

Mycology

Susceptibilities to amphotericin B and fluconazole of *Candida* species in Taiwan Surveillance of Antimicrobial Resistance of Yeasts 2006

Yun-Liang Yang^a, An-Huei Wang^b, Chih-Wei Wang^b, Wei-Ting Cheng^b,
Shu-Ying Li^c, Hsiu-Jung Lo^{b,*}
TSARY Hospitals

^aDepartment of Biological Science and Technology, National Chiao Tung University, Hsinchu 300, Taiwan, ROC

^bDivision of Clinical Research, National Health Research Institutes, Zhunan Town, Miaoli County 350, Taiwan, ROC

^cDivision of Laboratory Research and Development, Center for Disease Control, Taipei 115, Taiwan, ROC

Received 27 October 2007; accepted 16 January 2008

Abstract

Susceptibilities to amphotericin B and fluconazole of 964 *Candida* isolates collected in Taiwan Surveillance of Antimicrobial Resistance of Yeasts in 2006 were determined. There were 419 (43.5%) *Candida albicans*, 246 (25.5%) *Candida tropicalis*, 211 (21.9%) *Candida glabrata*, 62 (6.4%) *Candida parapsilosis*, 14 (1.5%) *Candida krusei*, and 12 (1.2%) others. Interestingly, 16 of the 17 amphotericin B-resistant isolates were non-*albicans* *Candida* species. The resistant rate to amphotericin B has decreased from 2.5% in 2002 to 1.8% in 2006. On the other hand, there were 132 *C. tropicalis*, 14 *C. krusei*, 10 *C. albicans*, and 9 *C. glabrata* isolates that had MICs to fluconazole ≥ 64 $\mu\text{g/mL}$. The prevalence of isolates with such high MICs was significantly higher than that in 2002 (17.1% versus 1.9%). Our results further indicate that most of the isolates with MICs to fluconazole ≥ 64 $\mu\text{g/mL}$ exhibited the “trailing” phenomenon.

© 2008 Elsevier Inc. All rights reserved.

Keywords: *Candida*; Susceptibility; Resistance

1. Introduction

Because of alterations in immune status and invasive hospital procedures (White et al., 1998; Yang and Lo, 2001), infections caused by opportunistic pathogens, such as yeasts, are becoming important causes of morbidity and mortality in immunocompromised patients. In the past 2 decades, nosocomial yeast infections have increased significantly worldwide. For example, the prevalence of nosocomial candidemia increased 27-fold from 1981 through 1993 at a major hospital in Taiwan (Chen et al., 1997; Hung et al., 1996). In the United States, yeast infection also ranks as the 4th most common cause of nosocomial bloodstream infection (Wisplinghoff et al., 2004). Several antifungal drugs have been applied to render the situation, and as a

result of broad prophylactic usages and long-term treatments with those drugs, the prevalence of drug resistance has become an important issue in various yeast infections, which have profound effects on human health (Marr et al., 2001; Pfaller et al., 2003; Yang et al., 2004b).

Candida species have various degrees of susceptibility to frequently used antifungal drugs. For instance, *Candida lusitanae* is relatively resistant to amphotericin B (Hadfield et al., 1987). *Candida krusei* is intrinsically resistant to fluconazole, and *Candida glabrata* is less susceptible or has a higher MICs to it than other *Candida* species (Akova et al., 1991; Orozco et al., 1998; Yang et al., 2004b). This phenomenon illustrates the importance of identification and surveillance of *Candida* species in the clinical settings.

In 1999 and again in 2002, 2 national surveys in Taiwan Surveillance of Antimicrobial Resistance of Yeasts (TSARY) have been conducted. The drug susceptibilities of the 632 and 909 isolates collected in 1999 and 2002, respectively, have been determined (Yang et al., 2004b, 2005b). Among them, 0.5% (1999) and 2.5% (2002) were resistant to

* Corresponding author. Tel.: +886-37-246-166x35516; fax: +886-37-586-457.

E-mail address: hjlo@nhri.org.tw (H.-J. Lo).

amphotericin B. There were 8.4% of the isolates in 1999 that had MICs to fluconazole ≥ 64 $\mu\text{g/mL}$, whereas only 1.9% in 2002 (Yang et al., 2004b, 2005b). In 2006, a follow-up survey was taken place, and 964 *Candida* isolates were collected and analyzed. The article is to report the result of the susceptibilities to antifungal drugs of *Candida* species in TSARY 2006 and the trends of the resistance in Taiwan from 1999 to 2006.

