure prediction[19], protein fold recognition[20] and microarray data analysis[21].

An SVM finds a nonlinear decision function in the input space by implicitly mapping the data into a linear separable higher dimensional feature space and separating the data there by maximizing the geometric margin and minimizing the training error at the same time. Yuan et al.⁵ implemented SVM^{light} to predict solvent accessibility. Here, we use LIBSVM version 2.6, which is a multi-class SVM developed by Chang and Lin.[22]



3. Server Development and Administration System Overview

Figure 2 shows the flowchart of RSA prediction server, and most of the process was built using the Perl language and CGI package. And the prediction result can automatically show on the webpage when the relative solvent accessibility prediction was finished.



4. World Wide Web Interface

4.1 Models trained by the three benchmark data sets

User can select an appropriate training dataset of the three benchmark datasets. The consuming time will increase depending on the dataset size.

4.2 Models of different state thresholds

User can choose a 2-state model (25%), 3-state model (9%;36%) or 10-state model (10%;20%;30%;40%;50%;60%;70%;80%;90%) for different purposes.

4.3 Query sequence

User can input their query sequence in FASTA format or directly upload the FASTA format file to the SAS prediction server.



4.4 Website

The RSA prediction server is available at <http://e100.life.nctu.edu.tw/~weilun/>. Figure3 shows the World Wide Web interface of RSA prediction server. Figure 4 shows the standard output webpage of prediction.

5. Results and Discussion

5.1 Prediction accuracies and MCC values using individual local descriptors with RS126 dataset in 2-state prediction model

In the first phase, the performance was evaluated in terms of Q_{total} , Q_b , Q_e and MCC (Table 7). Of the 8 local descriptors, 5 were ranging 65.73% to 75.33%. They are PSSM, amino acid composition, secondary structure profiles, character information and hydrophathy indexes in order.

5.2 Prediction accuracies and MCC values using comprehensive local descriptors with RS126 dataset in 2-state prediction model

In the second phase, we combined the top 5 best-performing local descriptors and 2 global descriptors in many kinds of combinations. In Table 8 we show the results.

Here, we found that PSSM involved more useful information in relative solvent accessibility prediction. The hydrophathy indexes performed worse than secondary structure profiles in the individual test, but when combining with PSSM, the hydrophathy indexes could increase the prediction accuracy more than the secondary structure profiles did. The combination of these three input vectors can reach the accuracy rate at 77.23% in RS-126 dataset. However, amino acid composition and amino acid properties also seem to offer duplicated information with PSSM, but the amino acid composition could slightly improve the prediction accuracy about 0.5%. The other feature vectors seem not to give useful information about prediction including global descriptors.

Overall, the best combinations of input vector were PSSM, secondary structure profiles, hydropathy indexes and amino acid composition in this stage. Compared with the individual performance of the 8 different descriptors, the increased accuracy was in the range 68.40% to 77.78%.

5.3 Prediction accuracies and MCC values of the benchmark test on the RS126, NM215 and KP480 datasets in 2-state prediction model and its comparison with other methods

The performances of prediction in this study are showed in Table9 and Table10. In this stage, the result of top-4 outperforming feature vectors is almost the same as the result of top-3 outperforming feature vectors. To reduce the training space, we took the top-3 outperforming feature vectors as our best work. They are PSSM, secondary structure profiles and hydropathy indexes. Our best work shows 60.2% accuracy for 3-state prediction (9%;36% thresholds) and 88.4%,81.4%,77.9% and 77.2% for the 2-state prediction with thresholds of 0%,5%,16% and 25% on the RS126 dataset, respectively. It shows slightly better prediction than other methods on the RS126 dataset expect for "Fuzzy k-NN".

5.4 MAE and Pearson's "r" values of the benchmark test on the RS126 and KP480 datasets in 10-state prediction model

We used the 10-state prediction model to simulate the real value prediction of solvent accessibility prediction. We took the same measured way to compare with the other real value prediction methods. All results were shown in Table11 and our best work achieved comparable results. The MAE value is 15.2 and Pearson's r is 0.506 in RS-126 dataset. We also transform the 10-state prediction into 2-state to give a rough evaluation. At the threshold of 20%, the accuracy achieved 72.8%, 74.4% in RS-126 and KP-480 dataset.

6. Conclusion

In this study, our best method achieved the similar performance as the researches did in the recent years. Hence, we can conclude that the three input vectors in our best work can catch enough information to predict relative solvent accessibility accurately.

Our research shows that local descriptors are sufficient for predicting accurately without global descriptors. Secondary structure profiles and hydropathy indexes both can be complementary to PSSM, but in individual case, hydropathy indexes can only achieve poor performance. Sequence information and character information offer overlapped information with PSSM.

In conclusion, our work achieves good prediction accuracy on the three benchmark datasets in 2-state, 3-state and 10-state model. And in the future work, we can apply our method in real value prediction.



Last position-specific scoring matrix computed, weighted observed percent

| | A | R | N | D | C | Q | E | G | H | I | L | K | M | F | P | S | T | W | Y | V |
|------|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| 1 T | -1 | -2 | -1 | -2 | -2 | -2 | -2 | -3 | -3 | -3 | -3 | -2 | -2 | -4 | -2 | 3 | 6 | -4 | -3 | -2 |
| 2 S | 0 | -3 | -2 | 1 | -3 | -1 | -2 | -3 | -3 | -3 | -3 | -2 | 4 | -4 | -3 | 5 | 2 | -5 | -4 | -1 |
| 3 P | -2 | 4 | -3 | -2 | -5 | 3 | 0 | -3 | -3 | -4 | 0 | 3 | -3 | -5 | 5 | -3 | -3 | -5 | -4 | -4 |
| 4 Q | -2 | 4 | -2 | -4 | -5 | 2 | -3 | 4 | 1 | -5 | -4 | 0 | 2 | -1 | -4 | -1 | 1 | 2 | 1 | -4 |
| 5 R | -2 | 4 | -1 | -2 | -5 | -3 | -2 | 3 | -2 | -3 | -3 | 3 | -4 | -2 | 4 | -1 | -2 | -5 | -5 | -5 |
| 6 E | -3 | 2 | 0 | -1 | -5 | 3 | 3 | -2 | 1 | -2 | -2 | 4 | -4 | -3 | -4 | -3 | 0 | -5 | -4 | -3 |
| 7 A | 3 | -2 | 3 | -5 | 4 | -4 | -4 | -2 | -5 | -1 | -1 | -4 | -3 | -5 | 6 | -3 | -2 | -6 | -5 | 2 |
| 8 T | -2 | 2 | 3 | 4 | -5 | -1 | 2 | -3 | 1 | -5 | -3 | 0 | 1 | -3 | -4 | 1 | 0 | -6 | -3 | -2 |
| 9 C | -2 | -6 | -3 | -6 | 11 | -5 | -6 | -2 | 0 | -3 | -3 | -5 | -4 | -5 | -5 | -3 | -3 | -5 | -5 | -3 |
| 10 T | -1 | -1 | 2 | -1 | -4 | -1 | -2 | -1 | -1 | -2 | -3 | -1 | 1 | -4 | 2 | 2 | 0 | -5 | -2 | 4 |
| 11 S | -2 | 0 | 2 | 0 | 9 | -2 | 1 | -2 | -2 | -3 | -3 | -1 | -2 | -4 | -3 | -1 | -3 | 0 | -3 | -1 |
| 12 E | 0 | -2 | 2 | 1 | 1 | 3 | 0 | -1 | -2 | -2 | -3 | -1 | -3 | 1 | 0 | -2 | -2 | -2 | 5 | -3 |
| 13 V | -2 | 1 | -2 | 2 | 1 | 0 | -1 | -3 | -3 | 0 | 1 | 2 | 0 | -1 | -3 | 0 | 1 | -4 | -1 | 0 |
| 14 S | -2 | -3 | 0 | 1 | 5 | 0 | 1 | -3 | 0 | -3 | -2 | -1 | 1 | -3 | 4 | 0 | -2 | -5 | -4 | 2 |
| 15 G | 1 | 1 | 0 | 1 | 2 | -2 | -1 | 2 | -1 | -2 | -3 | 0 | 1 | -4 | 1 | -2 | 1 | -4 | -4 | -1 |
| 16 C | -3 | -5 | -3 | -3 | 11 | -5 | -5 | -4 | -5 | -3 | -3 | -5 | -4 | -4 | -5 | -2 | -1 | -4 | -4 | -2 |
| 17 F | -2 | -1 | -2 | -1 | -4 | -1 | -3 | -2 | -1 | -4 | -3 | 0 | -2 | -3 | 6 | 1 | 3 | -5 | -4 | -4 |
| 18 K | -1 | 4 | 1 | 1 | -5 | 1 | -1 | -1 | 1 | -2 | -1 | 2 | 1 | -4 | 0 | -1 | -2 | -5 | -2 | -3 |
| 19 I | -2 | -2 | 2 | 2 | -5 | 1 | 4 | -3 | 2 | 2 | -2 | -3 | -2 | -3 | -3 | -1 | -2 | 0 | -2 | 0 |
| 20 Y | -3 | -3 | -3 | -2 | -5 | -2 | -1 | -3 | 0 | -2 | -1 | -2 | -2 | 3 | -3 | 0 | 0 | 3 | 7 | -2 |