2. Materials and methods

2.1. Organisms and media

Yeast isolates were collected according to previous studies (Lo et al., 2001; Yang et al., 2005b) from the 22 hospitals participating in TSARY from July to September in 2006. Each hospital was asked to submit all yeast pathogens from blood and the first 10 *Candida albicans* and 40 non-*albicans Candida* species isolates from nonsterile sites. In principle, only 1 isolate was accepted from each specimen. Nevertheless, when there were multiple species isolated from 1 specimen, 1 isolate from each species was analyzed. All the collected isolates were stored frozen at -70 °C in bead-containing Microbank cryovials (Pro-Lab Diagnostics, Austin, TX). After their arrival at the laboratory at National Health Research Institutes (NHRI), Taiwan, ROC, these isolates were 1st subcultured on Sabouraud dextrose agar (Becton Dickinson, Cockeysville, MD) to assess the purity and identification. Pure isolates were labeled and stored in vials containing 50% glycerol at -70 °C awaiting further analyses.

2.2. Identification

The identifications of the isolates were reassessed in the laboratory at the NHRI. The identification procedure for the yeast isolates was modified based on our previous report (Lo et al., 2001). All isolates were subjected to API-32C (bioMérieux, St. Louis, MO). The VITEK Yeast Biochemical Card (YBC, bioMérieux) would then be used for the identification of the isolates when the API-32C showed less than 90% confidence and for the isolates whose information was inconsistent to that provided by the hospitals. The sequence of internal transcribed spaces of ribosomal DNA (Leaw et al., 2006, 2007) was applied for further assessment when both API-32C and YBC failed.

2.3. Antifungal susceptibility testing

The MICs to amphotericin B and to fluconazole of each isolate were determined by the same in vitro antifungal susceptibility testing as those in TSARY 1999 and TSARY 2002 (Yang et al., 2004b, 2005b), according to the guidelines of M27A published in 1997 by the Clinical and Laboratory Standards Institute (CLSI, 1997). The RPMI medium 1640 (31800-022) provided by Gibco BRL was used for the testing. Strains from American Type Culture Collection

including *C. albicans* (ATCC 90028), *C. krusei* (ATCC 6258), and *Candida parapsilosis* (ATCC 22019) were used as the standard controls. The final growth of each isolate was measured by Biotrak II plate reader (Amersham Biosciences, Cambridge, UK) after incubation at 35 °C for 48 h. The MICs of some isolates were also measured by Etest (AB BIODISK, Solna, Sweden) to assess the results of the broth microdilution method.

The MICs to amphotericin B and to fluconazole were defined as the MICs of drugs capable of reducing the turbidity of cells to greater than 95% and 50%, respectively. For susceptibility to amphotericin B, isolates with MIC ≥ 2 $\mu\text{g/mL}$ were considered to be resistant, and those with MIC ≤ 1 $\mu\text{g/mL}$ were susceptible. For susceptibility to fluconazole, isolates with MIC ≥ 64 $\mu\text{g/mL}$ were considered to be resistant, whereas those with MIC ≤ 8 $\mu\text{g/mL}$ were susceptible. Isolates with MICs falling in between (16–32 $\mu\text{g/mL}$) were susceptible dose dependent. The MICs of 50% and 90% of the total population were defined as MIC₅₀ and MIC₉₀, respectively.

Among the phenomena associated with resistance, “trailing” describes the reduced but persistent growth that some isolates exhibit at drug concentrations above the MIC in broth dilution tests with azole antifungal agents, such as fluconazole (Lee et al., 2004). When the MIC of an isolate measured after 48-h incubation is approximately 4-fold higher than that at the 24-h point (Arthington-Skaggs et al., 2002), the isolate is defined to have trailing growth.

2.4. Database and analysis

The database for this study contained the characteristic information of each submitted isolate: hospital origin, location and type of the hospital, and identification and source of the isolate. The statistic significance of the differences in frequencies and proportions was determined by the χ^2 test with Yates' correction.

3. Results and discussion

3.1. Distribution of *Candida* species

The distribution of *Candida* species in TSARY 2006 (Table 1) was similar to that of 2 previous surveys in 1999 and 2002 (Yang et al., 2004b, 2005a). *C. albicans* was the most frequently isolated species, accounting for 43.5% of the total isolates. *Candida tropicalis* (25.5%) and *C. glabrata* (21.9%) were the 2 most frequently isolated non-*albicans Candida* species, followed by *C. parapsilosis* (6.4%), *C. krusei* (1.5%), and others (1.2%). When classified according to the sources (Table 1), there were 406 (42.1%) isolates from urine, 158 (16.4%) from sputum, 145 (15%) from blood, 43 (4.5%) from wound, 40 (4.1%) from pus, 40 (4.1%) from catheter tip, 38 (4%) from ascites, and 94 (9.8%) from other 45 different sites.

Different *Candida* species had various prevalence in different body sites. Although *C. albicans* was still the most

Table 1
The distribution of *Candida* species from different sources

Sources	<i>C. albicans</i>	<i>C. tropicalis</i>	<i>C. glabrata</i>	<i>C. parapsilosis</i>	<i>C. krusei</i>	Others	Total
Urine	129	109	145	13	5	5	406 (42.1) ^a
Sputum	70	65	19	2	2	0	158 (16.4)
Blood	82	32	20	9	1	1	145 (15.1)
Wound	21	6	6	5	1	4	43 (4.5)
Pus	20	9	2	6	2	1	40 (4.1)
Tip ^b	20	13	3	4	0	0	40 (4.1)
Ascites ^c	24	7	4	2	1	0	38 (3.9)
Others	53	5	12	21	2	1	94 (9.8)
Total	419 (43.5)	246 (25.5)	211 (21.9)	62 (6.4)	14 (1.5)	12 (1.2)	964

^a Number of isolates (percentage).

^b Catheter tips.

^c Body fluid, not urinary ascites or pus.

common single species (56.6%) causing candidemia in our study as well as others' (Cheng et al., 2004; Peman et al., 2005; Pfaller et al., 2007a; Yang et al., 2006), the prevalence of non-albicans *Candida* species had increased. Though *C. tropicalis* was the most common non-albicans *Candida* species in our collection, *C. glabrata* was the most common one from urine (145/277, 52.3%). The majority of *C. glabrata* (68.7%) was isolated from urine samples, consistent with the previous reports that *C. glabrata* is 2nd only to *C. albicans* as a cause of candiduria (Pfaller et al., 1999; Yang et al., 2004b).

3.2. Susceptibilities to amphotericin B of *Candida* species

The range of MICs to amphotericin B of the 964 isolates was from 0.125 to 2 µg/mL (Table 2). *C. krusei* was less susceptible to amphotericin B than any other species because the MIC₅₀ of this species was 1 µg/mL. The overall resistant rate to amphotericin B has decreased from 2.5% in TSARY 2002 (Yang et al., 2005b) to 1.8% in TSARY 2006. In recent years, fungal infections caused by non-albicans *Candida* species have increased dramatically (Abi-Said et al., 1997; Slavin et al., 1995; Walsh et al., 2004). The phenomenon was also reflected in the distribution of resistant isolates in others' (Slavin et al., 1995; Walsh et al., 1998) as well as our studies. All 3 amphotericin B-resistant isolates in TSARY 1999 were non-albicans *Candida* species (Yang et al., 2004b). Furthermore, 20 of the 23 amphotericin B-resistant isolates in

TSARY 2002 were also non-albicans *Candida* species (Yang et al., 2005b). Of the 17 amphotericin B-resistant isolates in TSARY 2006, 16 were non-albicans *Candida* species. They consisted of 12 *C. tropicalis*, 2 *C. krusei*, and 1 each of *C. glabrata* and *Candida curvatus*.

The prevalence of amphotericin B resistance of *C. krusei* (14.3%) or *C. tropicalis* (4.9%) was significantly higher than that of *C. glabrata* (0.5%) or *C. albicans* (0.2%) ($P < 0.05$). Among the frequently isolated non-albicans *Candida* species, no amphotericin B-resistant isolate of *C. parapsilosis* was detected in this study (Table 2), as well as in the 2 previous TSARYs (Yang et al., 2004b, 2005b). A total of 2, 2, and 5 *C. lusitanae* were collected in TSARY 1999, 2002, and 2006, respectively. Up-to-date amphotericin B resistance of *C. lusitanae* is not an issue for concern since all 9 *C. lusitanae* was susceptible to amphotericin B. The MIC₅₀ and MIC₉₀ of those isolates were 0.25 and 1 µg/mL, respectively.