Window of
15 rows

A R N D C Q E G H I L K M F P S T W Y V

| | | | | | | | | | | | | | | | | | | | | |
|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|--|
| 0.27 | 0.12 | 0.27 | 0.12 | 0.12 | 0.12 | 0.05 | 0.05 | 0.05 | 0.05 | 0.12 | 0.12 | 0.02 | 0.12 | 0.95 | 1.00 | 0.02 | 0.05 | 0.12 | | |
| 0.50 | 0.05 | 0.12 | 0.73 | 0.05 | 0.27 | 0.12 | 0.05 | 0.05 | 0.05 | 0.05 | 0.12 | 0.98 | 0.02 | 0.05 | 0.99 | 0.88 | 0.01 | 0.02 | 0.27 | |
| 0.12 | 0.98 | 0.05 | 0.12 | 0.01 | 0.95 | 0.50 | 0.05 | 0.05 | 0.02 | 0.50 | 0.95 | 0.05 | 0.01 | 0.99 | 0.05 | 0.05 | 0.01 | 0.02 | 0.02 | |
| 0.12 | 0.98 | 0.12 | 0.02 | 0.01 | 0.88 | 0.05 | 0.98 | 0.73 | 0.01 | 0.02 | 0.50 | 0.88 | 0.27 | 0.02 | 0.27 | 0.73 | 0.88 | 0.73 | 0.02 | |
| 0.12 | 0.98 | 0.27 | 0.12 | 0.01 | 0.05 | 0.12 | 0.95 | 0.12 | 0.05 | 0.05 | 0.95 | 0.02 | 0.12 | 0.98 | 0.27 | 0.12 | 0.01 | 0.01 | 0.01 | |
| 0.05 | 0.88 | 0.50 | 0.27 | 0.01 | 0.95 | 0.95 | 0.12 | 0.73 | 0.12 | 0.12 | 0.98 | 0.02 | 0.05 | 0.02 | 0.05 | 0.50 | 0.01 | 0.02 | 0.05 | |
| 0.95 | 0.12 | 0.05 | 0.01 | 0.98 | 0.02 | 0.02 | 0.12 | 0.01 | 0.27 | 0.27 | 0.02 | 0.05 | 0.01 | 1.00 | 0.05 | 0.12 | 0.00 | 0.01 | 0.88 | |
| 0.12 | 0.88 | 0.95 | 0.98 | 0.01 | 0.27 | 0.88 | 0.05 | 0.73 | 0.01 | 0.05 | 0.50 | 0.73 | 0.05 | 0.02 | 0.73 | 0.50 | 0.00 | 0.05 | 0.12 | |
| 0.12 | 0.00 | 0.05 | 0.00 | 1.00 | 0.01 | 0.00 | 0.12 | 0.50 | 0.05 | 0.05 | 0.01 | 0.02 | 0.01 | 0.01 | 0.05 | 0.05 | 0.01 | 0.01 | 0.05 | |
| 0.27 | 0.27 | 0.88 | 0.27 | 0.02 | 0.27 | 0.12 | 0.27 | 0.27 | 0.12 | 0.05 | 0.27 | 0.73 | 0.02 | 0.88 | 0.88 | 0.50 | 0.01 | 0.12 | 0.98 | |
| 0.12 | 0.50 | 0.88 | 0.50 | 1.00 | 0.12 | 0.73 | 0.12 | 0.12 | 0.05 | 0.05 | 0.27 | 0.12 | 0.02 | 0.05 | 0.27 | 0.05 | 0.50 | 0.05 | 0.27 | |
| 0.50 | 0.12 | 0.88 | 0.73 | 0.73 | 0.95 | 0.50 | 0.27 | 0.12 | 0.12 | 0.05 | 0.27 | 0.05 | 0.73 | 0.50 | 0.12 | 0.12 | 0.12 | 0.99 | 0.05 | |
| 0.12 | 0.73 | 0.12 | 0.88 | 0.73 | 0.50 | 0.27 | 0.05 | 0.05 | 0.50 | 0.73 | 0.88 | 0.50 | 0.27 | 0.05 | 0.50 | 0.73 | 0.02 | 0.27 | 0.50 | |
| 0.12 | 0.05 | 0.50 | 0.73 | 0.99 | 0.50 | 0.73 | 0.05 | 0.50 | 0.05 | 0.12 | 0.27 | 0.73 | 0.05 | 0.98 | 0.50 | 0.12 | 0.01 | 0.02 | 0.88 | |
| 0.73 | 0.73 | 0.50 | 0.73 | 0.88 | 0.12 | 0.27 | 0.88 | 0.27 | 0.12 | 0.05 | 0.50 | 0.73 | 0.02 | 0.73 | 0.12 | 0.73 | 0.02 | 0.02 | 0.27 | |
| 0.05 | 0.01 | 0.05 | 0.05 | 1.00 | 0.01 | 0.01 | 0.02 | 0.01 | 0.05 | 0.05 | 0.01 | 0.02 | 0.02 | 0.01 | 0.12 | 0.27 | 0.02 | 0.02 | 0.12 | |
| 0.12 | 0.27 | 0.12 | 0.27 | 0.02 | 0.27 | 0.05 | 0.12 | 0.27 | 0.02 | 0.05 | 0.50 | 0.12 | 0.05 | 1.00 | 0.73 | 0.95 | 0.01 | 0.02 | 0.02 | |
| 0.27 | 0.98 | 0.73 | 0.73 | 0.01 | 0.73 | 0.27 | 0.27 | 0.73 | 0.12 | 0.27 | 0.88 | 0.73 | 0.02 | 0.50 | 0.27 | 0.12 | 0.01 | 0.12 | 0.05 | |
| 0.12 | 0.12 | 0.88 | 0.88 | 0.01 | 0.73 | 0.98 | 0.05 | 0.88 | 0.88 | 0.12 | 0.05 | 0.12 | 0.05 | 0.05 | 0.27 | 0.12 | 0.50 | 0.12 | 0.50 | |
| 0.05 | 0.05 | 0.05 | 0.12 | 0.01 | 0.12 | 0.27 | 0.05 | 0.50 | 0.12 | 0.27 | 0.12 | 0.12 | 0.95 | 0.05 | 0.50 | 0.50 | 0.95 | 1.00 | 0.12 | |

15*20 scaled
inputs

Figure 1. An outline of the PSSM input vectors, which shows how the PSI-BLAST score matrices are processed.

| | | | Coil | Helix | Strand |
|----|---|---|-------------|--------------|---------------|
| 1 | G | C | 0.998 | 0.000 | 0.005 |
| 2 | K | E | 0.561 | 0.001 | 0.642 |
| 3 | I | E | 0.085 | 0.000 | 0.981 |
| 4 | T | E | 0.020 | 0.000 | 0.996 |
| 5 | F | E | 0.029 | 0.001 | 0.991 |
| 6 | Y | E | 0.080 | 0.005 | 0.969 |
| 7 | E | E | 0.340 | 0.010 | 0.770 |
| 8 | D | C | 0.944 | 0.005 | 0.089 |
| 9 | R | C | 0.907 | 0.029 | 0.030 |
| 10 | G | C | 0.962 | 0.020 | 0.006 |
| 11 | F | C | 0.885 | 0.019 | 0.070 |
| 12 | Q | C | 0.523 | 0.012 | 0.490 |
| 13 | G | E | 0.243 | 0.007 | 0.576 |
| 14 | R | E | 0.306 | 0.019 | 0.531 |
| 15 | H | E | 0.174 | 0.016 | 0.811 |
| 16 | Y | E | 0.131 | 0.056 | 0.756 |
| 17 | E | E | 0.236 | 0.054 | 0.551 |
| 18 | C | H | 0.223 | 0.342 | 0.231 |
| 19 | S | C | 0.454 | 0.216 | 0.134 |
| 20 | S | C | 0.582 | 0.206 | 0.066 |

} 15*3 scaled inputs

Figure 2. The input vectors called “secondary structure profiles”.

| AA | Hydropathy index | Scaled index |
|----|------------------|--------------|
| A | 1.8 | 1.8 / 4.5 |
| C | 2.5 | 2.5 / 4.5 |
| D | -3.5 | -3.5 / 4.5 |
| E | -3.5 | -3.5 / 4.5 |
| F | 2.8 | 2.8 / 4.5 |
| G | -0.4 | -0.4 / 4.5 |
| H | -3.2 | -3.2 / 4.5 |
| I | 4.5 | 4.5 / 4.5 |
| K | -3.9 | -3.9 / 4.5 |
| L | 3.8 | 3.8 / 4.5 |
| M | 1.9 | 1.9 / 4.5 |
| N | -3.5 | -3.5 / 4.5 |
| P | 1.6 | 1.6 / 4.5 |
| Q | -3.5 | -3.5 / 4.5 |
| R | -4.5 | -4.5 / 4.5 |
| S | -0.8 | -0.8 / 4.5 |
| T | -0.7 | -0.7 / 4.5 |
| V | 4.2 | 4.2 / 4.5 |
| W | -0.9 | -0.9 / 4.5 |
| Y | -1.3 | -1.3 / 4.5 |

| | | |
|----|---|------------|
| 1 | T | -0.7 / 4.5 |
| 2 | S | -0.8 / 4.5 |
| 3 | P | 1.6 / 4.5 |
| 4 | Q | -3.5 / 4.5 |
| 5 | R | -4.5 / 4.5 |
| 6 | E | -3.5 / 4.5 |
| 7 | A | 1.8 / 4.5 |
| 8 | T | -0.7 / 4.5 |
| 9 | C | 2.5 / 4.5 |
| 10 | T | -0.7 / 4.5 |
| 11 | S | -0.8 / 4.5 |
| 12 | E | -3.5 / 4.5 |
| 13 | V | 4.2 / 4.5 |
| 14 | S | -0.8 / 4.5 |
| 15 | G | -0.4 / 4.5 |
| 16 | C | 2.5 / 4.5 |
| 17 | P | 1.6 / 4.5 |
| 18 | K | -3.9 / 4.5 |
| 19 | I | 4.5 / 4.5 |
| 20 | Y | -1.3 / 4.5 |