3.3. Susceptibilities to fluconazole of *Candida* species from different sources

For the susceptibility to fluconazole, a total of 756 (78.4%), 43 (4.5%), and 165 (17.1%) isolates had MICs ≤8, 16 to 32, and ≥64 µg/mL, respectively. The MIC₅₀ and MIC₉₀ of these isolates were 1 and 64 µg/mL, respectively. Interestingly, among the isolates with MICs ≥64 µg/mL, those from sputum (27.2%) appear to have a higher ratio than

Table 2
The susceptibility to amphotericin B of *Candida* species

MICs (µg/mL)	<i>C. albicans</i>	<i>C. tropicalis</i>	<i>C. glabrata</i>	<i>C. parapsilosis</i>	<i>C. krusei</i>	Others	Total
0.125	16 (3.8) ^a	0	4 (1.9)	0	0	1 (8.3)	21 (2.2)
0.25	213 (50.8)	45 (18.3)	49 (23.2)	23 (37.1)	0	6 (50)	336 (34.8)
0.5	134 (31.9)	126 (51.2)	113 (53.6)	29 (46.8)	5 (35.7)	2 (16.7)	409 (42.4)
1	55 (13.1)	63 (25.6)	44 (20.8)	10 (16.1)	7 (50)	2 (16.7)	181 (18.8)
2	1 (0.2)	12 (4.9)	1 (0.5)	0	2 (14.3)	1 (8.3)	17 (1.8)
Total	419	246	211	62	14	12	964
MIC ₅₀	0.25	0.5	0.5	0.5	1	0.25	0.5
MIC ₉₀	1	1	1	1	2	1	1

^a Number of isolates (percentage).

those from ascites (7.9%), wound (11.6%), blood (13.1%), and urine (17.7%) ($P = 0.05$) (Table 3).

When comparisons were made among species, there were distinct variations in the fluconazole susceptibility. Of the 5 common *Candida* species, all *C. krusei* had MICs ≥ 64 $\mu\text{g}/\text{mL}$. In contrast, none of the *C. parapsilosis* did. These results were consistent with previous reports (Akova et al., 1991; Pfaller et al., 2000, 2004; Yang et al., 2004a, 2005b). Furthermore, susceptibilities among different species from blood also varied. All *C. albicans*, *C. glabrata*, and *C. parapsilosis* were susceptible to fluconazole, whereas 18 of the 32 (56.3%) *C. tropicalis* isolates had MICs ≥ 64 $\mu\text{g}/\text{mL}$ (Table 3).

Interestingly, *C. glabrata* having MICs to fluconazole ≥ 64 $\mu\text{g}/\text{mL}$ was lower than what has been reported in other geologic areas (Fleck et al., 2007; Pfaller et al., 2007b; St Germain et al., 2001). In the 3 TSARYs, the portions of isolates having MICs ≥ 64 $\mu\text{g}/\text{mL}$ were 8.3%, 1.6%, and

4.3% isolates from 1999, 2002, and 2006, respectively. It has been reported that continuous exposure to azoles seems to have a major impact on developing resistance to fluconazole in *Candida* species, especially for *C. glabrata* (Kontoyannis, 2002). Therefore, our previous observation that only 13% of the candidemia 92 patients had prior fluconazole treatments (Cheng et al., 2004) may explain the low fluconazole-resistant *C. glabrata* in Taiwan.

3.4. Trailing growth

Among the phenomena associated with resistance, trailing describes the reduced but persistent growth that some isolates exhibit at drug concentrations above the MIC in broth dilution tests with azole antifungal agents, such as fluconazole (Lee et al., 2004). Trailing may interfere the observation of resistance level in vivo (Arthington-Skaggs et al., 2002). Thus, we have also determined whether those 165 isolates with MICs ≥ 64 $\mu\text{g}/\text{mL}$ to fluconazole exhibit

Table 3
The susceptibility to fluconazole of *Candida* species from different sources

	MICs ($\mu\text{g}/\text{mL}$)	<i>C. albicans</i>	<i>C. tropicalis</i>	<i>C. glabrata</i>	<i>C. parapsilosis</i>	<i>C. krusei</i>	Others	Total
Urine	≤ 8	123 (95.3) ^a	53 (48.6)	117 (80.7)	13 (100)	0	5 (100)	311 (76.6)
	16–32	0	1 (0.9)	22 (15.2)	0	0	0	23 (5.7)
	≥ 64	6 (4.7)	55 (50.5)	6 (4.1)	0	5 (100)	0	72 (17.7)
	Subtotal	129	109	145	13	5	5	406
Sputum	≤ 8	69 (98.6)	25 (38.5)	13 (68.4)	2 (100)	0	0	109 (69)
	16–32	1 (1.4)	0	5 (26.3)	0	0	0	6 (3.8)
	≥ 64	0	40 (61.5)	1 (5.3)	0	2 (100)	0	43 (27.2)
	Subtotal	70	65	19	2	2	0	158
Blood	≤ 8	82 (100)	10 (31.2)	16 (80)	9 (100)	0	1 (100)	118 (81.4)
	16–32	0	4 (12.5)	4 (20)	0	0	0	8 (5.5)
	≥ 64	0	18 (56.3)	0	0	1 (100)	0	19 (13.1)
	Subtotal	82	32	20	9	1	1	145
Wound	≤ 8	21 (100)	2 (3.3)	5 (83.3)	5 (100)	0	3 (75)	36 (83.7)
	16–32	0	0	1 (16.7)	0	0	1 (25)	2 (4.7)
	≥ 64	0	4 (66.7)	0	0	1 (100)	0	5 (11.6)
	Subtotal	21	6	6	5	1	4	43
Pus	≤ 8	19 (95)	4 (44.4)	2 (100)	6 (100)	0	1 (100)	32 (80)
	16–32	0	0	0	0	0	0	0
	≥ 64	1 (5)	5 (55.6)	0	0	2 (100)	0	8 (20)
	Subtotal	20	9	2	6	2	1	40
Tip ^b	≤ 8	18 (90)	7 (53.8)	2 (66.7)	4 (100)	0	0	31 (77.5)
	16–32	0	1 (7.7)	0	0	0	0	1 (2.5)
	≥ 64	2 (10)	5 (38.5)	1 (33.3)	0	0	0	8 (20)
	Subtotal	20	13	3	4	0	0	40
Ascites ^c	≤ 8	24 (100)	5 (71.4)	4 (100)	2 (100)	0	0	35 (92.1)
	16–32	0	0	0	0	0	0	0
	≥ 64	0	2 (28.6)	0	0	1 (100)	0	3 (7.9)
	Subtotal	24	7	4	2	1	0	38
Others	≤ 8	51 (96.2)	2 (40)	10 (83.4)	20 (95.2)	0.0	1 (100)	84 (89.4)
	16–32	1 (1.9)	0.0	1 (8.3)	1 (4.8)	0.0	0.0	3 (3.2)
	≥ 64	1 (1.9)	3 (30)	1 (8.3)	0	2 (100)	0	7 (7.4)
	Subtotal	53	5	12	21	2	1	94
All	≤ 8	407 (97.1)	108 (43.9)	169 (80.1)	61 (98.4)	0	11 (91.7)	756 (78.4)
	16–32	2 (0.5)	6 (2.4)	33 (15.6)	1 (1.6)	0	1 (8.3)	43 (4.5)
	≥ 64	10 (2.4)	132 (53.7)	9 (4.3)	0	14 (100)	0	165 (17.1)
	Total	419	246	211	62	14	12	964

^a Number of isolates (percentage).

^b Catheter tips.

^c Body fluid, not urinary ascites or pus.

Table 4

The distribution of trailing growth of *Candida* species with MICs greater than 64 µg/mL

Sources	<i>C. albicans</i>		<i>C. glabrata</i>		<i>C. krusei</i>		<i>C. tropicalis</i>		Subtotal		Total
	No	Yes	No	Yes	No	Yes	No	Yes	No	Yes	
	Urine	3	3	5	1	5	0	13	42	26	
Sputum	0	0	0	1	2	0	14	26	16	27	43
Blood	0	0	0	0	1	0	6	12	7	12	19
Pus	0	1	0	0	2	0	0	5	2	6	8
Ascites	0	0	0	0	1	0	0	2	1	2	3
Wound	0	0	0	0	1	0	0	4	1	4	5
Tip	0	1	0	0	0	0	0	4	0	5	5
Others	0	2	0	2	1	0	3	1	4	5	9
Total	3	7	5	4	14	0	36	96	58	107	165

trailing, and the results are summarized in Table 4. High percentage (64.8%) of those isolates indeed exhibited the trailing phenomenon. When classified according to species, 72.7% (96/132) *C. tropicalis*, 70% (7/10) *C. albicans*, and 44.4% (4/9) *C. glabrata* isolates exhibited the trailing phenomenon, whereas none of the *C. krusei* isolates did.

3.5. Conclusion

In TSARY 1999, there were 14.7% *C. tropicalis* isolates having MICs ≥ 64 µg/mL (Yang et al., 2004b). In contrast, none of the 244 *C. tropicalis* from TSARY 2002 had MICs ≥ 64 µg/mL (Yang et al., 2005b). Surprisingly, in TSARY 2006, among the 246 *C. tropicalis* isolates, 132 (53.7%) had MICs ≥ 64 µg/mL. Recently, we have reported an association between fluconazole susceptibility and genetic relatedness among *C. tropicalis* isolates from TSARY 1999 (Chou et al., 2007; Wang et al., 2007). The DST140 was a predominant type of *C. tropicalis* isolates among those having MICs ≥ 64 µg/mL in TSARY 1999 (Chou et al., 2007). Furthermore, none of the 17 tested fluconazole-susceptible *C. tropicalis* isolates collected from TSARY 2002 were DST140 type (Chou et al., 2007). Interestingly, our preliminary result showed that DST140 was detected again among *C. tropicalis* isolates collected in TSARY 2006. This result may explain the dramatic fluctuation in fluconazole susceptibility among 3 TSARYs. Whether this is an endemic problem requires further investigation.