} **15*1 scaled inputs**

Figure 3. The input vectors called “hydropathy indexes”.

| AA | ID | |
|----|----|--|
| A | 1 | 1 T 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 |
| C | 2 | 2 S 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 |
| D | 3 | 3 P 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 |
| E | 4 | 4 Q 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 |
| F | 5 | 5 R 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 |
| G | 6 | 6 E 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 |
| H | 7 | 7 A 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 |
| I | 8 | 8 T 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 |
| K | 9 | 9 C 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 |
| L | 10 | 10 T 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 |
| M | 11 | 11 S 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 |
| N | 12 | 12 E 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 |
| P | 13 | 13 V 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 |
| Q | 14 | 14 S 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 |
| R | 15 | 15 G 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 |
| S | 16 | 16 C 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 |
| T | 17 | 17 P 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 |
| V | 18 | 18 K 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0 |
| W | 19 | 19 I 0 0 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 |
| Y | 20 | 20 Y 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 1 |

} 15*20 scaled inputs

Figure 4. The input vectors called “amino acid in binary coding”.

| Class | ID | AA | |
|------------------------------|----|----|------------------|
| Nonpolar, aliphatic R groups | 1 | G | |
| | 1 | A | 1 T 0 0 1 0 0 0 |
| | 1 | P | 2 S 0 0 1 0 0 0 |
| | 1 | V | 3 P 1 0 0 0 0 0 |
| | 1 | L | 4 Q 0 0 1 0 0 0 |
| | 1 | I | 5 R 0 0 0 1 0 0 |
| | 1 | M | 6 E 0 0 0 0 1 0 |
| Aromatic R groups | 2 | F | 7 A 1 0 0 0 0 0 |
| | 2 | Y | 8 T 0 0 1 0 0 0 |
| | 2 | W | 9 C 0 0 1 0 0 0 |
| Polar, uncharged R groups | 3 | S | 10 T 0 0 1 0 0 0 |
| | 3 | T | 11 S 0 0 1 0 0 0 |
| | 3 | C | 12 E 0 0 0 0 1 0 |
| | 3 | N | 13 V 1 0 0 0 0 0 |
| | 3 | Q | 14 S 0 0 1 0 0 0 |
| Positively charged R groups | 4 | K | 15 G 1 0 0 0 0 0 |
| | 4 | H | 16 C 0 0 1 0 0 0 |
| | 4 | R | 17 P 1 0 0 0 0 0 |
| Negatively charged R groups | 5 | D | 18 K 0 0 0 1 0 0 |
| | 5 | E | 19 I 1 0 0 0 0 0 |
| Others | 6 | | 20 Y 0 1 0 0 0 0 |

} 15*6 scaled inputs

Figure 5. The input vectors called “amino acid properties in binary coding”.

| AA | ID | | | AA | N | P |
|----|----|----|---|----|---|--------|
| A | 1 | 1 | T | A | 1 | 1 / 15 |
| C | 2 | 2 | S | C | 2 | 2 / 15 |
| D | 3 | 3 | P | D | 0 | 0 |
| E | 4 | 4 | Q | E | 2 | 2 / 15 |
| F | 5 | 5 | R | F | 0 | 0 |
| G | 6 | 6 | E | G | 1 | 1 / 15 |
| H | 7 | 7 | A | H | 0 | 0 |
| I | 8 | 8 | T | I | 0 | 0 |
| K | 9 | 9 | C | K | 0 | 0 |
| L | 10 | 10 | T | L | 0 | 0 |
| M | 11 | 11 | S | M | 0 | 0 |
| N | 12 | 12 | E | N | 0 | 0 |
| P | 13 | 13 | V | P | 1 | 1 / 15 |
| Q | 14 | 14 | S | Q | 1 | 1 / 15 |
| R | 15 | 15 | G | R | 1 | 1 / 15 |
| S | 16 | 16 | C | S | 3 | 3 / 15 |
| T | 17 | 17 | P | T | 2 | 2 / 15 |
| V | 18 | 18 | K | V | 1 | 1 / 15 |
| W | 19 | 19 | I | W | 0 | 0 |
| Y | 20 | 20 | Y | Y | 0 | 0 |

} 20 scaled inputs

Figure 6. The input vectors called “local amino acid composition”.

| Class | ID | AA | | | Class | N | P |
|------------------------------|----|----|----|---|------------------------------|---|--------|
| Nonpolar, aliphatic R groups | 1 | A | 1 | T | Nonpolar, aliphatic R groups | 4 | 4 / 15 |
| | 1 | P | 2 | S | Aromatic R groups | 0 | 0 |
| | 1 | V | 3 | P | Polar, uncharged R groups | 8 | 8 / 15 |
| | 1 | L | 4 | Q | Positively charged R groups | 0 | 0 |
| | 1 | I | 5 | R | Negatively charged R groups | 1 | 1 / 15 |
| | 1 | M | 6 | E | Others | 2 | 2 / 15 |
| | | | 7 | A | | | |
| Aromatic R groups | 2 | F | 8 | T | | | |
| | 2 | Y | 9 | C | | | |
| | 2 | W | 10 | T | | | |
| Polar, uncharged R groups | 3 | S | 11 | S | | | |
| | 3 | T | 12 | E | | | |
| | 3 | C | 13 | V | | | |
| | 3 | N | 14 | S | | | |
| | 3 | Q | 15 | G | | | |
| Positively charged R groups | 4 | K | 16 | C | | | |
| | 4 | H | 17 | P | | | |
| Negatively charged R groups | 4 | R | 18 | K | | | |
| | 5 | D | 19 | I | | | |
| Others | 5 | E | 20 | Y | | | |
| | 6 | | | | | | |

6 scaled inputs

Figure 7. The input vectors called “local amino acid properties composition”.

| AA | MaxACC | Scaled MaxACC |
|----|--------|---------------|
| A | 117.2 | 117.2 / 270.7 |
| C | 142.0 | 142.0 / 270.7 |
| D | 169.8 | 169.8 / 270.7 |
| E | 202.0 | 202.0 / 270.7 |
| F | 233.0 | 233.0 / 270.7 |
| G | 87.9 | 87.9 / 270.7 |
| H | 195.4 | 195.4 / 270.7 |
| I | 182.1 | 182.1 / 270.7 |
| K | 214.2 | 214.2 / 270.7 |
| L | 176.2 | 176.2 / 270.7 |
| M | 204.0 | 204.0 / 270.7 |
| N | 169.6 | 169.6 / 270.7 |
| P | 148.9 | 148.9 / 270.7 |
| Q | 199.4 | 199.4 / 270.7 |
| R | 242.9 | 242.9 / 270.7 |
| S | 135.2 | 135.2 / 270.7 |
| T | 152.8 | 152.8 / 270.7 |
| V | 162.4 | 162.4 / 270.7 |
| W | 270.7 | 270.7 / 270.7 |
| Y | 253.8 | 253.8 / 270.7 |
| | | |

| | | |
|----|---|---------------|
| 1 | T | 152.8 / 270.7 |
| 2 | S | 135.2 / 270.7 |
| 3 | P | 148.9 / 270.7 |
| 4 | Q | 199.4 / 270.7 |
| 5 | R | 242.9 / 270.7 |
| 6 | E | 202.0 / 270.7 |
| 7 | A | 117.2 / 270.7 |
| 8 | T | 152.8 / 270.7 |
| 9 | C | 142.0 / 270.7 |
| 10 | T | 152.8 / 270.7 |
| 11 | S | 135.2 / 270.7 |
| 12 | E | 202.0 / 270.7 |
| 13 | V | 162.4 / 270.7 |
| 14 | S | 135.2 / 270.7 |
| 15 | G | 87.9 / 270.7 |
| 16 | C | 142.0 / 270.7 |
| 17 | P | 148.9 / 270.7 |
| 18 | K | 214.2 / 270.7 |
| 19 | I | 182.1 / 270.7 |
| 20 | Y | 253.8 / 270.7 |

15 scaled inputs

Figure 8. The input vectors called “residue size”.

| AA | ID | | | AA | N | P |
|----|----|----|---|----|---|--------|
| A | 1 | 1 | T | A | 1 | 1 / 20 |
| C | 2 | 2 | S | C | 2 | 2 / 20 |
| D | 3 | 3 | P | D | 0 | 0 |
| E | 4 | 4 | Q | E | 2 | 2 / 20 |
| F | 5 | 5 | R | F | 0 | 0 |
| G | 6 | 6 | E | G | 1 | 1 / 20 |
| H | 7 | 7 | A | H | 0 | 0 |
| I | 8 | 8 | T | I | 1 | 1 / 20 |
| K | 9 | 9 | C | K | 1 | 1 / 20 |
| L | 10 | 10 | T | L | 0 | 0 |
| M | 11 | 11 | S | M | 0 | 0 |
| N | 12 | 12 | E | N | 0 | 0 |
| P | 13 | 13 | V | P | 2 | 2 / 20 |
| Q | 14 | 14 | S | Q | 1 | 1 / 20 |
| R | 15 | 15 | G | R | 1 | 1 / 20 |
| S | 16 | 16 | C | S | 3 | 3 / 20 |
| T | 17 | 17 | P | T | 3 | 3 / 20 |
| V | 18 | 18 | K | V | 1 | 1 / 20 |
| W | 19 | 19 | I | W | 0 | 0 |
| Y | 20 | 20 | Y | Y | 1 | 1 / 20 |

} **20 scaled inputs**

Figure 9. The input vectors called “global amino acid composition”.

| Class | ID | AA |
|------------------------------|----|----|
| Nonpolar, aliphatic R groups | 1 | G |
| | 1 | A |
| | 1 | P |
| | 1 | V |
| | 1 | L |
| | 1 | I |
| | 1 | M |
| Aromatic R groups | 2 | F |
| | 2 | Y |
| | 2 | W |
| Polar, uncharged R groups | 3 | S |
| | 3 | T |
| | 3 | C |
| | 3 | N |
| | 3 | Q |
| Positively charged R groups | 4 | K |
| | 4 | H |
| | 4 | R |
| Negatively charged R groups | 5 | D |
| | 5 | E |
| Others | 6 | |

| 1 | T | Class | N | P | |
|----|---|------------------------------|---|--------|--------------------------|
| 2 | S | Nonpolar, aliphatic R groups | 6 | 6 / 20 | } 6 called inputs |
| 3 | P | Aromatic R groups | 1 | 1 / 20 | |
| 4 | Q | Polar, uncharged R groups | 9 | 9 / 20 | |
| 5 | R | Positively charged R groups | 1 | 1 / 20 | |
| 6 | E | Negatively charged R groups | 1 | 1 / 20 | |
| 7 | A | Others | 2 | 2 / 20 | |
| 8 | T | | | | |
| 9 | C | | | | |
| 10 | T | | | | |
| 11 | S | | | | |
| 12 | E | | | | |
| 13 | V | | | | |
| 14 | S | | | | |
| 15 | G | | | | |
| 16 | C | | | | |
| 17 | P | | | | |
| 18 | K | | | | |
| 19 | I | | | | |
| 20 | Y | | | | |

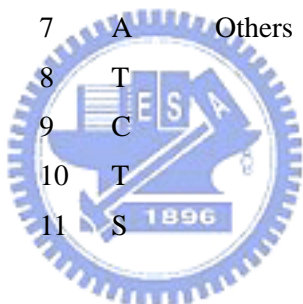


Figure 10. The input vectors called “global amino acid properties composition”.

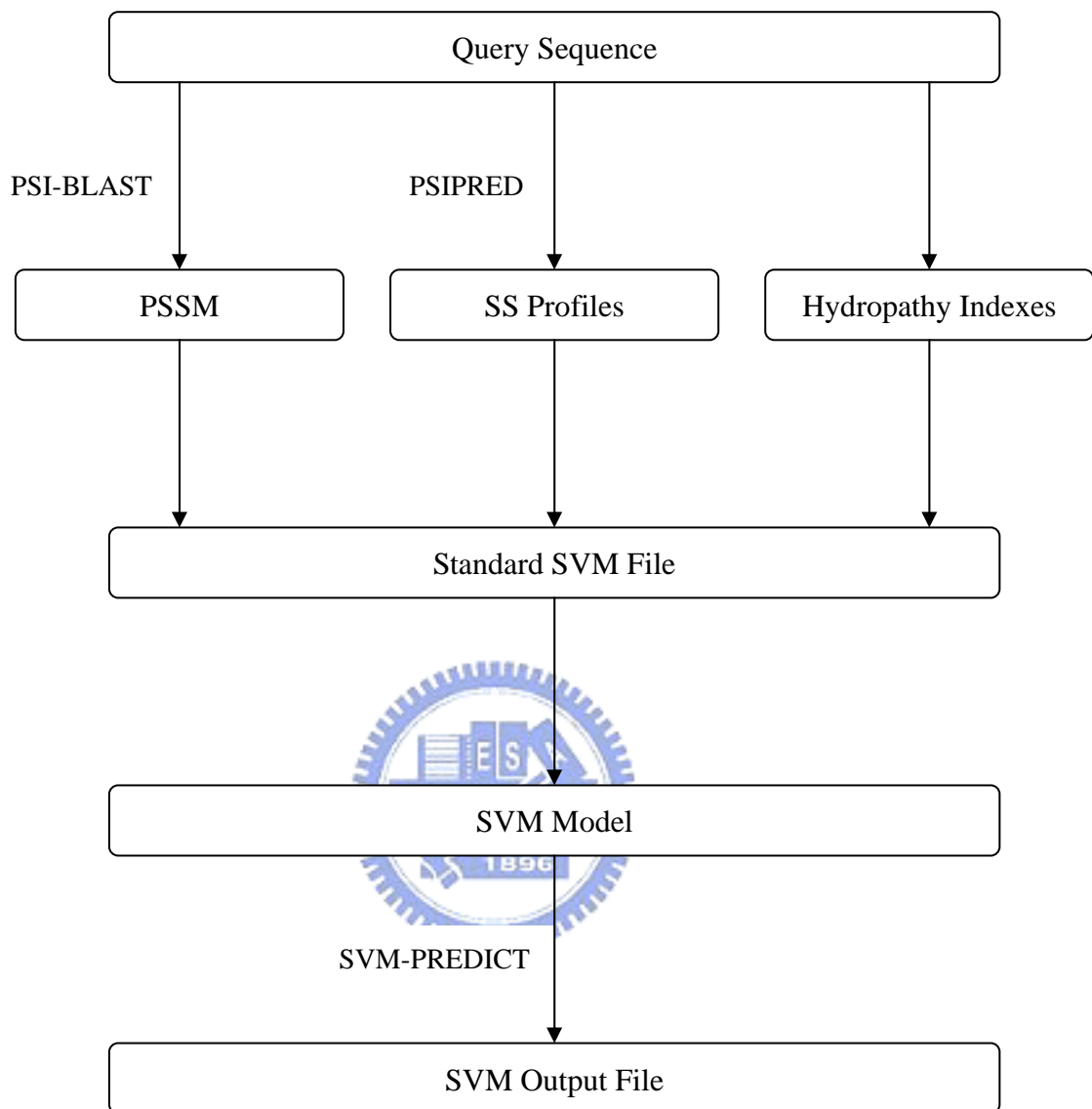


Figure 11. The figure shows the components of SAS prediction server, and most of the connection was built using the Perl language and CGI package.