The increasing rate of reduced susceptibility to fluconazole in *C. tropicalis* has considerable clinical importance, because this species is 1 of the most frequently isolated non-*albicans* *Candida* species (Cheng et al., 2004; Hung et al., 2005; Lo et al., 2005; Pfaller et al., 2000; Prasad et al., 1999). Furthermore, *C. tropicalis* develops drug resistance in the presence of fluconazole much more rapidly than *C. albicans* (Barchiesi et al., 2000; Calvet et al., 1997). In addition, *C. krusei* isolates with high MICs are also closely related (Chou et al., 2007; Wang et al., 2007). These findings may explain why, to fluconazole, *C. tropicalis* has a higher rate to have MICs ≥ 64 µg/mL than *C. albicans*. Here we have identified that 72.7% (96/132) *C. tropicalis* exhibited the

trailing phenomenon by a definition that the MIC of an isolate after 48-h incubation is 4-fold higher than that at the 24-h point. Nevertheless, whether the remaining 36 (14.6%) *C. tropicalis* isolates are “truly resistant” requires further investigation.

Acknowledgments

The authors would like to thank Bristol Myers Squibb (New Brunswick, NJ) and Pfizer (New York, NY) for supplying the amphotericin B and fluconazole, respectively. They would also like to acknowledge Dr. Y.C. Chen for her assistance and helpful suggestion. They also thank the 22 participating hospitals for providing clinical isolates and information regarding those isolates. They are Asia East Memorial Hospital, Buddhist Tzu-Chi General Hospital, Chang Gung Memorial Chiayi Christian Hospital, Hospital at Kaohsiung, Chang-Hwa Christian Hospital, Cheng Ching Hospital, Chung Shan Medical Dental College Hospital, Kaohsiung Military Hospital, Kaohsiung Medical College Chung-Ho Memorial Hospital, Kuan-Tien General Hospital, Lo-Hsu Foundation Lo-Tung Poh Ai Hospital, Miin Sheng General Hospital, National Cheng Kung University Hospital, Show Chwan Memorial Hospital, Sin-Lau Christian Hospital, St. Mary Hospital, Taipei Municipal Chung Hsiao Hospital, Taipei Municipal Hoping Hospital, Veterans General Hospital-Taichung, Veterans General Hospital-Kaohsiung, Zen Ai General Hospital, and Tungs' Taichung MetroHarbor Hospital, Sha Lu branch. We thank Dr. T.L. Lauderdale and Mr. S.H. Hsu for their technical assistance. This study was supported by the grant NHRI CL-096-PP07.

References

- Abi-Said D, Anaissie E, Uzun O, Raad I, Pinzcowski H, Vartivarian S (1997) The epidemiology of hematogenous candidiasis caused by different *Candida* species. *Clin Infect Dis* 24:1122–1128.
- Akova M, Akalin HE, Uzun O, Gur D (1991) Emergence of *Candida krusei* infections after therapy of oropharyngeal candidiasis with fluconazole. *Eur J Clin Microbiol Infect Dis* 10:598–599.
- Arthington-Skaggs BA, Lee-Yang W, Ciblak MA, Frade JP, Brandt ME, Hajjeh RA, Harrison LH, Sofair AN, Warnock DW (2002) Comparison of visual and spectrophotometric methods of broth microdilution MIC end point determination and evaluation of a sterol quantitation method for in vitro susceptibility testing of fluconazole and itraconazole against trailing and nontrailing *Candida* isolates. *Antimicrob Agents Chemother* 46:2477–2481.
- Barchiesi F, Calabrese D, Sanglard D, Falconi DF, Caselli F, Giannini D, Giacometti A, Gavaudan S, Scalise G (2000) Experimental induction of fluconazole resistance in *Candida tropicalis* ATCC 750. *Antimicrob Agents Chemother* 44:1578–1584.
- Calvet HM, Yeaman MR, Filler SG (1997) Reversible fluconazole resistance in *Candida albicans*: a potential in vitro model. *Antimicrob Agents Chemother* 41:535–539.
- Chen YC, Chang SC, Sun CC, Yang LS, Hsieh WC, Luh KT (1997) Secular trends in the epidemiology of nosocomial fungal infections at a teaching hospital in Taiwan, 1981 to 1993. *Infect Control Hosp Epidemiol* 18:369–375.