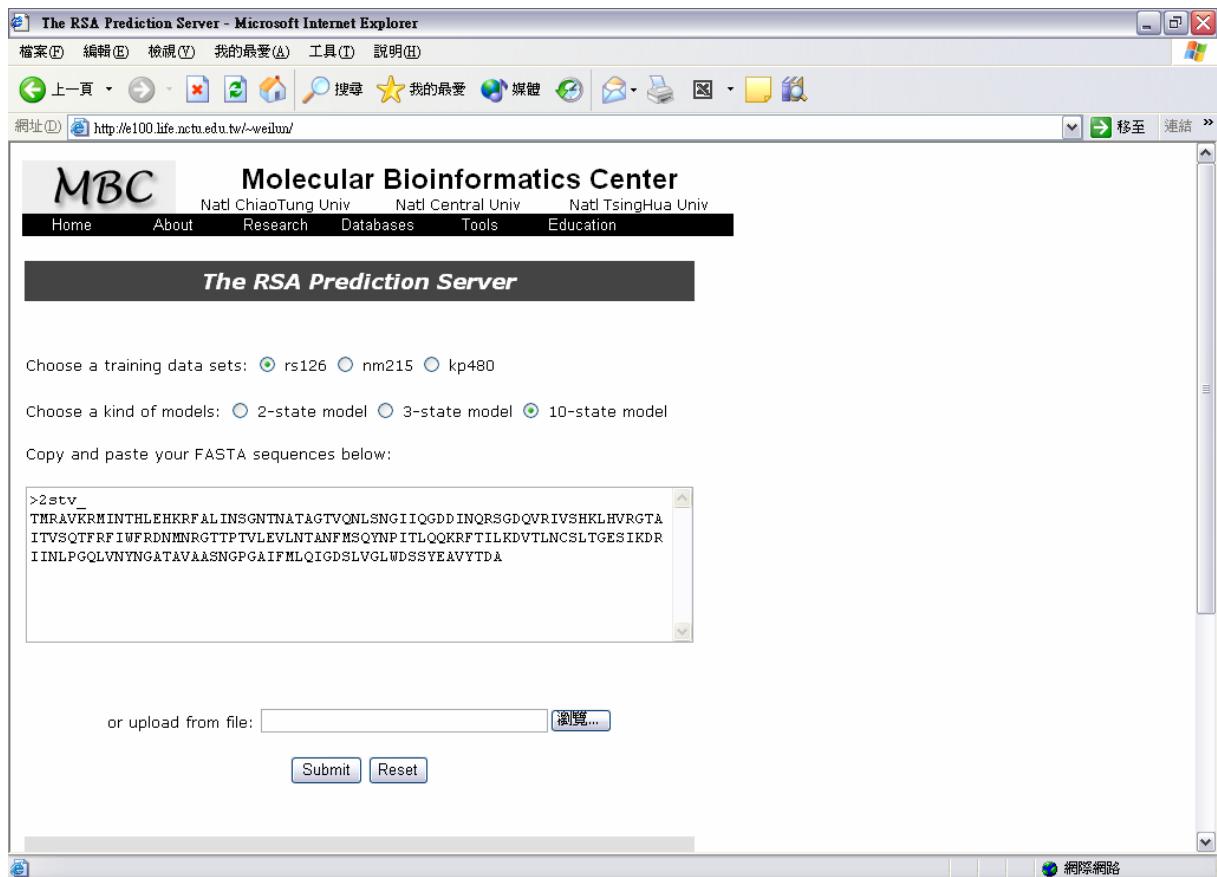
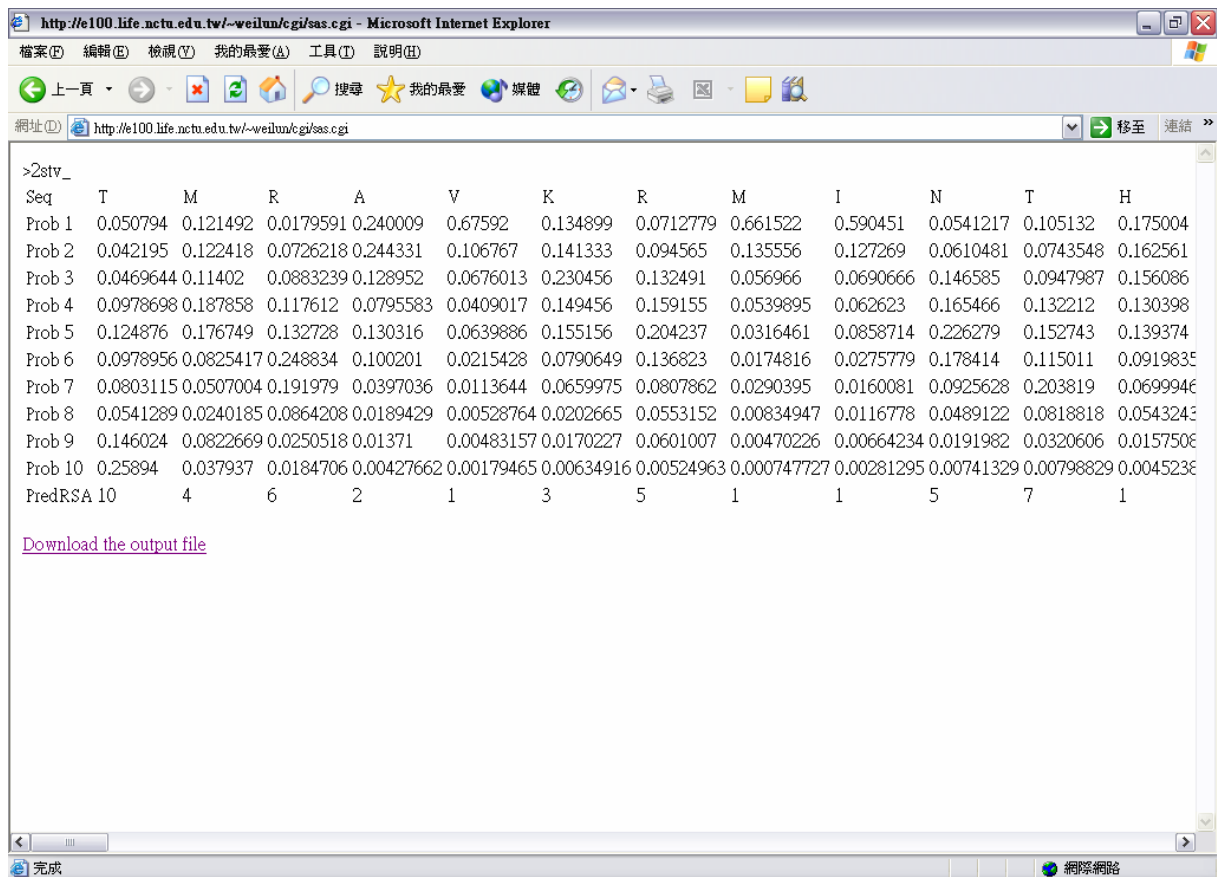


Figure 12. The World Wide Web interface of the RSA prediction server.



*Prob X: probability of the residue belongs to class X

*PredRSA: the predicted relative class

Figure 13. The standard output webpage of prediction.

Tables

Table 1. Maximum value of solvent accessible surface areas of each kind of residue calculated by MOLSIM program

| Amino acid | MaxACC |
|------------|--------|
| A | 117.2 |
| B | 160.0 |
| C | 142.0 |
| D | 169.8 |
| E | 202.0 |
| F | 233.0 |
| G | 87.9 |
| H | 195.4 |
| I | 182.1 |
| K | 214.2 |
| L | 176.2 |
| M | 204.0 |
| N | 169.6 |
| P | 148.9 |
| Q | 199.4 |
| R | 242.9 |
| S | 135.2 |
| T | 152.8 |
| V | 162.4 |
| W | 270.7 |
| X | 180.0 |
| Y | 253.8 |
| Z | 196.0 |



Table 2. The RS126 dataset used in this study

| | | | | |
|--------|--------|--------|--------|--------|
| 256b_a | 8abp_ | 1acx_ | 3ait_ | 2alp_ |
| 9api_b | 1cyo_ | 1bds_ | 1bmv_2 | 4bp2_ |
| 7cat_a | 1cc5_ | 1cdh_ | 3cla_ | 4cms_ |
| 6cpa_ | 4cpv_ | 1cse_i | 2cyp_ | 1eca_ |
| 3ebx_ | 1etu_ | 1fdl_h | 1fkf_ | 1iqz_ |
| 2fox_ | 2gbp_ | 1gdl_o | 2gn5_ | 4gr1_ |
| 6hir_ | 3hmg_b | 5hvp_a | 3icb_ | 1il8_a |
| 1158_ | 5ldh_ | 2lhb_ | 2ltn_a | 5lyz_ |
| 2mev_4 | 1ovo_a | 1paz_ | 2pcy_ | 3pgm_ |
| 1pyp_ | 2mhu_ | 1ppt_ | 1rhd_ | 4rhv_3 |
| 3rnt_ | 2rsp_a | 1s01_ | 4sgb_i | 2sns_ |
| 2stv_ | 1tgs_i | 6tmn_e | 1tnf_a | 2tsc_a |
| 2utg_a | 2wrp_r | 1bks_b | 1prc_c | 1prc_l |
| 2aat_ | 6acn_ | 8adh_ | 2ak3_a | 9api_a |
| 1azu_ | 1bbp_a | 1bmv_1 | 3blm_ | 2cab_ |
| 1cbh_ | 2ccy_a | 1cdt_a | 3cln_ | 4cpa_i |
| 6cpp_ | 1crn_ | 6cts_ | 5cyt_r | 6dfr_ |
| 5er2_e | 1fc2_c | 1dur_ | 1fnd_ | 1fxi_a |
| 1g6n_a | 1a45_ | 2gls_a | 1gp1_a | 1hip_ |
| 3hmg_a | 2hmz_a | 2i1b_ | 7icd_ | 9ins_b |
| 1lap_ | 1gdj_ | 1lmb_3 | 2ltb_b | 1mcp_l |
| 2or1_l | 2pab_a | 9pap_ | 4pfk_ | 2phh_ |
| 1r09_2 | 1mrt_ | 1rbp_ | 4rhv_1 | 4rhv_4 |
| 7rsa_ | 4rxn_ | 3sdh_a | 1sh1_ | 2sod_b |
| 2tgp_i | 3tim_a | 2tmv_p | 4ts1_a | 1ubq_ |
| 9wga_a | 1bks_a | 4xia_a | 1prc_h | 1prc_m |

Table 3. The NM215 dataset used in this study

| | | | | | | |
|-------|-------|-------|-------|-------|-------|-------|
| 119l_ | 2end_ | 1thv_ | 1pex_ | 1hsba | 1csn_ | 1dela |
| 1bnca | 3chy_ | 1xgsa | 1rro_ | 1mbd_ | 1fjma | 1gai_ |
| 1dktb | 1abrb | 2hhma | 1tml_ | 1php_ | 1ido_ | 1itg_ |
| 1ggga | 1cem_ | 3mdda | 1xyza | 1sbp_ | 1mml_ | 1nbab |
| 1knya | 1dxy_ | 1amm_ | 2liv_ | 1ubi_ | 1pmi_ | 1poc_ |
| 1ofga | 1gpc_ | 1cfya | 3sdha | 1ytw_ | 1slua | 1sra_ |
| 1pud_ | 1kuh_ | 1ede_ | 1atla | 2pgd_ | 1uxy_ | 1vhra |
| 1svpa | 1ospo | 1gtma | 1chma | 6gsva | 2arca | 1m85_ |
| 1vls_ | 1ra9_ | 1lki_ | 1erv_ | 1bbpa | 2pia_ | 2rspb |
| 2chsa | 1tfe_ | 1pdo_ | 1hgxa | 1csee | 8atcb | 1bib_ |
| 2sns_ | 1who_ | 1rgs_ | 1mai_ | 1ezm_ | 1bfg_ | 1dfji |
| 153l_ | 2gdm_ | 1thx_ | 1pgs_ | 1htp_ | 1cyx_ | 1gcb_ |
| 1btma | 3cox_ | 1xnb_ | 1rsy_ | 1mkaa | 1ftpa | 1jkw_ |
| 1dkza | 1afra | 2hpda | 1tupc | 1pioa | 1ifc_ | 1nox_ |
| 1gnd_ | 1ceo_ | 3minb | 1yasa | 1sftb | 1mola | 1pot_ |
| 1kpta | 1ecea | 1amp_ | 2mtac | 1uby_ | 1pne_ | 1std_ |
| 1onra | 1gpl_ | 1chd_ | 5p21_ | 256ba | 1smea | 1vid_ |
| 1pyta | 1lba_ | 1edg_ | 1atna | 2phla | 1vcaa | 2cba_ |
| 1tadc | 1pbc_ | 1hava | 1cmke | 6pfka | 2ayh_ | 2scpa |
| 1wba_ | 1rcf_ | 1lkka | 1esc_ | 1bdo_ | 2pspa | 1bmfq |
| 2ctc_ | 1tfr_ | 1pea_ | 1hlb_ | 1csga | 1bgc_ | 1dhr_ |
| 2tysa | 1bksb | 1rnl_ | 1maz_ | 1fds_ | 1deaa | 1gdoa |
| 1aba_ | 2hft_ | 1tib_ | 1phe_ | 1idaa | 1fua_ | 1knb_ |
| 1btn_ | 3grs_ | 1xvaa | 1rvaa | 1mlda | 1irk_ | 1noza |
| 1dosa | 1afwa | 2i1b_ | 1tys_ | 1plc_ | 1nar_ | 1ppn_ |
| 1gotb | 1cewi | 3nll_ | 1ysc_ | 1sig_ | 1poa_ | 1stfi |
| 1kte_ | 1ecpa | 1aoca | 2naca | 1udii | 1smpi | 1vin_ |
| 1opr_ | 1gsa_ | 1chka | 5ptp_ | 2abk_ | 1vhh_ | 2ccya |
| 1qapa | 1lcl_ | 1edt_ | 1axn_ | 2phy_ | 2bbvc | 2sil_ |
| 2tdx_ | 1pda_ | 1hfc_ | 1cnv_ | 7rsa_ | 2rn2_ | |
| 1whi_ | 1rec_ | 1ltsa | 1exnb | 1beo_ | 1bhmb | |

Table 4. The KP480 dataset used in this study

| | | | | |
|----------------|----------------|----------------|----------------|----------------|
| 154l-1-AUTO.1 | 1cbg-1-AS | 1ctn-1-AS.1 | 1edd-1-DOMAK | 2gep-3-AS |
| 1aazb-1-DOMAK | 1cbh | 1ctn-3-AS.1 | 1edmc-1-AUTO.1 | 1gflb-1-AS |
| 1acx | 1cc5 | 1ctu-1-AUTO.1 | 1eft-3-DOMAK | 1ghsb-1-GJB |
| 1add-1-AS | 1cdta | 1ctu-2-AUTO.1 | 1efud-2-AUTO.1 | 1gky-2-AS |
| 1adeb-2-AUTO.1 | 1cei-1-GJB | 1eu1a-4-AUTO.1 | 1epbb-1-DOMAK | 1gln-2-AS |
| 1ahb-2-GJB | 1celb-1-AUTO.1 | 1cyx-1-AUTO.1 | 1ese-1-AUTO.1 | 1gln-4-AS |
| 1alkb-1-AS | 1cem-1-GJB | 1daab-1-AS | 1esl-1-GJB | 1gmpb-1-DOMAK |
| 1amp-1-AS | 1cewi-1-DOMAK | 1daab-2-AS | 1etu | 1gnd-2-JAC |
| 1aorb-1-AS | 1cfb-1-AS | 1dar-3-AS | 1euu-2-JAC | 1gog-1-AS.1 |
| 1aorb-3-AS | 1cgu-2-GJB | 1dfji-1-AUTO.1 | 1fbab-1-DOMAK | 1gog-2-AS.1 |
| 1aozb-1-AS | 1cgu-3-GJB | 1dih-2-AS | 1fbl-1-AS | 1gog-3-AS.1 |
| 1aozb-2-AS | 1cgu-4-GJB | 1dik-1-AS.1 | 1fc2c | 1gp1a |
| 1aozb-3-AS | 2chbe-1-DOMAK | 1dik-2-AS.1 | 1fdlh | 1gp2g-2-AS |
| 1asw-1-AUTO.1 | 1chd-1-AS | 1dik-3-AS.1 | 1fdt-1-AS | 1gpc-1-AS |
| 1avhb-3-AS | 1chkb-2-AUTO.1 | 1dik-4-AS.1 | 1dur | 1gpmd-4-AS |
| 1avhb-4-AS | 1chmb-1-DOMAK | 1din-1-AS | 1find-1-AUTO.1 | 1gpmd-5-AS |
| 1ayab-1-GJB | 1cksc-1-AUTO.1 | 1dkza-1-JAC | 1find-2-AUTO.1 | 1grj-1-AS |
| 1azu | 1clc-1-AS.1 | 1dlc-1-AS.1 | 1fjmb-2-AS | 1grj-2-AS |
| 1bam-1-AS | 1clc-2-AS.1 | 1dlc-3-AS.1 | 1fkf | 1gtmc-2-AUTO.1 |
| 1bbpa | 1clc-3-AS.1 | 1dnpb-1-AUTO.1 | 1fnd | 1gtqb-1-AUTO.1 |
| 1bcx-1-DOMAK | 1cnsb-1-AUTO.1 | 1dnpb-2-AUTO.1 | 1fua-1-AUTO.1 | 1gym-1-AUTO.1 |
| 1bdo-1-AS | 1colb-1-DOMAK | 1dpgb-1-AUTO.1 | 1fuqb-1-AUTO.1 | 1han-1-AUTO.1 |
| 1bet-1-DOMAK | 1comc-1-DOMAK | 1dpgb-2-AUTO.1 | 1fuqb-2-AUTO.1 | 1han-2-AUTO.1 |
| 1bfg-1-DOMAK | 1cpcl-1-DOMAK | 1dsbb-2-AUTO.1 | 1fuqb-3-AUTO.1 | 1hcbg-1-AS |
| 1bmv1 | 1cpn-1-DOMAK | 1dts-1-AUTO.1 | 1fxia | 1hcr-1-DOMAK |
| 1bmv2 | 1cqa-1-AUTO.1 | 1dupa-1-AS | 1gal-2-AS | 1hip |
| 1bncb-1-AS | 1crn | 1dynb-1-AUTO.1 | 1gal-3-AS | 1hiws-1-AS |
| 1bncb-3-AS | 1csei | 1eca | 1gcb-2-AS | 1hjrd-1-AUTO.1 |
| 1bncb-4-AS | 1csmb-1-AUTO.1 | 1eceb-1-AUTO.1 | 1gcmc-1-AUTO.1 | 1hmpp-1-AUTO.1 |
| 1bovb-1-DOMAK | 1ctf-1-DOMAK | 1ecl-1-AS | 1gd1o | 1hnf-1-AS |
| 1brse-1-DOMAK | 2cthb-1-DOMAK | 1ecl-4-AS | 1gdj | 1hnf-2-AS |
| 1bsdb-1-DOMAK | 1ctm-2-DOMAK | 1ecpf-1-AUTO.1 | 2gep-2-AS | 1horb-1-AUTO.1 |

Table 4. The KP480 dataset used in this study (continued)

| | | | | |
|----------------|----------------|----------------|----------------|----------------|
| 1hplb-1-AS | 1leh3-3-AS | 1noz2-2-AUTO.1 | 1ptr-1-AUTO.1 | 1sfe-1-AS |
| 1hplb-2-AS | 1lib-1-DOMAK | 1oacb-1-AS.1 | 1ptx-1-AS | 1sfe-2-AS |
| 1hslb-2-DOMAK | 1lis-1-DOMAK | 1oacb-2-AS.1 | 1pyp | 1sftb-2-AS |
| 1htrp-1-AS | 1lki-1-AS | 1oacb-3-AS.1 | 1pyta-1-AS | 1sh1 |
| 1hvq-1-AUTO.1 | 1lmb3 | 1oacb-4-AS.1 | 1qbb-1-AUTO.1 | 1smnb-1-AUTO.1 |
| 1hxn-1-AS | 1lpba-1-DOMAK | 1onrb-1-AUTO.1 | 1qbb-2-AUTO.1 | 1smpi-1-AS |
| 1hyp-1-DOMAK | 1lpe-1-DOMAK | 1otgc-1-AS | 1qbb-3-AUTO.1 | 1spbp-1-AS |
| 1ignb-2-GJB | 1mai-1-JAC | 1ovb-1-GJB | 1qbb-4-AUTO.1 | 1sra-1-AS |
| 1il8a | 1masb-1-AUTO.1 | 1ovoa | 1qrdb-1-AUTO.1 | 1srja-1-DOMAK |
| 1ilk-1-AS | 1mdam-1-DOMAK | 1oxy-3-AS | 1r092 | 1stfi-1-DOMAK |
| 1ilk-2-AS | 1mdta-1-AS | 1oyc-1-AS | 1rbp | 1stme-1-AUTO.1 |
| 1inp-1-AS.1 | 1mdta-2-AS | 1paz | 1rec-1-DOMAK | 1svb-1-AS |
| 1inp-2-AS.1 | 1mdta-3-AS | 1pbp-2-DOMAK | 1rec-2-DOMAK | 1svb-2-AS |
| 1irk-1-AS | 1mjc-1-DOMAK | 1pbwb-1-AS | 1regy-1-AUTO.1 | 1tabi-1-DOMAK |
| 1irk-2-AS | 1mla-2-AS.1 | 1pda-2-AS | 1reqc-2-AS | 1taq-2-AS |
| 1isab-1-GJB | 1mmoh-1-AS | 1pda-3-AS | 1rhd | 1tcra-2-GJB |
| 1isab-2-GJB | 1mns-2-AS | 1pdnc-2-AS | 1rhgc-1-DOMAK | 1tfr-1-GJB |
| 1isub-1-DOMAK | 1mof-1-AS | 1pdo-1-GJB | 1rie-1-GJB | 1thtb-1-AUTO.1 |
| 1jud-1-GJB | 1mrrb-1-DOMAK | 1pga-1-DOMAK | 1ris-1-DOMAK | 1thx-1-AUTO.1 |
| 1kimb-1-AUTO.1 | 1mrt | 1pht-1-AUTO.1 | 1rls-1-DOMAK | 1tie-1-DOMAK |
| 1knb-1-AS | 1mspb-1-AS | 1pii-2-DOMAK | 1rlr-1-JAC | 1tif-1-AS |
| 1krca-1-AUTO.1 | 1nal4-1-AUTO.1 | 1pkyc-2-AUTO.1 | 1rlr-2-JAC | 1tig-1-AUTO.1 |
| 1krcb-1-AS | 1nar-1-DOMAK | 1pkyc-3-AUTO.1 | 1rpo-1-AUTO.1 | 1tml-1-AS |
| 1kte-1-AS | 1nbac-1-AS | 1pmi-2-GJB | 1rsy-1-AS | 1tndb-2-DOMAK |
| 1ktq-1-AUTO.1 | 1ncg-1-AUTO.2 | 1pnmb-2-AS | 1rvvz-1-AUTO.1 | 1tnfa |
| 1kuh-1-AS | 1ndh-1-AS | 1pnt-1-AS | 1s01 | 1tplb-3-AS |
| 1l58 | 1ndh-2-AS | 1poc-1-DOMAK | 1scud-1-AS | 1trb-2-AS |
| 1lap | 1nfp-1-AS | 1powb-1-DOMAK | 1scue-2-AS | 1trh-1-AS |
| 1latb-1-AUTO.1 | 1nga-2-AS.1 | 1powb-2-DOMAK | 1scue-3-AS | 1trkb-1-AS |
| 1lba-1-DOMAK | 1nkl-1-DOMAK | 1powb-3-DOMAK | 1seib-1-AUTO.1 | 1trkb-3-AS |
| 1lbu-1-AS | 1nol-1-AUTO.2 | 1ppi-2-AS | 1seib-2-AUTO.1 | 1tsp-1-AS |
| 1lbu-2-AS | 1nox-1-GJB | 1ppt | 1sesa-2-AS | 2tssb-2-DOMAK |