- Cheng MF, Yu KW, Tang RB, Fan YH, Yang YL, Hsieh KS, Ho M, Lo HJ (2004) Distribution and antifungal susceptibility of *Candida* species causing candidemia from 1996 to 1999. *Diagn Microbiol Infect Dis* 48:33–37.
- Chou HH, Lo HJ, Chen KW, Liao MH, Li SY (2007) Multilocus sequence typing of *Candida tropicalis* shows clonal cluster enriched in isolates with resistance or trailing growth of fluconazole. *Diagn Microbiol Infect Dis* 58:427–433.
- Clinical and Laboratory Standards Institute (formerly NCCLS) (1997) Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Approved Standard. M27A. Wayne, PA: CLSI.
- Fleck R, Dietz A, Hof H (2007) In vitro susceptibility of *Candida* species to five antifungal agents in a German university hospital assessed by the reference broth microdilution method and Etest. *J Antimicrob Chemother* 59:767–771.
- Hadfield TL, Smith MB, Winn RE, Rinaldi MG, Guerra C (1987) Mycoses caused by *Candida lusitanae*. *Rev Infect Dis* 9:1006–1012.
- Hung CC, Chen YC, Chang SC, Luh KT, Hsieh WC (1996) Nosocomial candidemia in a university hospital in Taiwan. *J Formos Med Assoc* 95:19–28.
- Hung CC, Yang YL, Lauderdale TL, McDonald LC, Hsiao CF, Cheng HH, Ho YA, Lo HJ (2005) Colonization of human immunodeficiency virus-infected outpatients in Taiwan with *Candida* species. *J Clin Microbiol* 43:1600–1603.
- Kontoyiannis DP (2002) Why prior fluconazole use is associated with an increased risk of invasive mold infections in immunosuppressed hosts: an alternative hypothesis. *Clin Infect Dis* 34:1281–1283.
- Leaw SN, Chang HC, Sun HF, Barton R, Bouchara JP, Chang TC (2006) Identification of medically important yeast species by sequence analysis of the internal transcribed spacer regions. *J Clin Microbiol* 44:693–699.
- Leaw SN, Chang HC, Barton R, Bouchara JP, Chang TC (2007) Identification of medically important *Candida* and non-*Candida* yeast species by an oligonucleotide array. *J Clin Microbiol* 45:2220–2229.
- Lee MK, Williams LE, Warnock DW, Arthington-Skaggs BA (2004) Drug resistance genes and trailing growth in *Candida albicans* isolates. *J Antimicrob Chemother* 53:217–224.
- Lo HJ, Ho AH, Ho M (2001) Factors accounting for mis-identification of *Candida* species. *J Microbiol Immunol Infect* 34:171–177.
- Lo HJ, Cheng HH, TSARY Hospitals (2005) Distribution of clinical yeasts in Taiwan. *Fung Sci* 20:83–91.
- Marr KA, Lyons CN, Ha K, Rustad TR, White TC (2001) Inducible azole resistance associated with a heterogeneous phenotype in *Candida albicans*. *Antimicrob Agents Chemother* 45:52–59.
- Orozco AS, Higginbotham LM, Hitchcock CA, Parkinson T, Falconer D, Ibrahim AS, Ghannoum MA, Filler SG (1998) Mechanism of fluconazole resistance in *Candida krusei*. *Antimicrob Agents Chemother* 42:2645–2649.
- Peman J, Canton E, Gobernado M (2005) Epidemiology and antifungal susceptibility of *Candida* species isolated from blood: results of a 2-year multicentre study in Spain. *Eur J Clin Microbiol Infect Dis* 24:23–30.
- Pfaller MA, Messer SA, Hollis RJ, Jones RN, Doern GV, Brandt ME, Hajjeh RA (1999) Trends in species distribution and susceptibility to fluconazole among blood stream isolates of *Candida* species in the United States. *Diagn Microbiol Infect Dis* 33:217–222.
- Pfaller MA, Jones RN, Doern GV, Sader HS, Messer SA, Houston A, Coffman S, Hollis RJ (2000) Bloodstream infections due to *Candida* species: SENTRY antimicrobial surveillance program in North America and Latin America, 1997–1998. *Antimicrob Agents Chemother* 44:747–751.
- Pfaller MA, Diekema DJ, Messer SA, Boyken L, Hollis RJ (2003) Activities of fluconazole and voriconazole against 1,586 recent clinical isolates of *Candida* species determined by Broth microdilution, disk diffusion, and Etest methods: report from the ARTEMIS Global Antifungal Susceptibility Program, 2001. *J Clin Microbiol* 41:1440–1446.