Table 4. The KP480 dataset used in this study (continued)

| | | | | |
|----------------|----------------|---------------|---------------|---------------|
| 1tul-1-JAC | 2afnc-2-AUTO.1 | 2i1b | 2utga | 4fisb-1-DOMAK |
| 1tupc-1-AUTO.1 | 2ak3a | 2ltna | 2wrpr | 4gr1 |
| 1ubq | 2alp | 2ltnb | 2yhx-3-DOMAK | 4pfk |
| 1udh-1-AUTO.1 | 2asr-1-DOMAK | 2mev4 | 3ait | 4rhv1 |
| 1umub-1-AS | 2bat-1-GJB | 2mtac-1-AS | 1cyo | 4rhv3 |
| 1vcab-1-AUTO.1 | 2bltb-2-AUTO.1 | 2nadb-2-AS.1 | 4bcl-1-DOMAK | 4rhv4 |
| 1vcab-2-AUTO.1 | 2bopa-1-DOMAK | 2npx-3-AS.1 | 3blm | 4rxn |
| 1vcc-1-AS | 2cab | 2olba-2-AS | 3cd4 | 4sdha |
| 1vhh-1-AS | 2ccya | 2olba-3-AS | 3chy-1-DOMAK | 4sgbi |
| 1vhrb-2-AUTO.1 | 2cmd-2-GJB | 2or11 | 3cla | 4ts1a |
| 1vid-1-JAC | 2cpo-1-AUTO.1 | 2paba | 3cln | 4xiaa |
| 1vjs-3-GJB | 2cyp | 2pgd-1-AUTO.1 | 3cox-1-AS.1 | 5cytr |
| 1vmob-1-AS | 2dkb-2-AS | 2pgd-2-AUTO.1 | 3cox-2-AS.1 | 5er2e |
| 1vnc-1-JAC | 2dlm-1-AS | 2phh | 3ecab-1-AS | 5ldh |
| 1vokb-1-AS | 2dlm-3-AS | 2phy-1-GJB | 3ecab-2-AS | 5lyz |
| 1vpt-1-JAC | 2dnja-1-AS | 2polb-1-AS | 1g6na | 5sici-1-DOMAK |
| 1wapv-1-AUTO.1 | 2ebn-1-AS | 2reb-1-DOMAK | 3hmga | 6acn |
| 1whi-1-AS | 2end-1-DOMAK | 2reb-2-DOMAK | 3hmgb | 6cpa |
| 1bksa | 2fox | 2rsla-1-GJB | 3icb | 6cpp |
| 1bkzb | 1iqz | 2rspa | 3inkd-1-DOMAK | 6cts |
| 1xvab-1-GJB | 2gbp | 2scpb-1-DOMAK | 3mddb-1-AS | 6dfr |
| 1yptb-1-AUTO.1 | 1a45 | 2sil-1-AS | 3mddb-2-AS | 6hir |
| 1yrna-2-AS | 2glsa | 2sns | 3mddb-3-AS | 6tmne |
| 1znbb-1-AS | 2gn5 | 2sodb | 3pgk-2-AS | 7cata |
| 1zymb-2-AUTO.1 | 2gsq-2-AS | 2spt-1-DOMAK | 3pgm | 7icd |
| 256ba | 2hft-1-AS | 2spt-2-DOMAK | 3pmgb-1-AS | 7rsa |
| 2aaib-2-DOMAK | 2hft-2-AS | 2tgi-1-DOMAK | 3pmgb-2-AS | 821p-1-DOMAK |
| 2aat | 2hhmb-1-DOMAK | 2tgpi | 3pmgb-3-AS | 8adh |
| 2abk-2-AS | 2hhmb-2-DOMAK | 2tmdb-3-AS | 3pmgb-4-AS | 9apia |
| 2admb-1-AUTO.1 | 2hipb-1-DOMAK | 2tmvp | 3rnt | 9apib |
| 2admb-2-AUTO.1 | 2hmza | 2trt-1-AUTO.1 | 3tima | 9pap |
| 2afnc-1-AUTO.1 | 2hpr-1-DOMAK | 2tsca | 4bp2 | 9wgaa |

Table 5. The hydropathy index of the standard amino acids.

| Amino acid | Hydropathy index |
|------------|------------------|
| A | 1.8 |
| C | 2.5 |
| D | -3.5 |
| E | -3.5 |
| F | 2.8 |
| G | -0.4 |
| H | -3.2 |
| I | 4.5 |
| K | -3.9 |
| L | 3.8 |
| M | 1.9 |
| N | -3.5 |
| P | 1.6 |
| Q | -3.5 |
| R | -4.5 |
| S | -0.8 |
| T | -0.7 |
| V | 4.2 |
| W | -0.9 |
| Y | -1.3 |



Table 6. The classification of the standard amino acids by R groups.

| Class | Amino acid |
|------------------------------|---------------------------------|
| Nonpolar, aliphatic R groups | G A P V L I M |
| Aromatic R groups | F Y W |
| Polar, uncharged R groups | S T C N Q |
| Positively charged R groups | K H R |
| Negatively charged R groups | D E |



Table 7. Prediction accuracies and MCC values using individual local descriptors with RS126 datasets in 2-state prediction model.

| Features | Threshold | Qtotal | Qb | Qe | MCC |
|---|-----------|--------|-------|-------|-------|
| PSSM | 25 | 75.33 | 76.38 | 74.11 | 50.43 |
| Hydropathy indexes | 25 | 65.73 | 80.43 | 48.61 | 30.79 |
| Secondary structure profile | 25 | 67.80 | 68.93 | 66.46 | 35.34 |
| Amino acid in binary coding | 25 | 70.59 | 74.49 | 66.06 | 40.70 |
| Amino acid properties in binary coding | 25 | 67.34 | 69.51 | 64.81 | 34.31 |
| Local amino acid composition | 25 | 58.22 | 72.67 | 41.39 | 14.82 |
| Local amino acid properties composition | 25 | 58.04 | 75.85 | 37.30 | 14.27 |
| Residue size | 25 | 55.8 | 96.19 | 8.68 | 10.18 |

Table 8. Prediction accuracies and MCC values using comprehensive local descriptors with RS126 datasets in 2-state prediction model.

| Features | Threshold | Qtotal | Qb | Qe | MCC |
|-------------------|-----------|--------|-------|-------|-------|
| PSSM+H | 25 | 76.45 | 78.96 | 73.52 | 52.57 |
| PSSM+SS | 25 | 75.98 | 79.06 | 72.40 | 51.60 |
| H+SS | 25 | 68.40 | 70.05 | 66.48 | 36.50 |
| PSSM+H+SS | 25 | 77.23 | 78.53 | 75.72 | 54.23 |
| PSSM+H+SS+AA | 25 | 77.78 | 79.44 | 75.84 | 55.29 |
| PSSM+H+SS+PROP | 25 | 77.40 | 79.15 | 75.36 | 54.53 |
| PSSM+H+SS+AA+PROP | 25 | 77.33 | 78.64 | 75.80 | 54.42 |
| PSSM+H+SS+GAA | 25 | 77.28 | 78.48 | 75.88 | 54.33 |
| PSSM+H+SS+GPROP | 25 | 77.15 | 78.29 | 75.82 | 54.07 |