- Pfaller MA, Messer SA, Boyken L, Hollis RJ, Rice C, Tendolkar S, Diekema DJ (2004) In vitro activities of voriconazole, posaconazole, and fluconazole against 4,169 clinical isolates of *Candida* spp. and *Cryptococcus neoformans* collected during 2001 and 2002 in the ARTEMIS global antifungal surveillance program. *Diagn Microbiol Infect Dis* 48:201–205.
- Pfaller MA, Boyken L, Hollis RJ, Kroeger J, Messer SA, Tendolkar S, Diekema DJ (2007a) In vitro susceptibility of invasive isolates of *Candida* spp. to anidulafungin, caspofungin, and micafungin: six years of global surveillance. *J Clin Microbiol* 46:150–156.
- Pfaller MA, Diekema DJ, Gibbs DL, Newell VA, Meis JF, Gould IM, Fu W, Colombo AL, Rodriguez-Noriega E (2007b) Results from the ARTEMIS DISK Global Antifungal Surveillance study, 1997 to 2005: an 8.5-year analysis of susceptibilities of *Candida* species and other yeast species to fluconazole and voriconazole determined by CLSI standardized disk diffusion testing. *J Clin Microbiol* 45:1735–1745.
- Prasad KN, Agarwal J, Dixit AK, Tiwari DP, Dhole TN, Ayyagari A (1999) Role of yeasts as nosocomial pathogens and their susceptibility to fluconazole and amphotericin B. *Indian J Med Res* 110:11–17.
- Slavin MA, Osborne B, Adams R, Levenstein MJ, Schoch HG, Feldman AR, Meyers JD, Bowden RA (1995) Efficacy and safety of fluconazole prophylaxis for fungal infections after marrow transplantation—a prospective, randomized, double-blind study. *J Infect Dis* 171:1545–1552.
- St Germain G, Laverdiere M, Pelletier R, Bourgault AM, Libman M, Lemieux C, Noel G (2001) Prevalence and antifungal susceptibility of 442 *Candida* isolates from blood and other normally sterile sites: results of a 2-year (1996 to 1998) multicenter surveillance study in Quebec, Canada. *J Clin Microbiol* 39:949–953.
- Walsh TJ, Hiemenz JW, Seibel NL, Perfect JR, Horwith G, Lee L, Silber JL, DiNubile MJ, Reboli A, Bow E, Lister J, Anaissie EJ (1998) Amphotericin B lipid complex for invasive fungal infections: analysis of safety and efficacy in 556 cases. *Clin Infect Dis* 26:1383–1396.
- Walsh TJ, Groll A, Hiemenz J, Fleming R, Roilides E, Anaissie E (2004) Infections due to emerging and uncommon medically important fungal pathogens. *Clin Microbiol Infect* 10(Suppl 1):48–66.
- Wang JS, Li SY, Yang YL, Chou HH, Lo HJ (2007) Association between fluconazole susceptibility and genetic relatedness among *Candida tropicalis* isolates in Taiwan. *J Med Microbiol* 56:650–653.
- White TC, Marr KA, Bowden RA (1998) Clinical, cellular, and molecular factors that contribute to antifungal drug resistance. *Clin Microbiol Rev* 11:382–402.
- Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, Edmond MB (2004) Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. *Clin Infect Dis* 39:309–317.
- Yang YL, Lo HJ (2001) Mechanisms of antifungal agent resistance. *J Microbiol Immunol Infect* 34:79–86.
- Yang YL, Cheng HH, Lo HJ (2004a) In vitro activity of voriconazole against *Candida* species isolated in Taiwan. *Int J Antimicrob Agents* 24:294–296.
- Yang YL, Ho YA, Cheng HH, Ho M, Lo HJ (2004b) Susceptibilities of *Candida* species to amphotericin B and fluconazole: the emergence of fluconazole resistance in *Candida tropicalis*. *Infect Control Hosp Epidemiol* 25:60–64.
- Yang YL, Ho YA, Cheng HH, Lo HJ (2005a) Distribution and susceptibility to amphotericin B and fluconazole of *Candida* spp. isolated from Taiwan. *Epidemiol Infect* 133:325–330.
- Yang YL, Li SY, Cheng HH, Lo HJ (2005b) Susceptibilities to amphotericin B and fluconazole of *Candida* species in TSARY 2002. *Diagn Microbiol Infect Dis* 51:179–183.
- Yang YL, Cheng HH, Lo HJ (2006) Distribution and antifungal susceptibility of *Candida* species isolated from different age populations in Taiwan. *Med Mycol* 44:237–242.