*H: hydropathy indexes

*SS: secondary structure profile

*AA: amino acid in binary coding

*PROP: amino acid properties in binary coding

*GAA: Global amino acid composition

*GPROP: Global amino acid properties composition

Table 9. Prediction accuracies of the benchmark tests on the RS126, NM215 and KP480 datasets in 2-state prediction model and its comparison with other methods

| Year | Method | State threshold | | | | | |
|------|------------------------------|-----------------|------|------|------|------|-------|
| | | 25% | 20% | 16% | 5% | 0% | 9,36% |
| 1994 | PHDacc(RS126) ^[3] | - | - | 74.8 | - | 86.0 | 57.5 |
| 2000 | Jnet(CB480)[7] | 76.2 | - | - | 79.8 | 86.5 | - |
| 2002 | BRNNs (PDB-1086)[4] | 77.2 | 77.5 | - | 81.2 | 86.5 | - |
| 2003 | PP(RS-126)[23] | - | - | 75.1 | - | - | 57.9 |
| 2003 | NETASA(NM-215)[24] | 70.3 | - | - | 74.6 | 87.9 | - |
| 2004 | SVMpsi (RS126)[13] | 76.8 | - | 77.8 | 79.8 | 86.2 | 59.6 |
| 2004 | SVMpsi(KP480)[13] | 78.7 | - | 80.7 | 82.4 | 87.4 | 64.5 |
| 2005 | Fuzzy k-NN(RS-126)[14] | 78.3 | - | 79.0 | 82.2 | 87.2 | 63.8 |
| 2005 | SVR12(RA-603)[10] | 76.7 | 76.8 | - | 76.5 | - | - |
| 2005 | WE(2148 chains)[1] | - | 80.0 | - | - | - | - |
| | PSSM+H+SS (RS126) | 77.2 | 77.6 | 77.9 | 81.4 | 88.4 | 60.2 |
| | PSSM+H+SS+AA (RS126) | 77.8 | 77.8 | 78.1 | 80.0 | 88.8 | 61.6 |
| | PSSM+H+SS (NM215) | 78.3 | 77.7 | 87.2 | 87.4 | 87.0 | 62.7 |
| | PSSM+H+SS (KP480) | 78.0 | | 78.5 | 81.3 | 87.0 | 62.3 |

Table 10. MCC values of the benchmark tests on the RS126, NM215 and KP480 datasets in 2-state prediction model and its comparison with other methods

| Year | Method | State threshold | | | | | |
|------|------------------------------|-----------------|------|------|------|------|----------------|
| | | 25% | 20% | 16% | 5% | 0% | 9,36% |
| 1994 | PHDacc(RS126) ^[3] | - | - | - | - | - | - |
| 2000 | Jnet(CB480)[7] | - | - | - | - | - | - |
| 2002 | BRNNs (PDB-1086)[4] | - | - | - | - | - | - |
| 2003 | PP(RS-126)[23] | - | - | 48.5 | - | - | 53.0 |
| 2003 | NETASA(NM-215)[24] | - | - | - | - | - | - |
| 2004 | SVMpsi (RS126)[13] | - | - | - | - | - | - |
| 2004 | SVMpsi(KP480)[13] | - | - | - | - | - | - |
| 2005 | Fuzzy k-NN(SCOP-229)[14] | 55.4 | - | 56.0 | 54.1 | 43.1 | 56.0;19.9;50.8 |
| 2005 | SVR12(RA-603)[10] | - | - | - | - | - | - |
| 2005 | WE(2148 chains)[1] | - | 60.0 | - | - | - | - |
| | PSSM+H+SS (RS126) | 54.2 | 55.2 | 54.7 | 49.7 | 22.2 | 52.0;17.0;48.4 |
| | PSSM+H+SS+AA (RS126) | 55.3 | 55.6 | 55.2 | 45.7 | 31.7 | 53.2;19.3;50.8 |
| | PSSM+H+SS (NM215) | 56.0 | 56.5 | 34.2 | 36.4 | 38.3 | 56.3;20.9;50.7 |
| | PSSM+H+SS (KP480) | 55.1 | | 56.6 | 53.2 | 35.8 | 55.2;19.9;50.9 |

Table 11. MAE and Pearson’s “r” values of the benchmark tests on the RS126 and KP480 datasets in 10-state prediction model

| Year | Method | Data Set | MAE | R | Accuracy |
|------|---------------------|------------|-----------|-----------|------------|
| 2003 | RVP ¹⁶ | RS-126 | 19.4 | 0.477 | 71%(16%) |
| | | Carugo-338 | 19.0 | 0.490 | |
| | | CB-502 | 18.8 | 0.482 | |
| 2004 | SVR[25] | CB-502 | 18.5 | 0.520 | |
| | | YH-1277 | 17.0 | 0.617 | 74.6%(15%) |
| | SABLE[5] | RA-603 | 15.3-15.8 | 0.64-0.67 | 77%(15%) |
| 2005 | SVR12 ¹⁸ | RA-603 | 15.8-16.6 | 0.62-0.63 | 75.8(15%) |
| | PSSM+H+SS | RS-126 | 15.2 | 0.506 | 72.8(20%) |
| | | KP-480 | 14.1 | 0.537 | 74.4(20%) |

參考文獻

1. Chen, H. and H.X. Zhou, *Prediction of solvent accessibility and sites of deleterious mutations from protein sequence*. Nucleic Acids Res, 2005. **33**(10): p. 3193-9.
2. Wang, Z. and J. Moulton, *SNPs, protein structure, and disease*. Hum Mutat, 2001. **17**(4): p. 263-70.
3. Rost, B. and C. Sander, *Conservation and prediction of solvent accessibility in protein families*. Proteins, 1994. **20**(3): p. 216-26.
4. Pollastri, G., et al., *Prediction of coordination number and relative solvent accessibility in proteins*. Proteins, 2002. **47**(2): p. 142-53.
5. Adamczak, R., A. Porollo, and J. Meller, *Accurate prediction of solvent accessibility using neural networks-based regression*. Proteins, 2004. **56**(4): p. 753-67.
6. Ahmad, S. and M.M. Gromiha, *Design and training of a neural network for predicting the solvent accessibility of proteins*. J Comput Chem, 2003. **24**(11): p. 1313-20.
7. Cuff, J.A. and G.J. Barton, *Application of multiple sequence alignment profiles to improve protein secondary structure prediction*. Proteins, 2000. **40**(3): p. 502-11.
8. Thompson, M.J. and R.A. Goldstein, *Predicting solvent accessibility: higher accuracy using Bayesian statistics and optimized residue substitution classes*. Proteins, 1996. **25**(1): p. 38-47.
9. Li, X. and X.M. Pan, *New method for accurate prediction of solvent accessibility from protein sequence*. Proteins, 2001. **42**(1): p. 1-5.
10. Wagner, M., et al., *Linear regression models for solvent accessibility prediction in proteins*. J Comput Biol, 2005. **12**(3): p. 355-69.
11. Naderi-Manesh, H., et al., *Prediction of protein surface accessibility with information theory*. Proteins, 2001. **42**(4): p. 452-9.
12. Yuan, Z., K. Burrage, and J.S. Mattick, *Prediction of protein solvent accessibility using support vector machines*. Proteins, 2002. **48**(3): p. 566-70.
13. Kim, H. and H. Park, *Prediction of protein relative solvent accessibility with support vector machines and long-range interaction 3D local descriptor*. Proteins, 2004. **54**(3): p. 557-62.
14. Sim, J., S.Y. Kim, and J. Lee, *Prediction of protein solvent accessibility using fuzzy k-nearest neighbor method*. Bioinformatics, 2005. **21**(12): p. 2844-9.
15. Kabsch, W. and C. Sander, *Dictionary of protein secondary structure: pattern recognition of hydrogen-bonded and geometrical features*. Biopolymers, 1983. **22**(12): p. 2577-637.
16. Altschul, S.F., et al., *Gapped BLAST and PSI-BLAST: a new generation of protein database search programs*. Nucleic Acids Res, 1997. **25**(17): p. 3389-402.
17. Jones, D.T., *Protein secondary structure prediction based on position-specific scoring*

- matrices*. J Mol Biol, 1999. **292**(2): p. 195-202.
18. Boser, B.E., Guyon, I.M. and Vapnik, V.N., *A training algorithm for optimal margin classifiers*. ACM Press, 1992: p. 144-152.
 19. Hua, S. and Z. Sun, *A novel method of protein secondary structure prediction with high segment overlap measure: support vector machine approach*. J Mol Biol, 2001. **308**(2): p. 397-407.
 20. Ding, C.H. and I. Dubchak, *Multi-class protein fold recognition using support vector machines and neural networks*. Bioinformatics, 2001. **17**(4): p. 349-58.
 21. Furey, T.S., et al., *Support vector machine classification and validation of cancer tissue samples using microarray expression data*. Bioinformatics, 2000. **16**(10): p. 906-14.
 22. C.C. Chang, C.J.L., *Training nu-Support Vector Classifiers: Theory and Algorithms*. Neural Computation, 2001. **13**(9): p. 2119-2147.
 23. Gianese, G., F. Bossa, and S. Pascarella, *Improvement in prediction of solvent accessibility by probability profiles*. Protein Eng, 2003. **16**(12): p. 987-92.
 24. Ahmad, S., M.M. Gromiha, and A. Sarai, *Real value prediction of solvent accessibility from amino acid sequence*. Proteins, 2003. **50**(4): p. 629-35.
 25. Yuan, Z. and B. Huang, *Prediction of protein accessible surface areas by support vector regression*. Proteins, 2004. **57**(3): p. 558-64.

